
Deciphering organic matter degradation in the continuous permafrost zone of Alaska based on biomarker analyses

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Submitted by: Carolin Luise Frauhammer²
E-Mail: carolin.frauhammer@uni-potsdam.de
Matriculation number: 797703

1st Supervisor: Dr. Jens Strauss¹
2nd Supervisor: Prof. Dr. Guido Grosse^{1,2}

¹ Alfred-Wegener-Institute Potsdam

² University of Potsdam



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Abbreviations

~	approximately	
°	degree	
°C	degree Celsius	273.15 K
°N	latitudinal position in degrees north the equator	
°W	longitudinal position in degrees west the prime median	
Ø	Diameter	
‰	per mille	
δ ¹³ C	stable carbon isotope ratio ¹³ C and ¹² C	‰ vs. VPDB
δ ¹⁸ O	stable oxygen isotope ratio ¹⁸ O and ¹⁶ O	‰ vs. SMOW
δD	stable hydrogen isotope ratio ² H and ¹ H	‰ vs. SMOW
¹⁴ C	radiocarbon age; radioactive carbon isotope ¹⁴ C and ¹² C	cal. ka BP
ACL	average chain length	
ACP	Arctic coastal plain	
AL	active layer (as permafrost layer)	
BP	before present (1950)	
b.s.l.	below surface level	cm
C	carbon	
cal.	calibrated	
CH ₄	methane	
cm	centimetre	10 ⁻² m
C/N	ratio between TOC and TN	
CO ₂	carbondioxide	
CPI	carbon preference index	
DLB	drained thermokarst lake basin core BAL-T0-DLB	
DTLB	drained thermokarst lake basin	
DOC	dissolved organic matter	mg L ⁻¹
e.g.	latin: exempli grata; for example	
EC	electrical conductivity	µS cm ⁻¹
eV	electronvolt	1.602*10 ⁻¹⁹ J
g	gram	10 ⁻³ kg
GHG	greenhouse gas	
HCPC	hierarchical clustering on principal components	
Gt	gigaton	10 ¹² kg
h	hour	3600 s
ha	hectare	10 000 m ²
ka	latin: kilo annus; thousand years	3.154 * 10 ¹⁰ s
L	thermokarst lake core BAL24-T0-L	
M	marine core BAL24-T0-M	
m ³	cubic meter	m ³
m/z	mass-to-charge ratio	
min	minute	60 s

Abbreviations

mL	millilitre	10^{-6} m^3
mg	milligram	10^{-3} g
mm	millimetre	10^{-3} m
MPa	megapascal	10^6 Pa
n.valid	number of valid cases	
nm	nanometre	10^{-9} m
ns	not statistically significant	
OM	organic matter	
p	p-value; probability for a statistical test	
p _{aq}	aquatic proxy	
PCA	principal component analysis	
PF	perennial frozen (as permafrost layer)	
pH	potential of hydrogen	
p _{wax}	terrestrial proxy	
R	Pearson's correlation coefficient	
R ²	coefficient of determination	
Sed	sediment	
SOC	soil organic carbon	
SMOW	standard mean ocean water	‰
T	talik (as permafrost layer)	
TC	total carbon	wt%
TIC	total inorganic carbon	wt%
TL	thermocarst lake	
TN	total nitrogen	wt%
TOC	total organic carbon	wt%
u	unified atomic mass unit	$1.661 \cdot 10^{-27} \text{ kg}$
UL	upland core BAL24-T0-UL	
vol%	per cent by volume	
VPDB	Vienna Pee Dee Belemnite	‰
wt%	per cent by weight	
µg	microgram	10^{-9} kg
µL	microliter	10^{-6} L
µS	microsiemens	10^{-6} S

Abstract

Human-induced climate change has led to a significant rise in global temperatures, with the Arctic warming nearly four times faster than the global average. This accelerated warming makes the Arctic one of the most vulnerable regions on Earth. A major component of this region is permafrost, permanently frozen grounds that covers around 22 % of the exposed land area of the northern Hemisphere. Permafrost regions store vast amounts of carbon and are therefore a key point in the carbon-climate-cycle, as the strong warming threatens the stability of the frozen ground and makes these storages available for microbial decomposition, which increases the greenhouse gas production. In particular thermo-karst processes – ground subsidence caused by the thawing of ice-rich permafrost – re-shape permafrost landscapes, resulting in degeneration features as thermokarst lakes, drained thermokarst lake basins and coastal erosion. This mobilizes previously frozen organic matter, altering its composition, reactivity and greenhouse gas potential. Thus, knowledge about the origin, quantity and quality are essential to estimate the future fate of organic matter in a changing permafrost landscape. This study addresses these knowledge gaps by analysing OM characteristics in northwestern Alaska. The aim of this study is to investigate (1) the paleoenvironment of the landscape and (2) the characterization of the OM by its quantity, source and quality. Therefore a multiproxy approach of bio-geochemistry, hydrochemistry, sedimentology and biomarker analyses for a transect along the four main features Upland, Thermokarst lake, Drained Thermokarst Lake Basin and Marine on the Baldwin Peninsula was used.

The deposition in this region started > 50 cal ka BP in a strongly aeolian regime. The sediments are mostly of coarse silt, indicating parent material at all four sites, and show with high water contents, organic-rich layers and high radiocarbon ages typically characteristics of late Pleistocene Yedoma, while the bottom layers show signs of an ancient fluvial environment and former thermokarst processes. Also, the biomarker analysis suggests a common terrestrial origin of the OM, also for the marine site, indicating a secondary marine infiltration, as well as a slightly aquatic influences, especially in the deeper layers, resulting from ancient thermokarst processes and the lake / marine phases. The carbon quantity decreases significantly from the Upland to the Marine site. It has a high quality without statistically significant differences between the sites.

Due to the high ice amount and carbon quality, the studied deposits are highly vulnerable to thawing under future warming and thus constitute a high greenhouse gas release potential.

Zusammenfassung

Der anthropogene Klimawandel hat zu einem signifikanten Anstieg der globalen Temperatur geführt, wobei sich die Arktis fast viermal so schnell wie der globale Durchschnitt erwärmt. Diese beschleunigte Erwärmung macht sie zu einer der vulnerabelsten Regionen der Welt. Einer der Hauptbestandteile dieser Region ist Permafrost, dauerhaft gefrorener Boden, der ~22 % der freiliegenden Landfläche der nördlichen Hemisphäre bedeckt. Permafrost-Regionen speichern große Mengen an Kohlenstoff und sind dadurch ein Kernpunkt im Kohlenstoff-Klima-Kreislauf, da die starke Erwärmung die Stabilität des gefrorenen Bodens und damit des Kohlenstoffes gefährdet. Besonders Thermokarstprozesse – Bodenabsenkung infolge des Taus von eisreichem Permafrost – verändern diese Landschaften und schaffen Strukturen wie Thermokarstseen, drainierte Thermokarstseebecken und Küstenerosion. Das wiederum mobilisiert zuvor eingefrorenes organisches Material, verändert dessen Zusammensetzung und Treibhausgaspotential. Daher ist es essenziell, mehr Wissen über die Charakteristik des organischen Materials zu erlangen, um dessen zukünftige Entwicklung in einer sich wandelnden Permafrost-Landschaft abzuschätzen. Diese Studie fokussiert sich auf diese Wissenslücke. Ziel ist die Untersuchung (1) der Paläoumwelt und Entstehung der Landschaft sowie (2) die Charakterisierung der OM anhand ihrer Quantität, Herkunft und Qualität. Dafür wird ein Multi-Proxy-Ansatz aus Bio-geochemie, Hydrochemie, Sedimentologie und Biomarker-Analyse entlang eines Transekts mit vier Hauptmerkmalen – Hochebene, Thermokarstsee, drainiertes Thermokarstseebecken und marine Ablagerungen – auf der Baldwin-Halbinsel angewendet. Die Ablagerung startet vor über 50 cal ka BP in einem stark äolischen Regime. Die Sedimente bestehen überwiegend aus grobem Schluff, was auf dasselbe Ausgangsmaterial an allen Standorten hinweist. Sehr hohe Eisgehalte, organikreiche Schichten und Radiokarbonalter > 30 cal ka BP deuten auf typische Eigenschaften spätpleistozänen Yedomas hin, während tiefere Schichten Spuren früherer fluvialer Bedingungen und Thermokarstprozesse aufweisen. Die Biomarker-Analyse bestätigt einen gemeinsamen terrestrischen Ursprung der OM, auch am marinen Standort, der terrestrisch abgelagert und später durch Erosion und Infiltration marin überprägt wurde. Die Kohlenstoffmenge nimmt vom Hochland zum marinen Standort deutlich ab. Sie weist eine hohe Qualität auf, ohne signifikante Unterschiede zwischen den Standorten. Aufgrund hoher Eismengen, möglicher Talikbildung und undegradierter Kohlenstoffe sind die untersuchten Ablagerungen sehr anfällig für Auftauen unter zukünftiger Erwärmung und stellen somit ein hohes Treibhausgas-Freisetzungspotenzial dar.

1. Introduction

1.1 Thematic relevance

Humanity and ecosystems are currently facing on the most pressing challenges due anthropogenic global warming. The climate has warmed at a rate unprecedented in at least the last 2 000 years by human's influence, with higher temperatures each of the last four decades than any previous one since 1850 (IPCC 2023c). According to the IPCC (2023a), the global surface temperature in 2011 – 2020 was around 1.1 °C above 1850 – 1900 due to human caused stronger radiative forcing, mainly by increasing concentrations of the greenhouse gases (GHG) carbon dioxide (CO₂), methane (CH₄) and nitrous oxide (N₂O). In 1900 – 2019, atmospheric CO₂ has risen as a minimum ten times faster than any previous in the last 800 000 years and likely four – five times faster than in the past 56 million years (IPCC 2023b). Regardless of emission scenarios, global temperatures are projected to rise by more than 2 °C by mid-century unless negative GHG emissions occur in the coming decades. Climate change has strong effects on all compartments of the earth systems, as sea-ice and ice-sheet loss, glacier retreat, global mean sea level rise due land ice loss and thermal expansion and the weather. So, additional warming will increase the frequency and intensity of climate extremes as heatwaves, heavy precipitations, drought and tropical cyclones (IPCC 2023c). Especially the polar regions are highly climate-sensitive areas on the planet, with the Arctic having warmed up to four times as fast as the global average in the last 30 years, as seen in Figure 1, and is projected to continue to warm with above two times the rate of global warming and an increase in the temperature of the coldest days three times the rate of global mean (Rantanen et al. 2022).

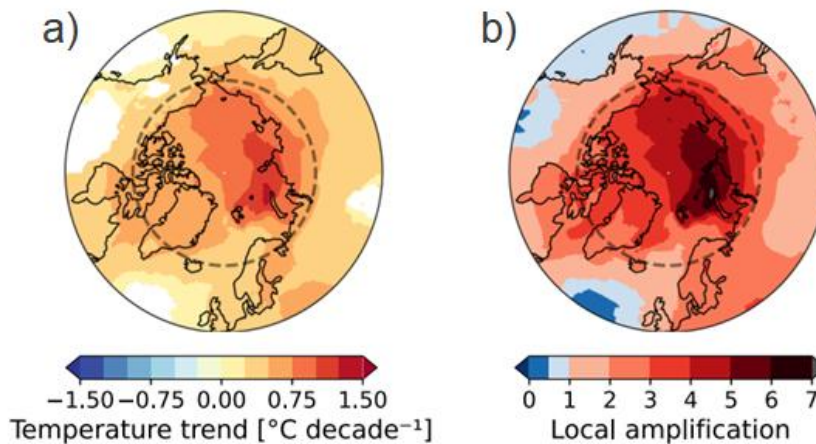


Figure 1. (a) Annual mean temperature trends and (b) local amplification ratio from 1979 – 2021. The Arctic circle (66.5 °N latitude) is indicated by the dashed line. From Rantanen et al. 2022.

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This phenomenon is called Arctic amplification, likely caused among others by increasing ocean heating and ice-albedo feedback (Screen and Simmonds 2010; Dai et al. 2019), Plank feedback (Pithan and Mauritsen 2014), lapse-rate feedback (Stuecker et al. 2018), inversion of the Arctic wintertime temperature (Bintanja et al. 2011), cloud feedback (Taylor et al. 2013), enhanced horizontal ocean heat transport (Beer et al. 2020) and transport of moist air into the arctic (Woods and Caballero 2016).

Permafrost soils, underlying 15 % of the terrestrial environment of the Northern Hemisphere, are particularly vulnerable to global warming, with increasing ground temperatures to record high levels up to 2 – 3 °C warmer than 30 years ago (IPCC polar regions 2019).

This warming causes a deepening of the active layer, the layer above the permafrost which is subject to annual thawing and freezing, up to complete thawing in the future (Schaeffer et al. 2011). The consequences of permafrost degradation are one the one hand regional impacts as change of ecosystems and hydrology, infrastructure damages and food security, especially for indigenous people living in these regions. Moreover, approximately 1 538 Gt carbon (C) are stored in the terrestrial permafrost region, which accounts for ~30 % of the global C pool in only ~15 % of the global land area. In addition, there are estimated 2 822 Gt C stored in subsea permafrost, resulting in 4 360 Gt C in total, as seen in Figure 2 (Strauss et al. 2025).

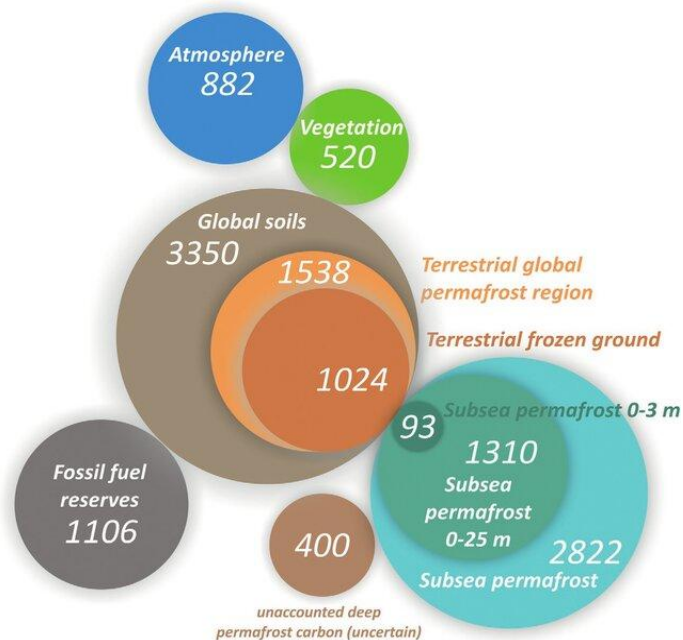


Figure 2. Atmospheric and terrestrial carbon stocks in relation to carbon stored in the permafrost zone, given in Gt, with the size of the circles proportional to the size of the carbon stocks. The global soil estimate (3350 Gt) is based on soils ≤ 3 m depth (2800 Gt) as well as deep permafrost > 3 m depth (500 Gt) and tropical peatlands (50 Gt; (Jackson et al. 2017)). Based on data from different IPCC reports, (Miesner et al. 2023) and (Hugelius et al. 2014; Hugelius et al. 2020). Adapted and updated from (Strauss et al. 2017). Figure from Strauss et al. (2025).

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The further Arctic warming is expected to make these deposits of organic-rich material trapped within permafrost more accessible for microbial decomposition, leading to heightened production of GHG as permafrost thaws, and thus in further climate warming; the positive permafrost carbon feedback, Figure 3 (Schuur et al. 2008; Schaeffer et al. 2011). One of the most abundant forms of permafrost degradation in ice-rich Arctic landscapes is thermokarst (Section 2.3), resulting from the thawing of excess ice-bearing permafrost or the melting of massive near-surface ice due active layer deepening (van Everdingen 2005; Morgenstern et al. 2011; Farquharson et al. 2022). The following surface lowering can lead to the formation of thaw lakes, called thermokarst lakes (TL) (van Everdingen 2005; Grosse et al. 2011), which grow, coalesce and eventually drain to drained thermokarst lake basins (DTLB), resulting in a lake-drained lake basin system (Fuchs et al. 2019). As thermokarst is already the dominant landscape change process in the Arctic with occupying ~20 % of the northern permafrost region, rising permafrost temperatures induce an increase in frequency and magnitude of thermokarst as well as a shift to landscapes dominated by DTLBs (Olefeldt et al. 2016; Jones et al. 2022; Jones et al. 2023). TLs have been shown to release organic carbon (OC) previously frozen in permafrost, especially since they can reach and thus mobilize old, deep permafrost carbon. DTLBs have the potential to reaggregate permafrost in a cold climate, though this potential will slow or not occur in a warming climate (Jones et al. 2022).

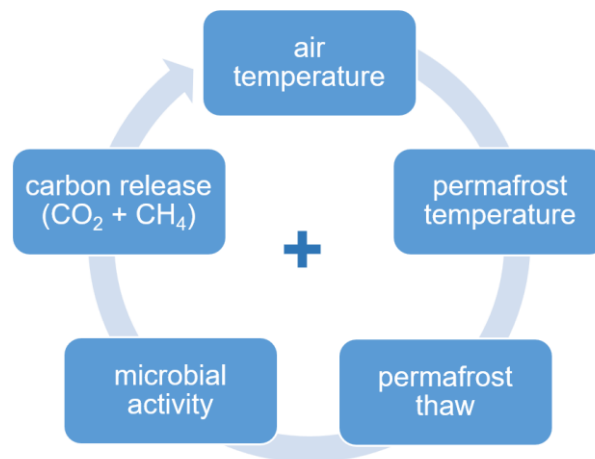


Figure 3. The permafrost-carbon feedback loop.

In addition, there is the process of talik formation (perennially thawed subsurface soils) by an adjustment of the active layer to a new thermal regime (Jorgenson et al. 2010), thermokarst related under a lake (subaquatic) due thermal interaction between surface water and permafrost (Parsekian et al. 2013), or non-lake environments (subaerial), when the summer thaw-depth exceeds the winter freeze-depth (Farquharson et al. 2022). The formation of talik

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is of special interest for the permafrost-carbon feedback, as it accelerates permafrost degradation and can mobilize old soil organic carbon (SOC) in deep permafrost (> 3 m), leading to a potential shift in the Arctic ecosystems from a C sink to a C source in long-term (Parazoo et al. 2018).

Many TLs and DTLBs are located close to the coast and are therefore strongly influenced by it and its changes (Hussey and Michelson 1966). The Arctic coastlines, which account for approximately 34 % of global coastlines (Lantuit et al. 2012), are with on average 0.5 m a^{-1} and with locally up to 47 m a^{-1} under the fastest eroding coasts (Lantuit et al. 2012; Obu et al. 2017; Malenfant et al. 2022), leading to a high release of c into the Arctic sea (Creel et al. 2024) and a change of the biochemistry of the permafrost soils by salt inundation (Jenrich et al. 2021).

Due to the resulting transition from a terrestrial to a marine permafrost environment through coastal erosion, marine sites represent a potentially end-member version along a permafrost degradation transect in the Arctic coastal plain (ACP). Thereby they serve as an expected future fate of degraded carbon from thermokarst affected soils. Especially knowledge about biochemical properties of the soils as well as processes mobilizing and decomposing deep, old carbon in the context of thermokarst, talik development and marine transgression are rare. As the mobilization and decomposition of organic matter (OM) also highly depends on soil properties as grain size (distribution), pH and C/N, investing these parameters is key to assess the quality and especially the potential GHG release of C from a permafrost affected landscape (Strauss et al. 2015). Additional biomarker analysis can support these by contributing information about the C sources as well as the quality (Fuchs et al. 2019).

1.2 Aim of the study

As there is sparse knowledge about C stocks, quality and mobility in thermokarst affected permafrost regions (Fuchs et al. 2019), this study uses a multiproxy approach of biogeochemistry, hydrochemistry, sedimentology and biomarker analyses to investigate the source and quality changes of OM affected by permafrost degradation along a landscape transect – from an undegraded upland, a thermokarst lake and drained thermokarst lake basin up to a marine site – to assess the potential vulnerability of the stored OM (Fuchs et al. 2019). The aim is to gain a further knowledge of the processes influencing the stability and mobility of OM in a changing permafrost landscape and of its potential future under further climate warming. Therefore, the four primary landscape units Upland (UL), Thermokarst Lake (L), Drained Lake Basin (DLB) and Marine (M), as well as the three permafrost layers contained

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therein, active layer (AL), perennial frozen (PF) and talik (T), were used to create a space-for-time approach along a geomorphological gradient of multiple successive permafrost degradation stages in the ACP. The resulting research objectives of this study are:

- 1) Reconstruction of the paleoenvironment of the landscape and the four landscape units
- 2) Characterisation of the OM with respect to differences between the four landscape units and the three permafrost layers, regarding:
 - a. Quantity of the stored OM
 - b. Source of the stored OM
 - c. Quality of the stored OM and its development along the transect and the degradation of the permafrost

2. Theoretical background

2.1 Periglacial environment and permafrost

Periglacial refers to conditions, processes and landforms associated with cold, non-glacial environments. These environments are dominated by frost action processes and often underlain by permafrost (van Everdingen 2005). Permafrost itself is defined as any ground with a maximum of 0 °C for at least two consecutive years, this can be soil or rock, frozen or unfrozen (van Everdingen 2005). The thickness of permafrost varies between 350 and 650 m in the continuous permafrost zone of the Northern Hemisphere and between 1 and 50 m in the discontinuous zone (Strauss et al. 2021a). In the Northern Hemisphere, the permafrost region covers around 22 % of the terrestrial environment, as seen in Figure 4. This includes more than 60 % of the Russian Federation, up to 46 % of the Canadian land surface, large areas of Alaska, Tibet and China, Greenland, northern Scandinavia, and Antarctic islands and dry valleys (Obu et al. 2019). Depending on the geographic continuity in the landscape, permafrost is categorized into four zones: the continuous zone with permafrost underlying 90 – 100 %, the discontinuous zone with permafrost underlying 50 – 90 %, the sporadic zone with permafrost underlying 10 – 50 % and isolated patches with permafrost underlying 0 – 10 % of the area.

Large quantities of permafrost were formed mostly during the late Pleistocene (126 – 11.7 ka BP) in the lowlands and hills of the northern circumpolar region as a remnant of the paleoclimate. The formation was syngenetic, deposition and permafrost development at the same time, as the Yedoma permafrost (Section 2.2), and epigenetic, permafrost aggregation in existing deposits, as mostly in uplands (Péwé 1975; Kanevskiy et al. 2011; Schirrmeister

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et al. 2011b; Kanevskiy et al. 2014; Ping et al. 2015). During the Holocene (<11.7 ka BP), a more modern permafrost formed under contemporary conditions by deglaciation and vegetation succession under cold climatic conditions, mostly in the sub-Arctic and boreal zone (Zoltai and Tarnocai 1975; Ping et al. 2015). The upper layer of the permafrost, which is subject to annual thawing and freezing, is called the active layer. The active layer thickness (the depth to which it thaws in summer) depends on air temperature, vegetation, drainage, soil or rock type, moisture, snow cover as well as degree and orientation of the slope (van Everdingen 2005). It ranges in the continuous zone from a few decimetres to more than 2 m, whereby it can exceed several meters in the discontinuous zone (Schuur et al. 2008).

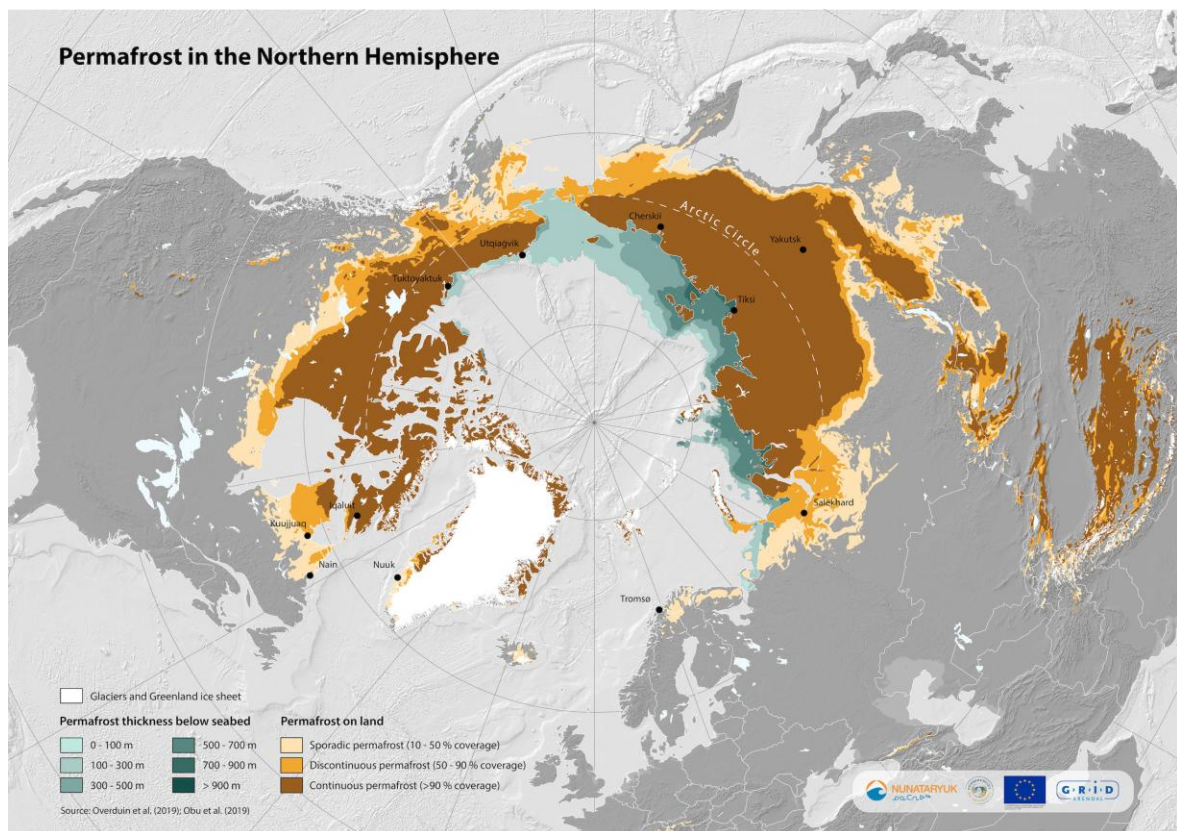


Figure 4. Distribution and extent of the terrestrial and subsea Northern Hemisphere permafrost. GRID-Arendal based data from Obu et al. (2019) and Overduin et al. (2019) adapted by G. Fylakis for the NUNATARYUK project in collaboration with GRID Arendal, 2020.

2.2 Yedoma permafrost

The Yedoma permafrost is a late Pleistocene Ice Complex, formed syngenetic under continental conditions in polygonal tundra with deposits up to 50 m thick. The Yedoma domain covers approximately 2 587 000 km² (11.3 % of the permafrost region) in Russia, Canada and Alaska, where estimated 480 000 km² of this domain is underlain by Yedoma deposits (Strauss et al. 2021b). Yedoma deposits are fine-grained, organic-bearing and ice supersaturated (50 – 95 %) permafrost soils with large syngenetic ice wedges, segregated ice lenses

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and sub horizontal ice bands (Strauss et al. 2012; Schirrmeister et al. 2013; Schirrmeister et al. 2020). Yedoma formed polygenetic due alluvial, fluvial and niveo-aeolian transport, ponding water accumulation, frost weathering and post-depositional processes with syngenetic ice growth, determined by local conditions (Schirrmeister et al. 2020). Due the high amount of ground ice, Yedoma is very extremely vulnerable to permafrost degradation and disturbances (Section 2.3) (Strauss et al. 2017).

2.3 Permafrost degradation, thermokarst and coastal erosion

Permafrost soils are highly sensitive to climate, thus natural or artificial disturbances lead to the degradation of permafrost resulting in a decrease of permafrost thickness or areal extend (Jorgenson 2013). The disturbances can be categorized into (1) presses; impacts due changes on a decadal to century-scale as top-down thaw (active layer deepening), paludification, aridification or changes in the vegetation structure, and (2) pulses; abrupt changes due one-time or episodic short-term events, as wildfires, thermokarst, flooding or rapid soil erosion, which can trigger each other (Grosse et al. 2011).

Thermokarst is the most abundant and recognisable process of permafrost degradation. It results of the thawing of ice-rich permafrost or the melting of excess ice, leading to surface settlement. The initiation depends on hydrology, soil properties, vegetation, geomorphology and disturbance interactions (Grosse et al. 2011; Morgenstern et al. 2011; Jorgenson 2013). The resulting surface depression can be occupied by water, resulting in a thermokarst lake. After the initiation, TLs grow, and potential coalesce by shoreline erosion and further thawing due downward and laterally heat flows and finally drain (partially or completely). The drainage can happen by a changing lake water balance or external factors as melting of ground ice, creation of a drainage pathway, headward gully erosion, river, lake or stream tapping, bank overtopping, coastal erosion or anthropogenic disturbances (Hinkel et al. 2007; Grosse et al. 2013; Jones et al. 2020b; Irrgang et al. 2022; Jones et al. 2022). The resulting DTLBs are optimal habitats for peat accumulation as vegetation can settle and in a cold climate, this basins can refreeze, followed by permafrost development (Jones et al. 2022).

A further consequence of thermokarst processes is the formation of taliks by local anomalies in thermal, hydrological or hydrochemical conditions (van Everdingen 2005). As they can also form sub-aerial due to wildfires or active layer adjustment to a new thermal regime when the summer thaw-depth exceeds the winter freezing-depth (Rey et al. 2020; Farquharson et al. 2022), TLs lead to the formation of sub-aquatic taliks under them and at their

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margins, as they transfer heat from the water body into the underlying ground, especially when the water depth exceeds the maximum thickness of winter ice cover (Burn 2002, 2005; Grosse et al. 2013).

Taliks are therefore thermal disturbances penetrating through permafrost, leading to fast thawing (Burn 2020; Rey et al. 2020; Ohara et al. 2022) and are thus crucial for the permafrost-carbon budget (Section 2.4). The resulting thermokarst landscape occupies ~20 % of the northern permafrost region, TLs and DTLBs are its most abundant recognisable form, with a two – three times higher area of DTLBs than TLs. Often, several TLs and DTLBs generations initiated during the transition between the Pleistocene and the Holocene and during the Holocene Thermal Maximum (11 – 5 ka BP) are present (Romanovskii et al. 2004; Grosse et al. 2013; Fuchs et al. 2019).

As the concentration of TLs and DTLBs increases with the proximity to the coast (Hussey and Michelson 1966), the ACP is strongly influenced by the sea. 34 % of the earth's coasts are affected by permafrost and/or seasonal sea ice cover (Lantuit et al. 2012) and therefore highly influenced by sea-level rise and coastal erosion, which is mainly driven by mechanical thermo-abrasion from wave action and thermo-denudation from insolation and summer warmth (Aré 1988a, 1988b). This leads to a transition from a terrestrial to a marine environment. As the coastal permafrost soils get occupied by the sea, salty seawater inundates them and changes the biogeochemical properties and the mobilization potential of OM into the Arctic ocean (Tagliapietra et al. 2009; Creel et al. 2024; Jenrich et al. 2025).

2.4 Permafrost and carbon

Permafrost carbon is the carbon stored in soils of the northern circumpolar region and in mountain permafrost regions (Schuur et al. 2013). The northern circumpolar region also contains C pools in subsea shelves, originally terrestrial during the last ice ages, which have been little researched yet, but are highly valued (Miesner et al. 2023).

The permafrost is the largest terrestrial C pool, with ~ 30 % of the global C pool on land in only ~15 % of the area (Figure 5). An estimated 1 538 Gt C are stored in the terrestrial permafrost region, with alone 327 – 466 Gt contained in Yedoma deposits in Alaska and Siberia (Strauss et al. 2017; Strauss et al. 2025). A further 2 822 Gt C are expected in subsea permafrost soils (Miesner et al. 2023).

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The main sources of C are plant litter inputs and microbial residues (Ping et al. 2015). Even so the C input is low, the much lower output due the cold and wet soil conditions on permafrost and the long deposition since the Pleistocene enabled this huge C pool. The C accumulation is determined by the landscape topography, vegetation, hydrology, permafrost conditions and the climate (Strauss et al. 2025). The low temperatures of the permafrost soils and the resulting impeding of water movement create a cold and wet environment and thus low decomposition rates due inhibited biochemical processes. The C is locked from the active carbon cycling and preserved like in a freezer for over thousands of years. The ongoing deposition, which buries the OM into deep permafrost deposits, recent peat accumulation which stores OM in the long-term in the soils, and cryoturbation (soil movement due thaw- and freeze-cycles), which relocates and redistributes OM into deeper layers of the perennial frozen soil, stabilize the OM and have led to the large C accumulation and storages in permafrost soils, as seen in Figure 5 (Schuur et al. 2008; Ping et al. 2015; Strauss et al. 2025).

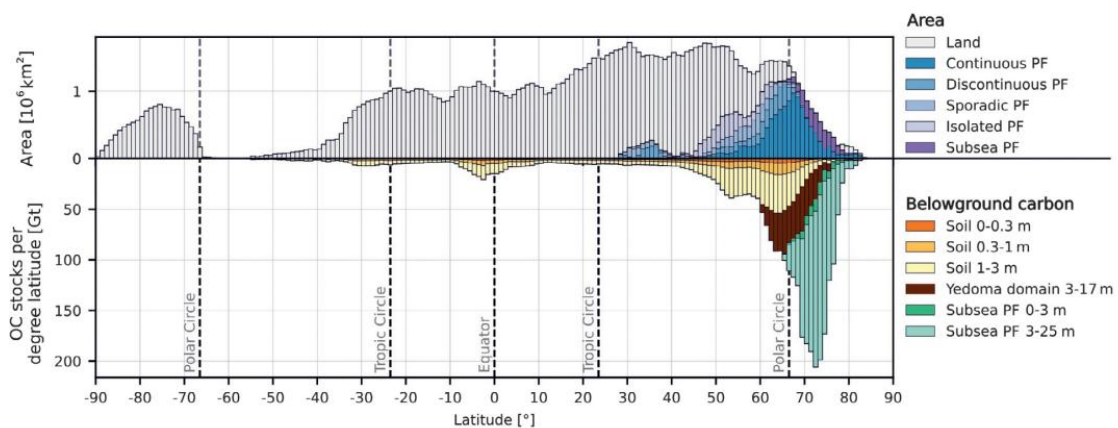


Figure 5. Latitudinal distribution of belowground organic carbon in relation to land mass. Updated data from Poggio et al. (2021), Miesner et al. (2023) and Strauss et al. (2017, 2021b). Figure from Strauss et al. (2025).

This stored OM can be decomposed by microorganisms, as it gets accessible and mobilized due thawing (of the surface layer and the subsurface due abrupt thawing events like thermokarst and talik formation) and erosion. Consequently, CO_2 and CH_4 , depending on the hydrologic conditions, get released to the atmosphere. Thereby, permafrost C plays an essential role for the high-northern carbon cycle (Olefeldt et al. 2016).

Especially thermokarst is important for this budget (Heslop et al. 2015). Figure 6 shows schematically the carbon cycling in TLs and DTLBs. TLs emit high amounts of CH_4 , especially in the initiation phase, originated from the thawed permafrost soils underneath and at the margins of the lakes (taliks) and decomposition of the young OM in the lakes, as well as due shoreline erosion (Walter et al. 2006; Kessler et al. 2012; Walter Anthony and Anthony 2013; Anthony et al. 2014; Heslop et al. 2015; Walter Anthony et al. 2016), followed by CO_2

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sequestration in the later lake phases due sedimentation. Though it is unclear, if this compensates the GHG emissions. The formed taliks under and at the margins of the lakes can then reach and mobilize old, deep carbon that wouldn't be tapped with only gradual top-down thawing (Walter Anthony et al. 2018).

Anthony et al. (2014) estimated that $28 \pm 12\%$ of the thawed C under permafrost lakes is converted into GHG over the lakes' millennial-scale lifespans. The C balance in DTLBs is still uncertain and strongly depends on the hydrologic conditions, vegetation, depth of the active layer and the time since drainage. DTLBs can act as a C source, especially shortly after the drainage by releasing CO_2 and small amounts of CH_4 due microbial decomposition. Since the basins favour peat accumulation and a fast succession from fen-type wetland to bog and tundra wetlands, they can then become a C sink, supplemented through new permafrost aggregation in a cold climate (Wickland et al. 2009; Anthony et al. 2014). The estimated C fluxes of DTLBs are one – three magnitudes smaller than in TKLs, with high CO_2 uptake offset and CH_4 emissions at the beginning, which declines with time (Walter Anthony et al. 2018; Zona et al. 2010; Zona et al. 2012; Treat et al. 2021; Jones et al. 2022). Thereby, thermokarst landscapes are a dynamic, uncertain component in the permafrost carbon budget, as they act as a C source through thaw and erosion processes on one hand and can

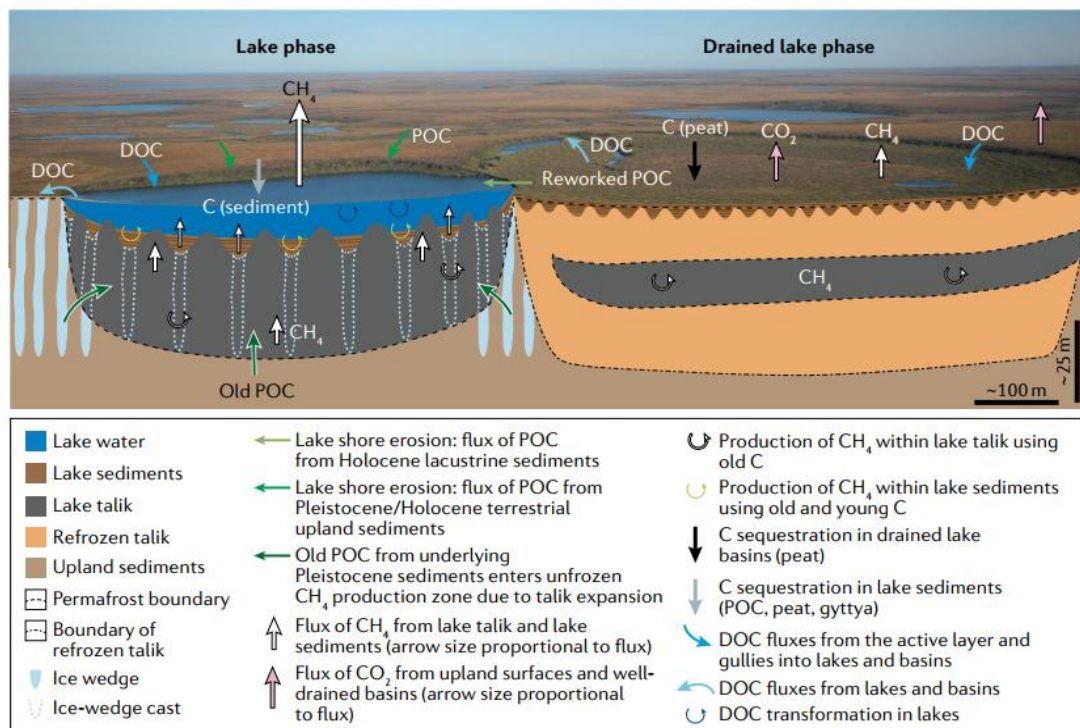


Figure 6. Carbon cycling of a lake and drained lake basin system in lowland permafrost. Oblique aerial photograph of a thermokarst lake and drained lake basin in ice-rich permafrost on the northern Seward Peninsula, Alaska. Shown in cross section are idealized schematics of the underlying talik and permafrost configurations associated with the lake and drained lake basin phases, and key surface and subsurface carbon (C) cycling components indicated by the arrows. Background image courtesy of Lawrence Plug, Figure by Jones et al. (2022).

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be a C sink in DTLBs on the other (Anthony et al. 2014; Fuchs et al. 2019).

2.5 Permafrost in a warming climate

Responding to climate warming, permafrost ground temperatures are on high records with increasing rates of up to 0.4 - 0.8 °C per decade since the 1980s (Smith et al. 2022). Between 2007 and 2016, the global permafrost temperatures have increased 0.29 ± 0.12 °C, with higher rates for the continuous (0.39 ± 0.15 °C) than for the discontinuous (0.20 ± 0.10 °C) zone, indicating ongoing thawing and active layer thickening in the warmer regions as the heat is being absorbed by the phase change of ice to water, instead of directly rising the ground temperatures (Biskaborn et al. 2019). This is of particular concern as it mobilizes the stored OM due active-layer deepening, thermokarst development, riverbank and coastal erosion, thaw subsidence, forest and tundra fires (Strauss et al. 2025). The permafrost warming followed by gradual and abrupt thaw brings back the C into the active C cycling and enhances microbial decomposition causing CO₂ and CH₄ release into the atmosphere, which in turn causes further climate warming, called the “permafrost carbon feedback” loop, Figure 3 (Schuur et al. 2022). As a result, it is likely that the northern permafrost region transitions over the long term from a C sink to a C source, with substantial C losses after 2100 (McGuire et al. 2018). The sixth assessment report of the IPCC estimates the permafrost GHG feedback from CO₂ per degree of global warming at the end of the century is 18 (3.1 – 41) Gt C °C⁻¹ and 2.8 (0.7 – 7.3) Gt CO₂-eq °C⁻¹ for CH₄ (IPCC 2023b). Schuur et al. (2022) states that the thawing of terrestrial permafrost could release 5 – 15 % of its C pool during this century, leading to calculated net CO₂ and CH₄ (as CO₂ equivalents) emissions by 2100 of 55 – 232 Gt C-CO₂-eq. This is in the range of an industry nation as Russia, OECD, USA and China (scaled to 100 a), while these calculations do not yet include abrupt thaw processes as thermokarst, which could add another 40 % to the projections of C release (Schuur et al. 2022).

Previous studies found that the recent climate warming and rising permafrost temperatures have already led to an increase in the frequency and magnitude of thermokarst (Fedorov et al. 2014; Lantz and Kokelj 2008; Jorgenson et al. 2006), and thus an increase in TLs and DTLBs, with the number of water bodies increasing by 10.7 %, while the area is decreasing due the drainage of large water bodies by 14.9 % (Jones et al. 2011). Further, Jones et al. (2022) states a shift from TLs to DTLBs. While DTLBs have under cold climate the potential of peat accumulation and refreezing and thus of acting as a net C sink, the future warming inhibits this permafrost re-aggregation and causes persist taliks, which can mobilize old and

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deep permafrost carbon, contrary to active-layer deepening, which mostly impacts shallow, young carbon (Jones et al. 2022; Parazoo et al. 2018). Additionally, warming permafrost and thus subsiding permafrost landscapes are accelerating the coastal erosion (Biskaborn et al. 2019; Lim et al. 2020; Jones et al. 2020a). This is among further factors as reduction of sea-ice extent and increasing open water period duration (Perovich et al. 2020), rising air and water surface temperatures (Ballinger et al. 2020; Timmermans and Labe 2020), sea-level rise (Moon et al. 2020) and stronger wave action (Casas-Prat and Wang 2020), resulting in some of the highest coastal erosion rates on Earth with up to 47 m a^{-1} , as seen in Figure 7 (Irrgang et al. 2022).

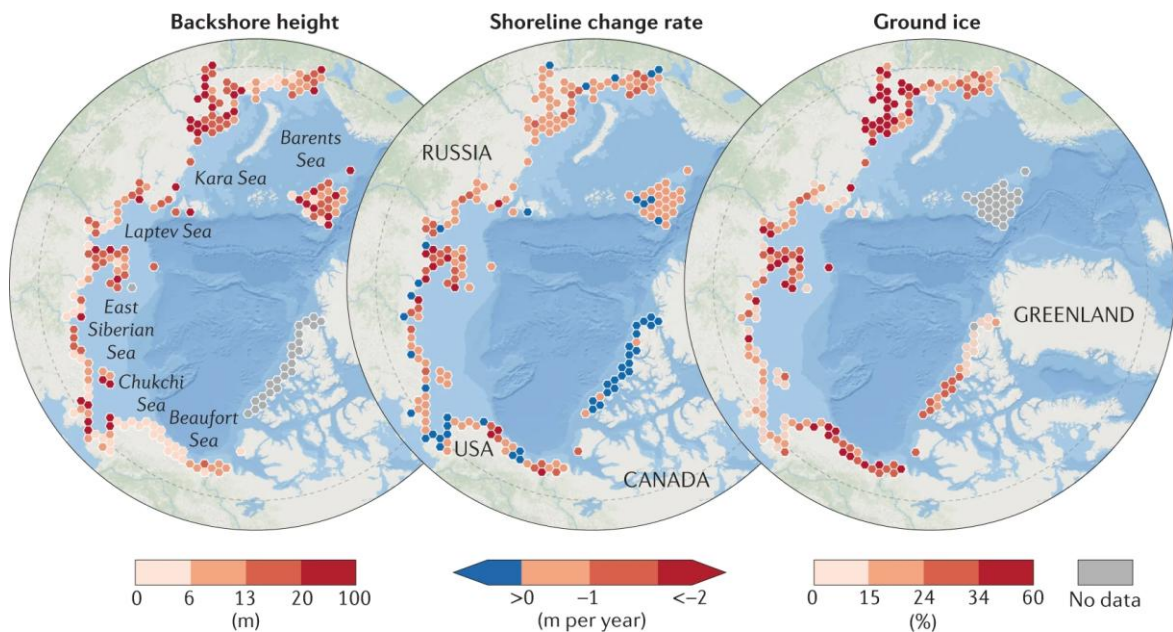


Figure 7. Change of Backshore height, shoreline change rate and ground ice content along the Arctic coast. Data used from the Arctic Coastal Dynamics database (Lantuit et al. 2020) and summarized over a $100 * 10^3 \text{ m}$ extent. Figure from Irrgang et al. (2022).

Creel et al. (2024) found that under medium emission scenarios, the Arctic coastline will lose $6\,638 \text{ km}^2$ of land due to erosion, permafrost subsidence and sea-level rise by 2100. Respectively, the estimated OC disturbance is 0.453 Gt. Also, taliks are expected to highly increase in frequency and extend, e.g. Parazoo et al. (2018) simulated that from 2100 to 2300, taliks will widespread form in the northern high latitudes, followed by a C sink-to-source transition. In the discontinuous zone, taliks will form across up to 70 % of the area by 2030 and reach thicknesses of 12 m by 2090. Figure 8 shows this C source to sink transition.

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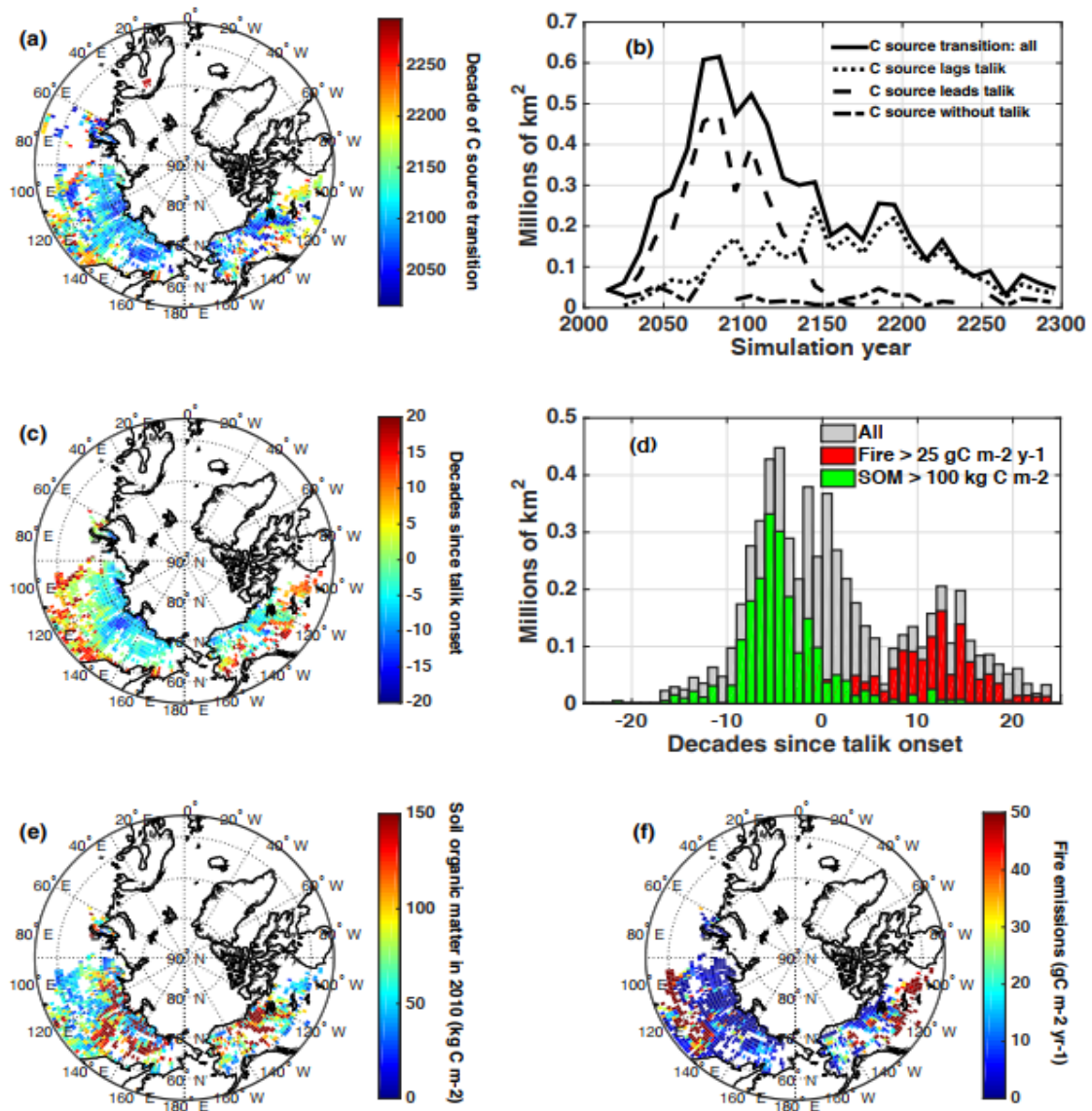


Figure 8. C sink to source transition from 2010 to 2300 in relation to talik onset, soil C, and fire emissions under a Representative Pathway 8.5 warming scenario through 2100 and an Extended Concentration Pathway 8.5 through 2300. (a) Map of the decade of shift to C source. (b) Land area that transitions in the late 21st century and is driven by regions where the C source leads talik onset (dashed). (c) Decadal time lag from talik onset to C source shift. (d) Trimodal distribution of permafrost area as a function of decadal time lag (negative lags related to high soil organic matter (green bars and map in e) and large positive lags related to fires (red bars and map in f)). Figure from Parazoo et al. (2018).

3. Study area

3.1 Regional setting

The study area is located near Kotzebue (Qikiqtaġruk), on the north coast of the Baldwin Peninsula in north-west Alaska, surrounded by Kotzebue Sound. It is in the continuous permafrost zone, near the transition into the discontinuous zone (Jorgenson 2013; Obu et al. 2019) and part of the Bering Land Bridge (Nitze et al. 2020). The region is characterized by a subarctic continental climate with a mean annual air temperature of $-4.4\text{ }^{\circ}\text{C}$ and 289 mm annual precipitation (Kotzebue Climate station, Station USW00026616, NOAA, 1991 – 2020). The average snowfall accumulation is 163 cm a^{-1} , with snow persisting until middle to end of May (Macander et al. 2015). The measured ground temperatures range between -3.53 and $-0.8\text{ }^{\circ}\text{C}$ (Biskaborn et al. 2019).

The Baldwin Peninsula is a middle Pleistocene push-moraine complex at elevations from sea level to ~ 50 m a.s.l. with marine, fluvial and glaciogenic sediments, partly covered by loess deposits, formed during the Anaktuvuk River glaciation 500 to 600 ka BP. The region is underlain by ice rich (> 40 vol%) Yedoma permafrost with large syngenetic Pleistocene ice wedges as well as signs of strong degradation (Karlstrom 1964; Huston et al. 1990; Jorgenson et al. 2008; Jones et al. 2011; Jones et al. 2012; Jongejans et al. 2018).

It contains to the major lake districts of Alaska with lakes mostly in DTLBs from previous lake generations (Arp and Jones 2009; Jones et al. 2011; Jongejans et al. 2018; Nitze et al. 2020). The vegetation is dominated by a shrubby tundra (Walker et al. 2005) with floating vegetation on the lake margins and above water (Parsekian et al. 2011).

3.2 Sampling transect

This study covers a transect of ~ 1600 m from the inland to the coast with the four landscape units Yedoma upland (UL), thermokarst lake (L), drained lake basin (DLB) and marine (M), with elevations between 0 and 35 m a.s.l. shows the digital elevation model of the study area with the four sampling sites. The drained lake basin is a former Thermokarst lake on the Schaeffer-Tessier Family Native allotment (Illivak) near Kotzebue. It is originally made up of two lakes which coalesced till 1974 and increased with expansion rates of $0.25 - 0.35\text{ m a}^{-1}$ up to an area of 0.54 ha in 2021 and bank heights of 2 – 4 m. The estimated average depth before the drainage is 3 – 4 m, the estimated mean permafrost thaw subsidence magnitude due active sub-lake thermokarst processes 6.5 m (Jones et al. 2023).

3 Study area

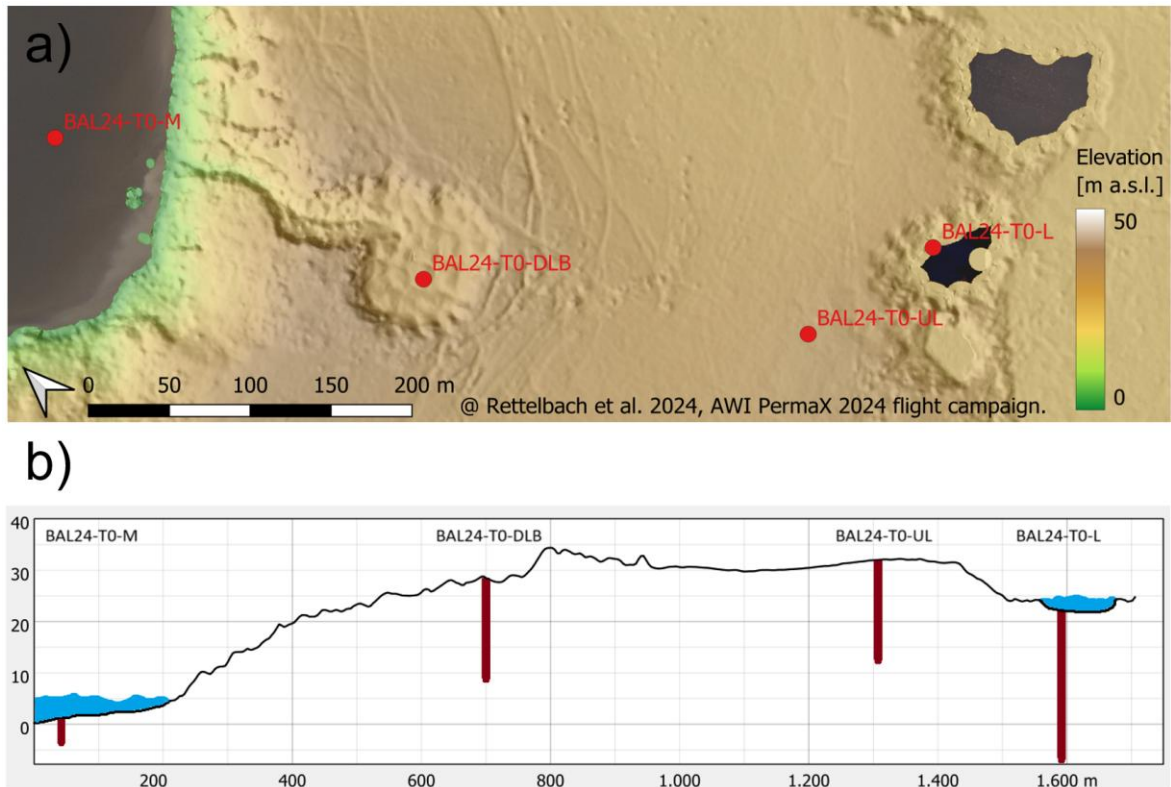


Figure 9. (a) Digital elevation model and (b) horizontal profile of the study transect showing the marine site, the drained lake basin, the upland and the thermokarst lake. Core lengths are in the scale 1:10. Data from the AWI PermaX 2024 flight campaign (Rettelbach et al. 2024). Created with QGIS (QGIS Development Team 2024).

The drainage took place on 29th June 2022 and could be observed with the Native Tribal Health Consortium's LEO Network (Tessier et al. 2022). The resulting drainage tunnel, Figure 10, is 125 m long and eroded up to 14 m into the ice-rich permafrost, releasing estimated 18 500 m³ water and 8 500 m³ material through erosion and thaw into Kotzebue Sound. As beavers were present the days before and strongly influence thermokarst lake dynamics, they likely highly contributed to the drainage (Jones et al. 2020c; Jones et al. 2023).

3 Study area

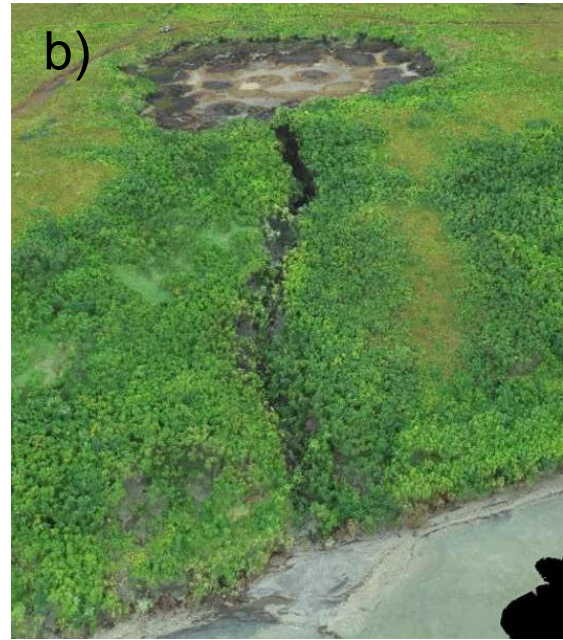
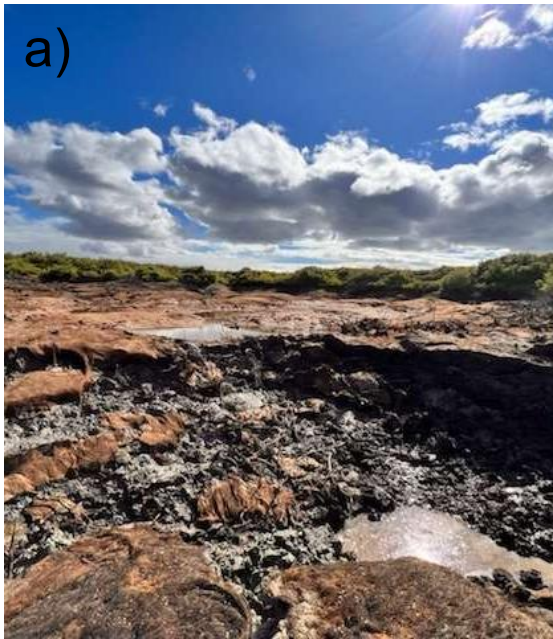


Figure 10. (a) The drained lake basin after the drainage, picture by Susan Tessier, (b) a drone photo by Ben Jones from the drained lake basin with its gully in summer 2022, (c) the drained lake basin in summer 2024, picture by Maija Marushchak, and (d) the erosion gully in summer 2024, picture by Lutz Schirrmeister.

4. Methods

To answer the research questions regarding the environmental history, OM sources, quantity and quality and its development due degradation, this study involved fieldwork, laboratory work and a statistical analysis.

4.1 Fieldwork

The field work took place in March 2024 during the AK-Land_2024_NWAlaska_spring Expedition (07.03.2024 – 28.03.2024) near Kotzebue on the Baldwin Peninsula, Northwest Alaska, led by Jens Strauss, as a collaboration of the University of Alaska Fairbanks and the Alfred Wegener Institute Helmholtz Centre for Polar and Marine research (AWI). For this master thesis, four cores under different landscape conditions (Table 1) were drilled on a transect from the inland to the coast (Figure 9). All sites were covered with a layer of 0.5 – 28.5 cm snow. The thermokarst lake (BAL24-T0-L) and marine site (BAL24-T0-M) were also covered by ice and water. The depth of the lake was 5.76 cm (1.26 cm ice and 4.5 cm liquid water), the ice on the marine site was 92 cm thick with 21 cm liquid water beneath. For the two frozen cores from the upland and the drained lake basin (BAL24-T0-UL and BAL24-T0-DLB), a “Snow-, Ice- and Permafrost-Establishment” (SIPRE, Ø 7.5 cm) was used. The unfrozen cores were drilled with a Vibracoring-system (WINK Vibracorer, Ø 7.5 cm) for the BAL24-T0-L and a Push Corer (Ø 7 cm) for BAL24-T0-M. The Cores BAL24-T0-UL and BAL24-T0-DLB were packed into whirlpacks and core foil, BAL24-T0-L in an aluminium tube, BAL24-T0-M in a plastic tube. They were labelled and stored below 0 °C in styrofoam boxes for the transportation to AWI Potsdam, where they were stored at around -20 °C until further analysis.

Table 1. Characteristics of the four sites.

Sample ID	Landscape unit	Coordinates	Core length [cm]	Sample number	Coring device
BAL24-T0-UL	Upland permafrost ground	66.903288°N, 162.508495°W	213	39	SIPRE (Ø 7.5 cm)
BAL24-T0-L	Thermokarst lake ground	66.903148°N, 162.506359°W	311	61	VIBRA (Ø 7.5 cm)
BAL24-T0-DLB	Drained thermokarst lake basin	66.905016°N, 162.511771°W	214	42	SIPRE (Ø 7.5 cm)
BAL24-T0-M	Marine ground	66.907028°N, 162.513997°W	55	12	PUSH (7 cm)
Total				155	

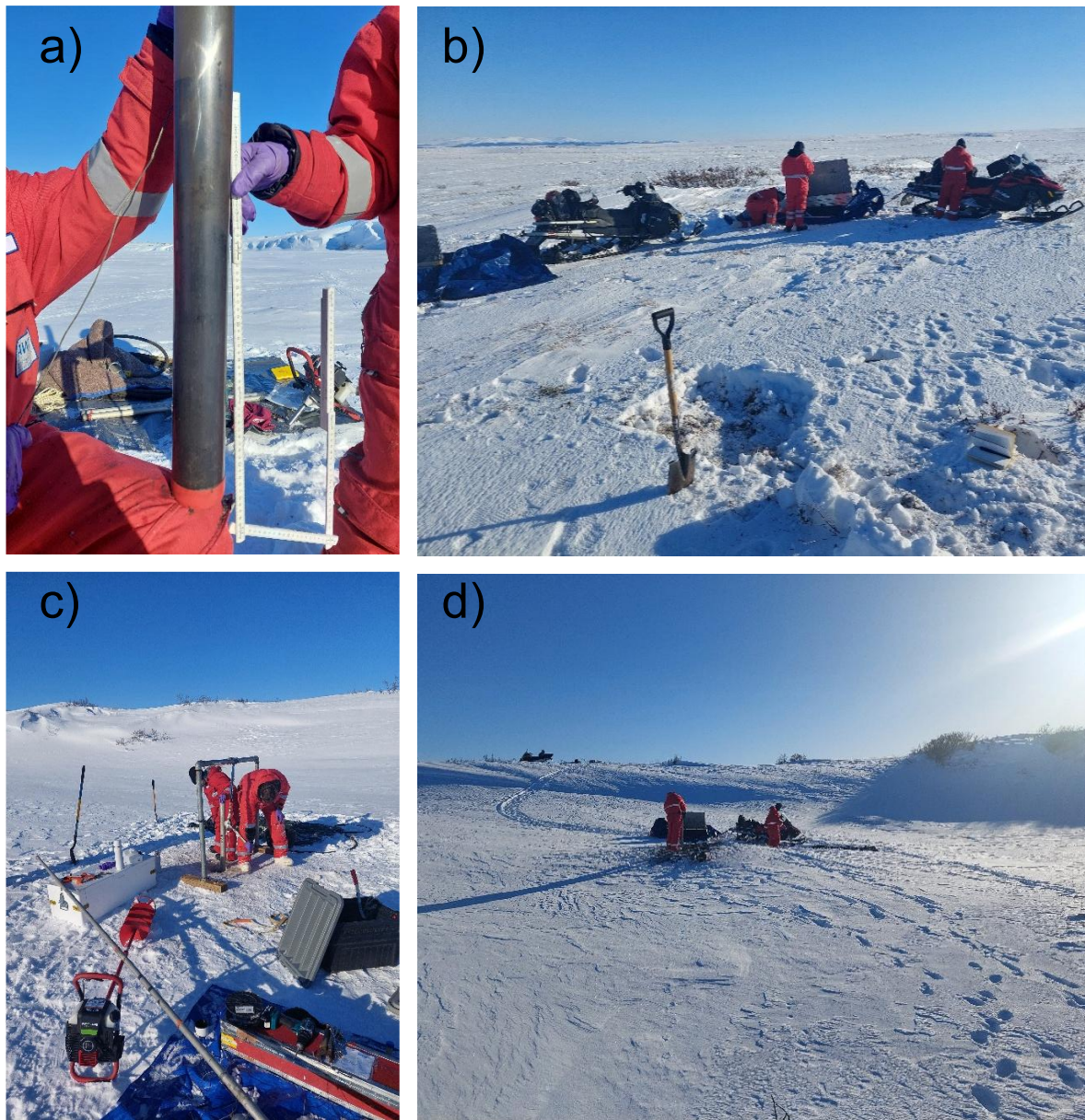


Figure 11. Impression of the field work during the AK-Land 2024 NWAlaska spring Expedition. (a) The marine core, BAL24-T0-M, (b) the upland study site, (c) the procedure of the virbracoring at the thermokarst lake site and (d) the drained thermokarst lake basin; Images from Fabian Seemann.

4.2 Laboratory analysis

A multi-proxy approach was used for answering the different objectives, containing geochronology and sedimentology, biogeochemistry and hydrochemistry and n-alkane biomarker analysis. Figure 12 shows a schematic of the laboratory work.

The cores were subsampled in a climate chamber with around $-8\text{ }^{\circ}\text{C}$. They were cut in half lengthwise with a band saw, resulting thawed and refrozen material was removed with knives. Afterwards the profiles were photographed and described sedimentological and cryolithological according to French and Shur (2010). One core half was packed in core foil

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and whirlpacks for the archive. The other was subsampled in approximately 2 – 10 cm increments for laboratory analyses, according to their cryostructure. From each core, three to five samples from the upper, middle and lower parts were taken for the biomarker analysis. The subsamples were then packed in labelled whirlpacks, respectively pre-burned, weighted

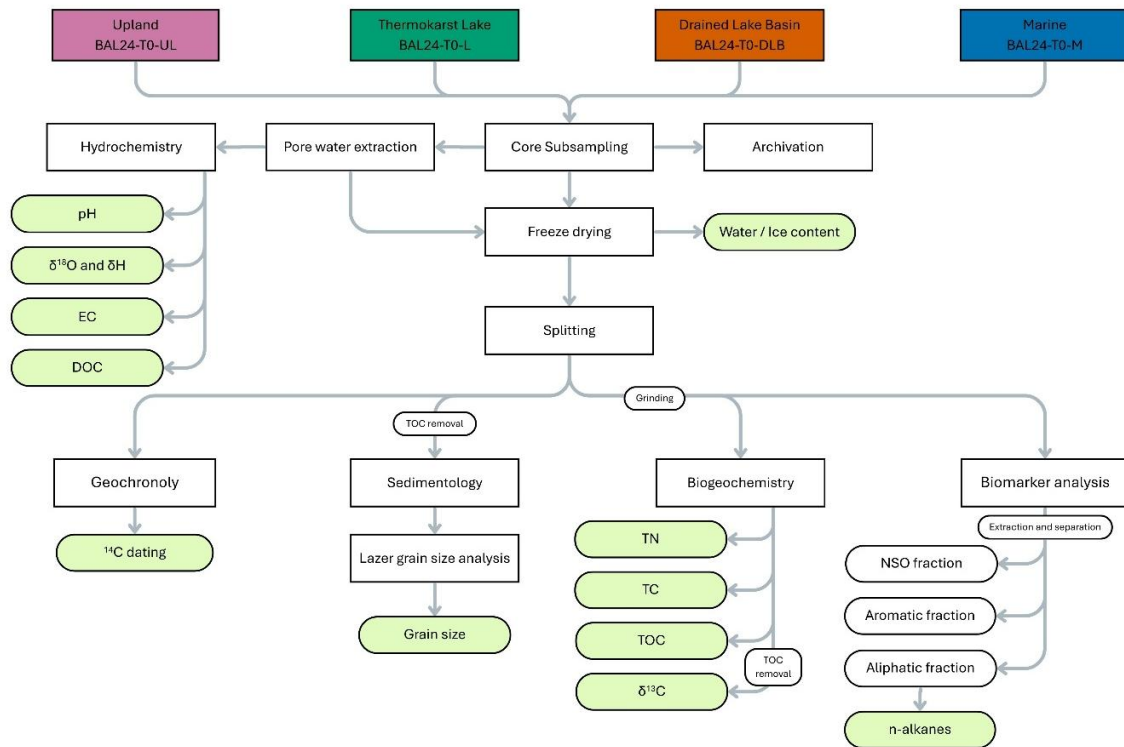


Figure 12. Flowchart of the used laboratory methods.

glass jars for biomarker analysis, weighted, and stored at -20°C . Figure 13 shows the subsampling of BAL24-T0-UL in the climate chamber at AWI.

A total of 155 samples was obtained for sedimentology and biogeochemistry analysis, from which 63 were additionally analysed for hydrochemistry, 14 for geochronology and 20 for biomarker analyses.

4.2.1 Hydrochemical analysis

63 of the subsamples were analysed for the hydrochemical parameters electrical conductivity (EC), pH-values, dissolved organic carbon (DOC) and stable water isotopes ($\delta^{18}\text{O}$ and δD). Investigating the properties of the porewater is key to understand the hydrochemical composition of the OM and the paleoenvironmental conditions as temperature, origin of water and genesis processes.



Figure 13. (a) Transported cores stored in whirlpacks and (b) cryolithological description and subsampling of BAL24-T0-UL in the climate chamber at AWI Potsdam.

4.2.1.1 Porewater extraction

For the porewater extraction, the samples were thawed at 6 °C in the whirlpacks. Rhizomes (*RHIZONS MOM 5 and 10 cm, Rhizospheres Research Products*) were inserted into the samples and taped airtight. Then a 30 mL syringe was stacked on the rhizomes and filled with the porewater by drawing up and creating a vacuum in the whirlpack for 1 – 1.5 h (Figure 14). The extracted porewater was injected into labelled glass bottles. This procedure was repeated three – five times.



Figure 14. Porewater extraction.

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4.2.1.2 Electrical conductivity and pH-value

The EC reflects the concentration of dissolved ions in the porewater. It can serve as an indicator of mineralization of the OM and thus the degradation of permafrost, as well as hydrological processes and the marine influence, as ions and salts get released into the porewater due permafrost degradation. Also repeated thaw-freeze processes can lead to an ion concentration, for example at the thawing front. The pH-value influences chemical processes and can provide information about the microbial activity and solubility of organic matter; low pH-values can limit decomposition rates (Grosse et al. 2011). As the analysis of the pH-value changes the chemistry of the porewater, it is essential to first measure the electrical conductivity. The analysis was carried out with 3 mL of the extracted porewater in a *Seal Analytics, Orion Versa Star Pro* laboratory robot

4.2.1.3 Oxygen and hydrogen isotopes

The stable water isotopes are the ratio of $^{18}\text{O}/^{16}\text{O}$ for $\delta^{18}\text{O}$ and $^2\text{H}/^1\text{H}$ for δD , respectively. As the chemical reaction or phase transition of water leads to a fractionation of the isotopes, and therefore a change in the isotopic ratio, they can be used to indicate the temperature, evaporation or moisture source of the water when infiltrated into the soil. E.g. clouds consist of the lighter isotopes due their higher vapour pressure and lakes or the ocean of the heavier isotopes. Further, the isotope fractioning is temperature depending, whereby it can be used as temperature indicator.

For the stable water isotope analytics, around 10 mL of the cooled porewater was measured with a *Finnigan MAT Delta-S Mass Spectrometer*. The values were obtained by the average of two measurements. The results were expressed in permille (‰), using the internal Standard Mean Ocean Water (SMOW) as reference (Craig 1961):

$$\delta_{sample, SMOW} = \left(\frac{R_{sample}}{R_{SMOW}} - 1 \right) * 1000, \quad (1)$$

where R_{sample} and R_{SMOW} are the isotopic ratios in the sample and SMOW reference and $\delta_{sample, SMOW}$ is the isotopic abundance in the sample relative to SMOW (Craig 1961).

4.2.1.4 Dissolved organic carbon content

The dissolved organic carbon (DOC) reflects the microbial activity and the release of OM into the environment (high = high availability, low = low mobility and stable conditions). The DOC was measured as non-purgeable organic carbon (NPOC, direct method) under the

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assumption that no significant volatile organics were in the sample. For this, 10 mL of the porewater was acidified with 50 μL of 30 % hydrochloric acid (HCL) to convert the inorganic carbon (IC) component into CO_2 via carbonic acid. Afterwards the CO_2 is removed from the sample solution by purging and DOC is directly measured as NPOC by oxidation at 680 $^\circ\text{C}$ followed by detection. For the analysis, 10 mL of the porewater was mixed with 50 μL of 30 % HCL to preserve the samples until the measurement. The measurement was carried out with a *Shimadzu TOC-VCPH*, the results of three to five injections were averaged. The detection limit was 0.3 mg L^{-1} .

4.2.2 Sedimentological analysis

For further analysis, all samples were freeze-dried (*Zirbus Sublimator*) at -40 $^\circ\text{C}$ and 0.02 MPa for 48 h and then weighted again. The water / ice content and grain size were measured for the sedimentological analysis.

4.2.2.1 Water / Ice content

The absolute water / ice content was obtained due the weight difference between the frozen and the freeze-dried samples after Equation (2):

$$\text{abs. water content [wt\%]} = \frac{m_w - m_d}{m_w} * 100, \quad (2)$$

where m_w is the netto wet weight [g] and m_d the netto dry weight [g].

4.2.2.2 Grain size analysis

The grain size and its distribution give information about the transport medium and the mechanism of deposition during the sedimentation. This is important to reconstruct the sedimentological and depositional history of the study site. Further, the grain size influences soil properties and is therefore important for interpreting the biogeochemical results.

For preparation of the grain size analysis, samples with to high peat content were separated out and the OM of the remained samples was removed via oxidation. Therefore, 100 mL of 3 % hydrogen peroxide (H_2O_2) and 2 – 3 drops of 25 % ammonia (NH_3) were added to the freeze-dried samples. They were then placed on a shaker, which got slowly heated up to 60 $^\circ\text{C}$ after a few days, for ~ 4 weeks to allow all OM to react (Figure 15). 4 – 5 times a week, further 10 mL 30 % H_2O_2 was added, as well as ammonia or acetic acid to ensure the pH remains at near-neutral (6 – 8).

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After all organic material reacted, the samples were washed out with purified water to remove the H_2O_2 , centrifuged (*Heraeus Multifuge 3s*, *Heraeus Cryofuge 8500i*, *Heraeus Megafuge 40*) to remove supernatant liquid, freeze-dried and then manually homogenized. Approximately 1 g of the samples were mixed with 0.5 g dispersing agent (tetra-sodium pyrophosphate, $Na_4P_2O_7$) and 0.0001 % ammonia solution (NH_3) in a plastic jar and shaken (*RS12 Rotoshake*, *Gerhardt*) for at least 24 h for complete dispersion. The samples were then divided into 8 subsamples with a particle concentration of 5 – 15 % with a rotary cone sample divider (*Laborette 27*, *Fritsch*). To prevent damages on the laser, particles >1 mm were sieved, weighted and afterwards included in the grain size distribution, if the proportion was significant compared to the total sample.

The grain size analysis was carried out with a *Malvern Mastersizer 3000* laser. This measures the light impulses on multiple scatter light detectors after particle refraction due a red laser (633 nm) and a blue light emitting diode (470 nm). Three measurements per subsample were taken and the results averaged. The grain size statistics as the mean, mode, sorting and skewness were calculated after Folk and Ward (1957) with the software GRADISTAT v 8.0 (Blott and Pye 2001), using geometric scaling after the international ISO 14688-1:2017 (ISO 2017), with clay ≤ 2 μm , silt 2 – 63 μm and sand 63 μm – 2 mm.

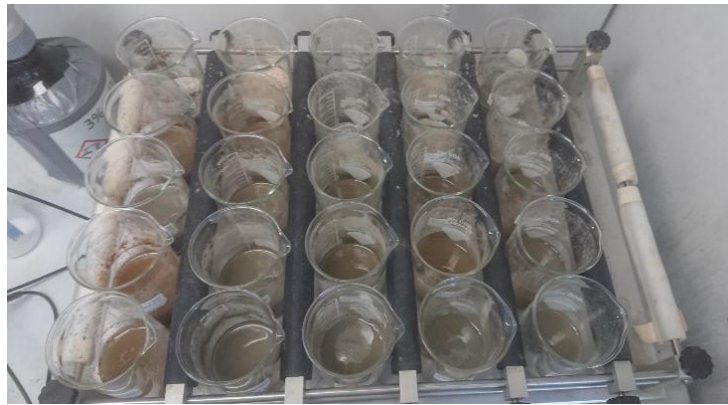


Figure 15. Sediment samples on the shaker.

4.2.3 Geochronology

Radiocarbon dating uses the decay of the radioactive carbon isotope ^{14}C into ^{14}N to determine the age of an organism. During live, the ^{14}C concentration in an organism remains in equilibrium with the environment, while after death, the original ^{14}C content decays with $5\,700 \pm 30$ a as half-life time and thus the remaining content of ^{14}C indicates the time since death (Hajdas et al. 2021).

14 of the samples were selected for geochronology. The freeze-dried samples were picked

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for macrofossils as leaves and branches under the microscope. The radiocarbon age was determined using the accelerator mass spectrometer *MICADAS (Mini CARbon Dating System, Ionplus)* at AWI Bremerhaven. To obtain a chemically unmodified fraction of the samples, they were pre-treated with the acid-base-acid (also known as acid-alkali-acid) treatment to remove contaminants as carbonates, organic acids and dissolved atmospheric carbon dioxides (Vries and Barendsen 1954; Mollenhauer et al. 2021). The samples were converted into CO₂ by combustion at 950 °C with an elemental analyser (*Elementar vario Isotope*), graphitized with an automated graphitization system (*AGE-3; Ionplus AG*) and then pressed into cathodes via pneumatic press, which were inserted into the linear magazine for measurement (Mollenhauer et al. 2021). The detection limit was >50 ka BP.

Afterwards the calibration software *CALIB 7.1* (Reimer et al. 2013) was used to express the determined radiocarbon ages in calibrated kilo years before present (cal. ka BP).

4.2.4 Biogeochemical analysis

The biogeochemical analysis gives important information about the characteristics of the OM, as the degree of decomposition and the vulnerability, as well as about the paleoenvironmental conditions. For this, the freeze-dried samples were homogenized by grinding (*Pulverisette 5 planetary mill, Fritsch*) for the further analysis.

4.2.4.1 Total organic carbon and nitrogen

The elemental analysis bases on combusting chromatography. For the carbon fractions, two 50 mg sub-samples per samples were weighted into stainless steel capsules and measured with an elemental analyser (*ELEMENTAR soli ToC cube*, Figure 16a). Between the three

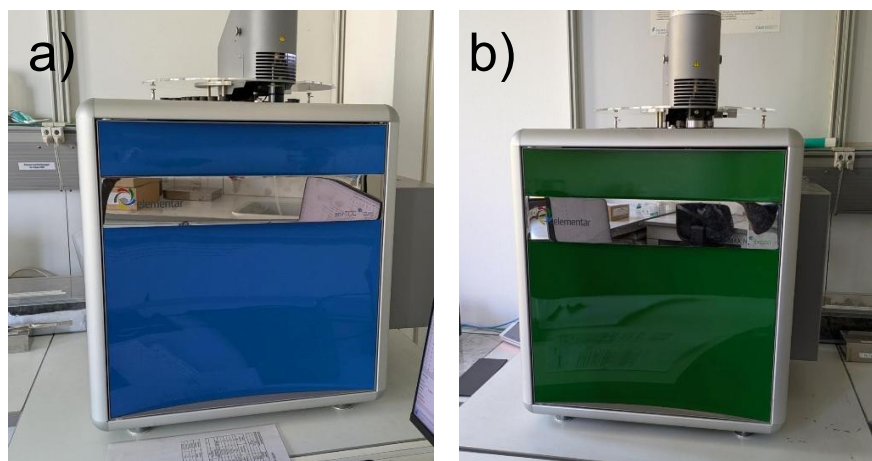


Figure 16. (a) The Soli TOC cube and (b) the rapid Max N exceed.

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fractions total inorganic carbon (TIC), total organic carbon (TOC) and residual organic carbon (ROC) was distinguished with a temperature ramping method after DIN 19539 in a flow of oxygen-containing gas. With a heating rate of 70 °C min⁻¹, the sub-samples were heated to 400 °C for TOC (holding time 230 s), 600 °C for ROC (holding time 120 s) and finally 900 °C for TIC (holding time 150 s).

Total nitrogen (TN) was measured with a *Rapid Max N exceed* (Elementar, Figure 16b) using the Dumas method (Buckee 1994). 50 mg of the samples were weighted into stainless steel capsules and heated up to 900 °C. Oxygen was introduced with a flow rate of 120 mL min⁻¹ for samples with a TOC content ~ 1 wt%, 140 mL min⁻¹ for a TOC content ~20 % and respectively 160 mL min⁻¹ for samples with a TOC content ~40 wt%. To detect background signals, blank capsules and calibration standards were included in the measurement for both procedures (

Table 2a for TOC and

Table 2b for TN). The detection limit of both analysers is <0.1 wt%.

Table 2. Insertion and weighing-in instruction for soli TOC cube (left) and for Rapid Max N exceed (right).

Number of runs	Description	Weigh-in	Number of runs	Description	Weigh-in
1x	RunIn	set to 1 mg	1x	RunIn	set to 100 mg
2x	Blank	set to 1 mg	2x	RunIn (N 10.313 %)	~250 mg
3x	CaCO ₃ 6% (TC 6 %)	20 mg	2x	Blank	set to 100 mg
3x	IVA 2176 (TOC 15.57 %)	50 mg	4x	EDTA 5:45 (N 0.960 %)	50 mg
1x	CaCO ₃ 6% (TC 6 %)	20 mg	1x	BS 1 (N 0.216 %)	50 mg
1x	CaCO ₃ 12% (TC 12 %)	15 mg	1x	BS 2 (N 0.064 %)	50 mg
1x	EDTA 5:45 (TOC 4.11 %)	100 mg	1x	IVA 2150 (N 0.490 %)	50 mg
1x	IVA 2176 (TOC 15.57 %)	50 mg	1x	IVA 2156 (N 1.360 %)	50 mg
15x (2)	Samples	50 mg	36x (2)	Samples	50 mg
	Control block			Control block	
15x (2)	Samples	50 mg	1x	Blank	set to 100 mg
	Control block				
2x	Blank	set to 1 mg			

The measured TOC and TN content was further used to calculate C/N as the quotient of TOC and TN. This ratio is an indicator of degree of OM decomposition. A lower C/N indicates a higher a degree of decomposition, as the degradation changes the elemental composition of the OM components because carbon gets more removed than nitrogen during decomposition

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(Kuhry and Vitt 1996; Schirrmeister et al. 2011b; Strauss et al. 2015). Furthermore, C/N can be used to distinguish the OM source, between algal (4 – 10) and land-plant (>20) input (Meyers 1994). As the source identifiers from Meyers (1994) are given in atomic C/N, they were converted into wt% with $C/N (wt\%) = C/N (atomic) * \frac{12.011 u}{14.007 u}$, where 14.007 u and 12.011 u are the atomic weights of N and C. This results in the values of 3.4 – 8.6 for aquatic algae and ≥ 17.1 for land plants. But as C/N is influenced by the grain size, with coarser fractions often include more visible plant remains, while finer, clay-rich fractions tend to have lower nitrogen levels because clay minerals can absorb ammonia and reduce its measurable concentration, it is important to use additionally indicators for the OM source, such as $\delta^{13}C$ or the biomarker proxies (Section 4.2.5.2) (Meyers 1994, 1997).

4.2.4.2 Stable carbon isotopes

The stable carbon isotopes ($\delta^{13}C$) are used to identify the source and decomposition state of the OM. Plants differ in their carbon isotope fractionation, e.g. marine (-22 to -20 ‰ vs. VDPB) vs. lacustrine and continental (\sim -27 ‰ vs. VDPB) plants, allowing $\delta^{13}C$ to serve as a proxy for vegetation input, especially as it is compared to C/N not influenced by the grain-size (Meyers 1994, 1997). Furthermore, microbial decomposition preferentially uses the lighter ^{12}C isotope, resulting in less negative $\delta^{13}C$ values in more degraded material (Heyer et al. 1976). Thus, $\delta^{13}C$ values help to infer both past vegetation and the preservation state of OM in sediments.

The measurement of $\delta^{13}C$ was carried out with a *Finnigan MAT Delta-S* mass spectrometer combined with a *FLASH elemental analyser* and a *CONFLO IV* gas mixing system. To remove carbonate, samples were treated with 20 mL HCL (1.3 mol) and heated at 97 °C for 3 h prior measuring. Afterwards, the samples were washed with purified water to a neutral pH-value, filtered, dried at 50 °C and weighted in silver capsules depending on the sample TOC content (Equation (3).

$$target\ sample\ weight\ [mg] = \frac{20}{TOC\ [wt\%]} \quad (3)$$

The sub-samples were combusted at 1020 °C in a O₂ atmosphere with chrome dioxide (CrO₂) as oxidant, to convert the OC into CO₂, which then was transferred to the mass spectrometer via the *CONFLO IV* gas mixing system. With the calculated ratio of ^{13}C to ^{12}C , the $\delta^{13}C$ value was expressed in parts per thousand with the permille (‰) notation, using the internal standard Vienna Pee Dee Belemnite (VPDB), after equation (4):

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$$\delta^{13}C_{sample, VPDB} = \left(\frac{I^{13}R_{sample}}{I^{13}R_{VPDB}} \right) - 1, \quad (4)$$

where $I^{13}R_{sample}$ and $I^{13}R_{VPDB}$ are the $^{13}C/^{12}C$ ratios in the sample and VPDB reference and $\delta^{13}C_{sample, VPDB}$ is the isotopic abundance in the sample relative to VPDB (Hoffman and Rasmussen 2022).

4.2.5 Biomarker analysis

Biomarkers are characteristic molecular markers which contain important information about the OM origin and preservation. Thus, biomarkers are a key indicator for paleoenvironmental reconstruction as well as proxies for the carbon source and biogeochemical processes as microbial degradation (Meyers 1997; Otto and Simpson 2005; Strauss et al. 2015). This study focuses on the aliphatic n-alkanes, lipids that contain unbranched, long-chain hydrocarbons with mostly odd-carbon numbers. They are an important component of plant leaf-waxes and get preserved in sediments through deposition. The main source of these hydrocarbons in sediments are algae, bacteria and aquatic vascular plants within lakes, and terrestrial vascular plants (Meyers 2003). As they have a low sensibility to microbial degradation, n-alkanes are quite robust proxies for the organic matter source (Cranwell et al. 1987; Eglinton and Eglinton 2008; Zech et al. 2013). For the analysis of the lipid biomarkers, 20 samples were used: five from BAL24-T0-UL and BAL24-T0-DLB, seven from BAL24-T0-L and three from BAL24-T0-M.

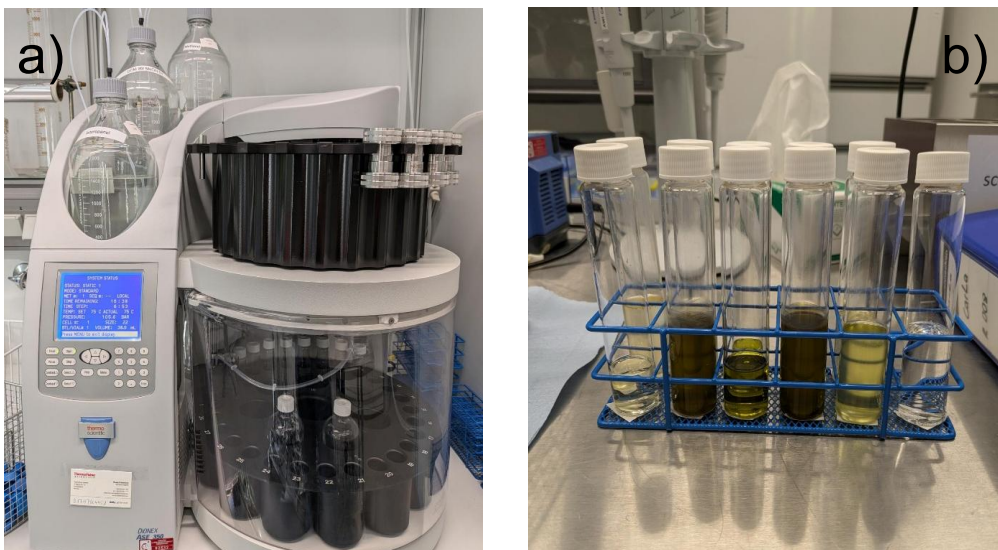


Figure 17. (a) The ASE 350 extractor and (b) the dissolved compounds.

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4.2.5.1 Measurement

For the lipid biomarker analysis, samples were extracted with an accelerated solvent extraction (*ASE 350, Dionex*, Figure 17a), using a mixture of Dichloromethane and Methanol (DCM + 1%MeOH, 99:1) as extraction solvent. For this, approximately 8 g of freeze-dried and grinded sediments were weighted in the extraction cells. The samples were heated over 5 min to 75 °C, followed by a static phase of 20 min. The pressure was constant at 5 MPa. Afterwards the extraction (Figure 17b), the dissolvent compounds were concentrated with a Rocket Synergy vacuum evaporator (*Genevac SP Scientific*) at 42 °C and further dried by evaporation under a stream of nitrogen. For quantification of the compounds, 80 µL each of 4 internal standards with a concentration of 100 µg mL⁻¹ were added to the samples:

- 5α-androstane (C₁₉H₃₂) for aliphates
- ethylpyrene (C₁₈H₁₄) for aromatics
- 5α-androstan-17-on (C₁₉H₃₀O) for nitrogen-, sulfur-, oxygen- (NSO-) containing compounds
- erucic acid (C₂₂H₄₂O₂) for the NSO-fatty fraction

Afterwards, a large excess of n-hexane (C₆H₁₄) was added to remove the n-hexane-insoluble fraction (“Asphaltene precipitation”) and the remaining n-hexane-soluble portion was separated into unipolar (aliphatic and aromatic hydrocarbons) and polar (hetero (NSO-) compounds) fractions using medium-pressure liquid chromatography (*MPLC, Köhnen-Willsch Chromatographie*, (Radke et al. 1980), Figure 18a). Therefore 300 µL of the samples dissolved in 1 ml n-hexane were injected into the MPLC, where the unpolar fractions were eluted with n-hexane, the polar with DCM.

The following measurement of the fractions was carried out with a gas chromatography-mass spectrometry (GC-MS, *Trace 1310 + ISQ 7000, Thermo Fisher Scientific*, Figure 18b).

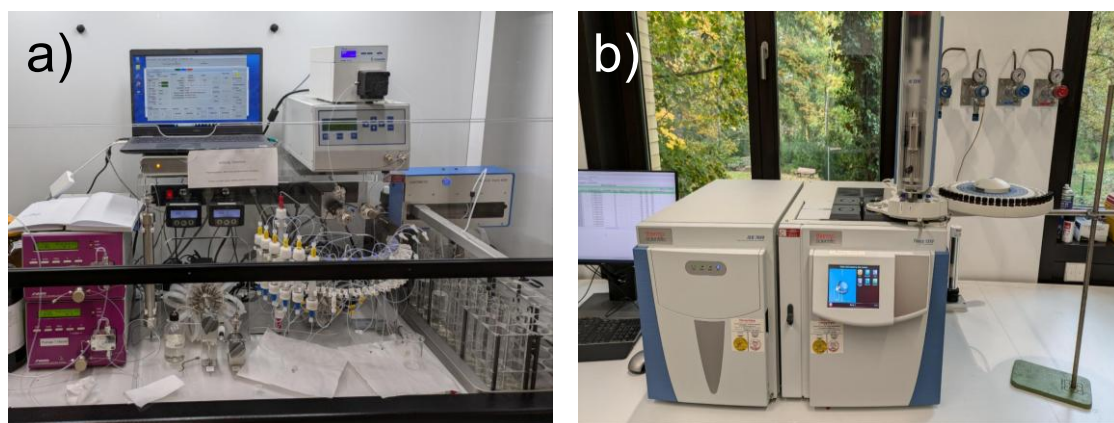


Figure 18. (a) The MPLC and (b) the GC-MS.

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The samples were injected in the GC system and vaporized by heating from 50 °C to 300 °C with a heating rate of 10 °C min⁻¹ and an isothermal holding time of 10 min, and then led through a fused silica capillary column (SLB-5ms Sigma Aldrich, 60 m length, 0.25 mm diameter), using helium as carrier gas (constant flow of 1 mL/min). For the measurement, the GC was heated with a rate of 3 °C min⁻¹ from the initial 50 °C to 310 °C, followed by an isothermal holding time of 30 min. For following compound analysis, the coupled mass spectrometer operated in electron impact ionization mode at 70 eV and 230 °C, recording a mass spectra $m/z-1$ from 50 to 600 u at a scan rate of 2.5 scan s⁻¹. The n-alkanes were quantified in the GC-MS chromatogram using the Xcalibur software (Thermo Fisher Scientific) by comparing the peak area of targeted compounds with the peak area of the internal standard (5 α -androstande).

4.2.5.2 Biomarker Indices

Five biomarker indices were calculated from the n-alkane concentrations. They serve as proxies for (1) the quality (total long-chain alkane concentration and CPI), and (2) the aquatic vs. terrestrial influence and thus the source of the OM (ACL, p_{aq} and p_{wax}).

Total long-chain alkane concentration

The total long-chain alkane concentration indicates the quality of the OM, with higher concentrations standing for a better quality, and was calculated as the sum of the whole range of detected long-chain n-alkanes (C₂₁₋₃₅):

$$\text{Total alkane concentration}_{C_{21-35}} = \sum C_i, \quad (5)$$

where i is the carbon number index and C is concentration.

Carbon preference index

The carbon preference index (CPI) is the ratio of odd- to neighbouring even-numbered alkanes introduced by (Bray and Evans 1961), as a proxy of the degradation and alternation by quantifying the odd over even predominance of the carbon chains, as microbial degradation produces more even chains (Zech et al. 2013). Thus, high values reflect typically fresh, undegraded soils, whereas the lower, the more degraded the soil (Bray and Evans 1961; Marzi et al. 1993). The revised calculation by Marzi et al. (1993) and the interval C₂₃₋₃₃ was used:

$$CPI_{C_{23-33}} = \frac{\sum \text{odd } C_{23-31} + \sum \text{odd } C_{25-33}}{2 * \sum \text{even } C_{24-32}}, \quad (6)$$

where n is the starting dominating chain length/2, m is the ending dominating chain length/2, i is the index of the carbon number, and C is the concentration.

Average chain length

The n-alkane average chain length (ACL) is calculated by the concentration-weighted sum of the different carbon chain lengths, introduced by Poynter and Eglinton (1990). While long-chain n-alkanes are typical for higher plants, with grasses and herbs dominated by 31 and 33 carbon atoms and trees and shrubs by 27 and 29 carbon atoms, short-chain n-alkanes are typical for bacteria and algae (Poynter and Eglinton 1990; Ficken et al. 1998; Zech et al. 2010; Zech et al. 2013), as seen in Figure 19. We used the long-chain interval C_{23-33} :

$$ACL_{C_{23-33}} = \frac{\sum i * C_i}{\sum C_i}, \quad (7)$$

where i is the carbon number index and C is the concentration.

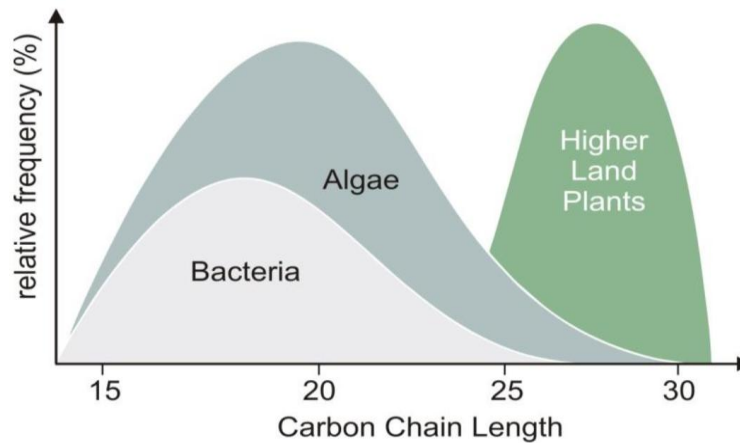


Figure 19. Relative frequency of n-alkane chain-length in bacteria, algae and higher land plants. From Strauss et al. (2015), modified after Killops and Killops (2009).

Aquatic organic matter proxy

The aquatic OM proxy p_{aq} was developed by Ficken et al. (2000) as followed:

$$P_{aq} = \frac{C_{23} + C_{25}}{C_{23} + C_{25} + C_{29} + C_{31}} \quad (8)$$

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and can be used to indicate the aquatic influence of plant inputs by the relative proportion of mid-chain length to long-chain length homologues with values for submergent and floating plants > 0.4 , emergent plants $0.1 - 0.4$ and terrestrial plants < 0.1 (Ficken et al. 2000).

Waxy organic matter proxy

The waxy OM proxy p_{wax} , derived by Zheng et al. (2007), reflects the relative proportion of waxy hydrocarbons derived from emergent macrophytes and terrestrial plants to total hydrocarbons and is additionally used, as the p_{aq} was derived for tropical and temporal regions (Jongejans et al. 2020):

$$P_{wax} = \frac{C_{27} + C_{29} + C_{31}}{C_{23} + C_{25} + C_{27} + C_{29} + C_{31}}, \quad (9)$$

with values for submerged or floating macrophytes < 0.5 , for emergent macrophytes between 0.6 and 0.7, and for terrestrial plants > 0.75 (Jongejans et al. 2020).

4.3 Statistical analysis

To evaluate the obtained data, the results were statistically analysed. All analysis and visualisations of the results were carried out with the free software *R v 2023.12.1.402* (Posit team 2024). To receive a first overview over the data, descriptive statistics as the minimum, maximum, mean and median was used.

As the vibracoring led to a soil compaction of 144 cm for BAL24-T0-L, especially in the upper layers, a normalized exponential depth correction after the classic compaction model of Athy (1930) with the correction method of Lisiecki and Herbert (2007) was applied for this core. For this, it was assumed that the porosity decreases, and therefore the compression increases exponentially with depth, as the top is drilled the longest and has softer soil. This is resulting in the correction formula:

$$z_t = D_p * \frac{1 - e^{-z_m/\lambda}}{1 - e^{-L_c/\lambda}}, \quad (10)$$

where z_t is the true depth, z_m any measured depth within the core liner, D_p the penetration depth of the barrels in the sediment, L_c the recovered core length and λ the exponential e-folding scale.

A λ of 100 cm was used, based on previous thermokarst und Yedoma studies regarding the water content, grain size and porosity (Anthony et al. 2014; Strauss et al. 2017). Figure A. 6 shows the resulting core correction curve for BAL-T0-L.

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4.3.1 Correlation

A correlation matrix after Pearson was calculated to obtain an initial overview of the relationships between the parameters with the *corrplot* package (Wei and Simko 2024). Further, the relations of degradation proxies were analysed by scatterplots with linear regression models and the Pearson correlation coefficient with the *ggpubr* package (Kassamara 2023).

4.3.2 Significance testing

The statistical distribution was tested via the Shapiro-Wilk normality test and a quantile-quantile plot. As the distribution was non-normal, the non-parametric Kruskal-Wallis rank sum test was applied to compare the biogeochemical and biomarker parameters between the four different profiles, as well as the non-parametric Mann-Whitney-Wilcoxon test for pairwise comparison, using the packages *ggpubr* (Kassamara 2023) and *ggplot2* (Wickham 2016). The same was carried out between the three permafrost layers AL, PF and T.

4.3.3 Clustering analysis

Finally, a principal component analysis (PCA) was applied on the scaled data, to reduce the data dimension and to indicate underlying patterns in the parameters, especially regarding the questions if these patterns correspond to the cores and permafrost layers. It was performed once with all parameters and once with the biomarker proxies just as supplementary data, not included in the calculation. A PCA serves to transform the variables into a smaller and uncorrelated amount of essential feature, the principal components, which mainly explain the variance in the data, by minimizing the loss of information. This can be used for a more efficient interpretation and visualization of the data (Abdi and Williams 2010; Jolliffe and Cadima 2016; Greenacre et al. 2022). Afterwards, an agglomerative (bottom-up) hierarchical clustering on principal components (HCPC) was made based on the results of the PCA to group the data in clusters of objects that are like each other while the objects in different clusters are quite different from each other. This bottom-up method begins by treating each data point as an individual cluster. Clusters are then successively merged according to their similarity, ultimately resulting in one single cluster encompassing all objects (Bridges 1966; Shetty and Singh 2021; Uchenna and Olusola 2024). The PCA and HCPC were carried out with the R package *FactoMineR* (Lê et al. 2008) and visualized in a biplot of individuals and

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variables with the package *factoextra* (Kassamara and Mundt 2020). To investigate the properties of the resulting clusters, boxplots of the different groups were created and compared with each other.

5. Results

5.1 Core description

The sedimentological and cryolithological description as well as photos of the cores during subsampling in the climate chamber can be seen in Appendix Figure A. 1, Figure A. 2, Figure A. 3 and Figure A. 4.

5.2 Sedimentology

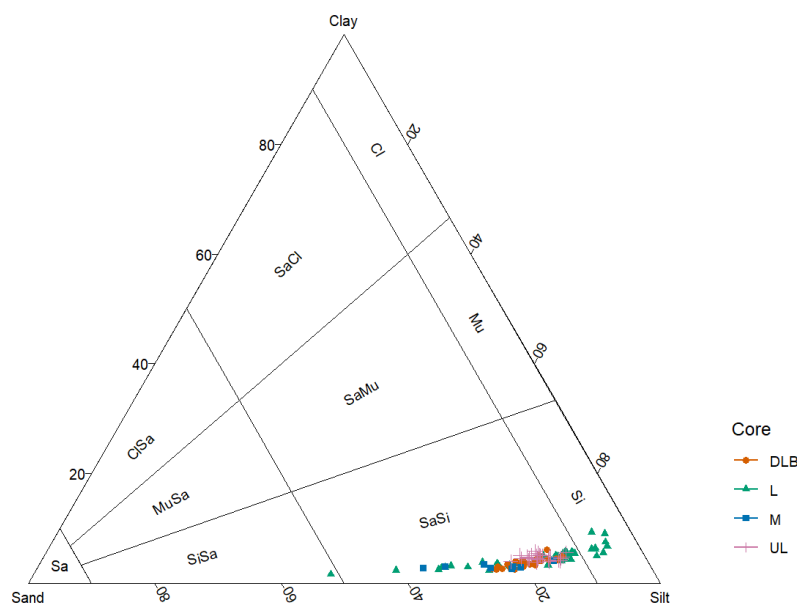


Figure 20. Sand-Silt-Clay diagram after Folk and Ward (1957) for the four different cores.

Figure 20 shows that most of the samples are in the section of sandy silt, also confirmed by the fact that most of the samples consist of coarse silt. Only the lake core has samples in the silt and silty sand fraction.

Upland – BAL24-T0-UL

BAL24-T0-UL has a unimodal grain size distribution and is dominated by silt with a percentage from 74.35 to 82.28 vol%, clay is the lowest fraction with < 6 vol%. The highest volume share is at 122 cm b.s.l. and 40.1 μm , but Figure 21 shows a relatively constant grain size with depth. All samples are classified as poorly sorted coarse silt. The mean grain size is similar for all depths and ranges from 18.22 (204.5cm b.s.l.) to 26.39 μm (112 cm b.s.l.), as seen in Figure 22. Bulked biogeochemical and hydrochemical parameters for the four cores against depth. The background colours indicate the active layer (yellow), perennial

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frozen ground (brown), the talik (grey) and the ice wedge (blue), according to the cryolithologically observation due the subsampling. Red arrows for the radiocarbon dating indicate infinite ages. The absolute ice content varies very strong, especially between 27 and 128 cm b.s.l. It ranges from 99.92 wt% at 128 cm b.s.l. down to 26.96 wt% at 39.5 cm b.s.l. with a mean of 56.63 wt%.

Thermokarst Lake – BAL24-T0-L – originally unfrozen

BAL24-T0-L consists of poorly sorted coarse silt, with a shift towards medium silt between 150 and 350 cm b.s.l. and very coarse silt at the bottom. The silt fraction is highest with a share of 47.10 – 88.24 vol%, the clay fraction the lowest with 4.6 vol% in average. The distribution is mostly unimodal, with a slight second peak in middle sand. The highest share of 9.6 vol% for 66.9 μm is at 444.7 cm b.s.l. (Figure 21). The mean grain size, as seen in Figure 22, ranges from 10.43 – 61.17 μm , with a mean of 22.92 μm . It is relatively homogeneous in the upper core and increasing from 230 cm b.s.l with a peak of 61.17 μm at 444.7 cm b.s.l. The absolute water content shows strong variations in the upper part with wt% from 37.47 at the top to 67.48 at 155.4 cm b.s.l., while it gets more homogeneous in the lower part, ranging between 22.3 and 35.45 vol% at the bottom.

Drained Lake Basin – BAL24-T0-DLB

The sediments of BAL24-T0-M are mainly classified as poorly sorted coarse silt with some samples consisting of very coarse silt at the bottom. Silt ranges between 72.01 – 82.00 vol%, while the clay content is not higher than 6 vol%. Figure 21 shows a maximum volume share with 6.3 vol% and a grain size of 40.1 μm at 144.5 cm b.s.l. but the unimodal distribution is quite constant over depth.

The mean grain size is on average 26.19 μm and shows a slightly increasing trend with depth, ranging from 18.79 to 32.36 μm . The absolute water / ice content is highest at top with 72.31 wt% and rapidly decreasing to 38.31 wt% at 25 cm b.s.l. after which it stabilized between 27.59 and 37.63 wt% (Figure 22).

Marine core – BAL24-T0-M – originally unfrozen

The marine samples are classified as poorly sorted coarse to very coarse silt, dominated by silt with a share between 61.18 and 81.16 % and a clay content smaller than 4 vol%. The distribution is unimodal and quite constant over depth with a peak at 30.5 cm b.s.l. and a share of 7.05 vol% for the grain size 40.1 μm . The mean grain size ranges from 22.51 to

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39.61 μm with a slightly increase with depth. The absolute water content decreases from 47.55 wt% at the top down to 24.11 wt% at 49.5 cm b.s.l.

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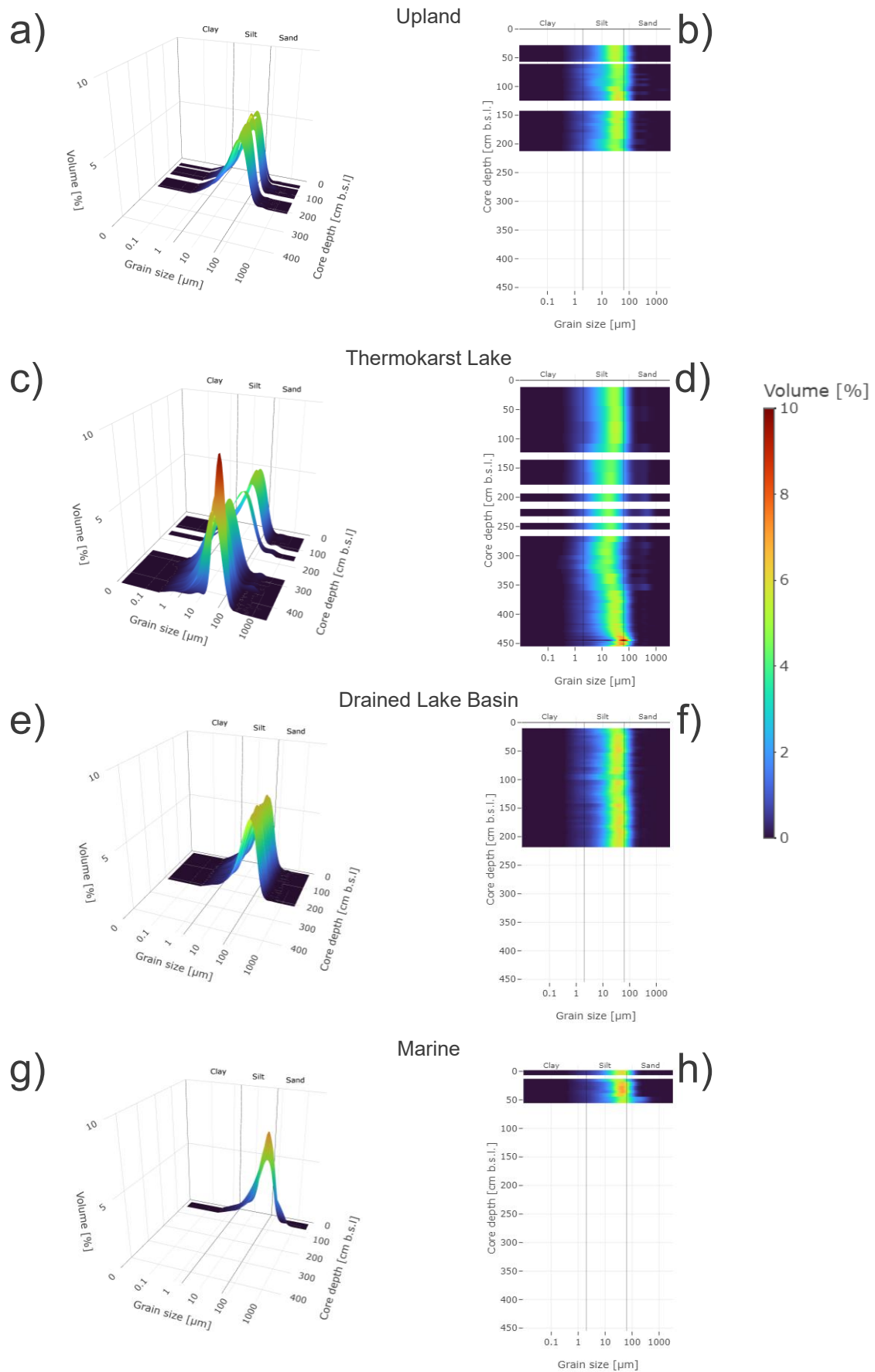


Figure 21. Grain size distribution of the four cores against profile depth with a view from (a, c, e, g) the side and (b, d, f, h) above.

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5.3 Geochronology

Table 2. Calibrated radiocarbon ages.

Core	Mean Depth [cm b.s.l.]	¹⁴ C ages mean [cal. ka BP]	¹⁴ C ages min. [cal. ka BP]	¹⁴ C ages max. [cal. ka BP]	Range [ka]
UL	21.0	0.086	0.032	0.14	0.108
	62.0	30.2915	30.175	30.408	0.233
	101.5	34.385	34.198	34.572	0.374
	146.0	34.4765	34.29	34.663	0.373
	199.0	32.7775	32.269	33.286	1.017
L	6.0	1.7815	1.721	1.842	0.121
	35.5	0.2975	0.287	0.308	0.021
	97.0	0.3965	0.36	0.433	0.073
	138.5	0.4805	0.458	0.503	0.045
	199.5	2.27	2.24	2.3	0.060
	242.5	11.9155	11.829	12.002	0.173
	309.0	1.5335	1.517	1.55	0.033
DLB	12.0	1.1155	1.054	1.177	0.123
	57.0	> 50.000			
	107.0	> 50.000			
	153.0	> 50.000			
	211.5	> 38.086			
M	11.5	0.8	0.684	0.916	0.232
	53.5	28.221	27.678	28.764	1.086

Note. Ages >50
are infinite.

All four cores show a trend with a young sediment layer on top <2 cal. ka BP. While BAL24-T0-L has young sediments in all depth ranging between 0.3 and 2 cal. ka BP, except a peak at 242.5 cm b.s.l. with 12 cal. ka BP, the other three cores have older sediments below the surface layer. There the calibrated ages of BAL24-T0-UL and BAL24-T0-M are ~30 cal. ka BP, while BAL24-T0-DLB has the oldest sediments with infinite ages > 50 cal. ka BP.

5.4 Bulked biochemical parameters

A visualization of the bulked parameters for all for cores is shown in Figure 22. The corresponding statistical parameters can be seen in Table 3, Table 4, Table 5 and Table 6.

Upland – BAL24-T0-UL

BAL24-T0-UL has the highest values for TOC, TN and C/N in the upper 40 cm b.s.l. with maximum values up to 44.42 wt%, 2.01 wt% and 42.02 respectively. Underneath, the values stay relatively constant around 5 wt%, 0.4 wt% and 7 with a slightly increasing trend with depth and a small peak at 59 cm b.s.l. $\delta^{13}\text{C}$ rises strongly in the upper 40 cm b.s.l. from around -30 to -27 ‰ vs. VPDB, where after it slowly approaches values around -26‰ vs. VPDB in the depth.

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The values of the water isotopes $\delta^{18}\text{O}$ and δD are decreasing in the upper half from -15 and -112 ‰ vs. SMOW, except a jump above the ice wedge, followed by a strong decrease in the ice wedge layers after which the values stay around -20 and -150 ‰ vs. SMOW. The DOC value varies between 5 and 250 mg L⁻¹ in the upper part till the end of the ice wedge, where after it rises to the maximum of 1707 mg L⁻¹. The same is true for the EC with an increase from 43 to 2379 $\mu\text{S cm}^{-1}$ below the ice wedge. The pH-value is the lowest in the top, increasing with depth to its maximum in the ice wedge and decreasing again afterwards.

Table 3. Biogeochemical and hydrochemical parameters for BAL24-T0-UL.

BAL24-T0-UL	Parameter	n.valid	min	med	max	mean	sd
Biogeochemistry	TOC [wt%]	39	1.85	4.75	44.42	8.42	10.88
	TN [wt%]	39	0.11	0.39	2.01	0.50	0.40
	C/N [-]	39	9.07	12.26	42.02	14.28	6.23
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	32	-29.76	-26.27	-25.81	-26.58	0.85
Hydrochemistry	$\delta^{18}\text{O}$ [‰ vs. SMOW]	14	-20.36	-18.14	-14.63	-17.83	2.10
	δD [‰ vs. SMOW]	14	-160.21	-139.43	-112.28	-137.89	17.11
	mean d excess	14	-3.06	4.61	10.76	4.74	3.85
	DOC [mg L ⁻¹]	15	5.78	159.10	1707.00	457.70	566.58
	EC [$\mu\text{S cm}^{-1}$]	15	43.41	205.90	2379.00	679.89	831.28
	pH [-]	15	4.87	6.74	7.43	6.58	0.69

Thermokarst Lake – BAL24-T0-L

TOC in BAL24-T0-L has strong variations in the upper half with a range between 4.4 and 24.4 wt%, after which it decreases continuous with depth to 0.6 wt%. TN and C/N show the same pattern with values between 0.11 to 1.28 wt% and 9.26 to 21.08, respectively, with 6 samples below the detection limit. $\delta^{13}\text{C}$ varies over depth with a minimum of -28.61 ‰ vs. VPDB at 318.5 cm b.s.l. $\delta^{18}\text{O}$ and δD are highest at the top and the bottom with values around -17 and -134 ‰ vs. SMOW, respectively and are continuous decreasing towards the bottom part with a final increase at the bottom. DOC, EC and pH stay constant around 155 mg L⁻¹, 623 $\mu\text{S cm}^{-1}$ and 7.8 over the profile.

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Table 4. Biogeochemical and hydrochemical parameters for BAL24-T0-L.

BAL24-T0-L	Parameter	n.valid	min	med	max	mean	sd
Biogeochemistry	TOC [wt%]	61	0.60	2.79	24.40	6.30	6.57
	TN [wt%]	61	0.01	0.21	1.28	0.39	0.36
	C/N [-]	55	9.26	13.61	21.08	14.08	2.85
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	55	-28.61	-26.80	-24.74	-26.78	0.73
Hydrochemistry	$\delta^{18}\text{O}$ [‰ vs. SMOW]	17	-18.95	-18.56	-17.03	-18.30	0.62
	δD [‰ vs. SMOW]	17	-152.39	-147.70	-133.24	-146.07	6.01
	mean d excess	17	-1.17	-0.11	6.95	0.29	1.85
	DOC [mg L ⁻¹]	22	113.80	151.55	213.50	155.35	29.63
	EC [$\mu\text{S cm}^{-1}$]	22	91.00	626.40	862.60	623.25	176.29
	pH [-]	22	7.22	7.74	8.22	7.75	0.23

Drained Lake Basin – BAL24-T0-DLB

TOC and TN drop in the upper 15 cm b.s.l. from the maximum of 18.4 and 1.6 wt% and remain at values below 8.1 and 0.1 wt%, whereby two samples are below the detection limit. C/N stays below 15.9 with small variations. $\delta^{13}\text{C}$ ranges between -26.3 and -24.5 ‰ vs. VDPB with strong fluctuations over depth. $\delta^{18}\text{O}$ and δD have their maximum at the top with -17.7 and -112.2 ‰ vs. SMOW with a strong decrease afterwards up to -19.6 and -146.4 ‰ vs. SMOW. DOC, EC and pH have small changes over the profile around 189.2 mg L⁻¹, 909.4 $\mu\text{S cm}^{-1}$ and 8.

Table 5. Biogeochemical and hydrochemical parameters for BAL24-T0-DLB.

BAL24-T0-DLB	Parameter	n.valid	min	med	max	mean	sd
Biogeochemistry	TOC [wt%]	43	1.19	2.76	18.38	3.12	2.69
	TN [wt%]	43	0.01	0.22	1.57	0.24	0.23
	C/N [-]	41	10.19	12.25	15.88	12.50	1.29
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	37	-26.34	-25.31	-24.45	-25.37	0.51
Hydrochemistry	$\delta^{18}\text{O}$ [‰ vs. SMOW]	12	-19.57	-18.71	-14.25	-17.70	2.02
	δD [‰ vs. SMOW]	12	-146.35	-142.17	-112.15	-135.20	12.82
	mean d excess	12	1.30	6.46	14.60	6.40	4.13
	DOC [mg L ⁻¹]	17	46.63	211.25	312.25	189.22	71.11
	EC [$\mu\text{S cm}^{-1}$]	17	733.90	923.70	1071.00	909.42	103.35
	pH [-]	17	7.62	7.97	8.41	7.96	0.18

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Marine – BAL24-T0-M

The values for TOC and TN are slightly decreasing with depth from 3 and 0.2 wt% at the top to 1.5 and 0.1 wt% at the bottom, with 5 samples in the bottom under the detection limit for TN. C/N varies between 11.9 and 19.7 over the profile. $\delta^{13}\text{C}$ is increasing in the depth, with a small decrease at the bottom around -26.7 ‰ vs. VDPB. $\delta^{18}\text{O}$, δD , DOC and pH have values around -13.3‰ vs. SMOW, -102.1‰ vs. SMOW, 0.1 mg L⁻¹ and 7.7, while EC is increasing with depth up to 19020 $\mu\text{S cm}^{-1}$.

Table 6. Biogeochemical and hydrochemical parameters for BAL24-T0-M.

BAL24-T0-M	Parameter	n.valid	min	med	max	mean	sd
Biogeochemistry	TOC [wt%]	12	1.14	1.95	3.03	1.95	0.64
	TN [wt%]	12	0.01	0.12	0.21	0.09	0.08
	C/N [-]	7	11.85	14.48	19.74	14.88	2.52
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	9	-26.63	-26.32	-25.88	-26.27	0.28
Hydrochemistry	$\delta^{18}\text{O}$ [‰ vs. SMOW]	3	-13.42	-13.28	-13.14	-13.28	0.14
	δD [‰ vs. SMOW]	3	-102.51	-102.25	-101.57	-102.11	0.48
	mean d excess	3	3.57	3.74	5.09	4.13	0.84
	DOC [mg L ⁻¹]	12	0.01	0.12	0.21	0.09	0.08
	EC [$\mu\text{S cm}^{-1}$]	3	17290.00	19380.00	20390.00	19020.00	1581.04
	pH [-]	3	7.47	7.83	7.99	7.76	0.27

5 Results

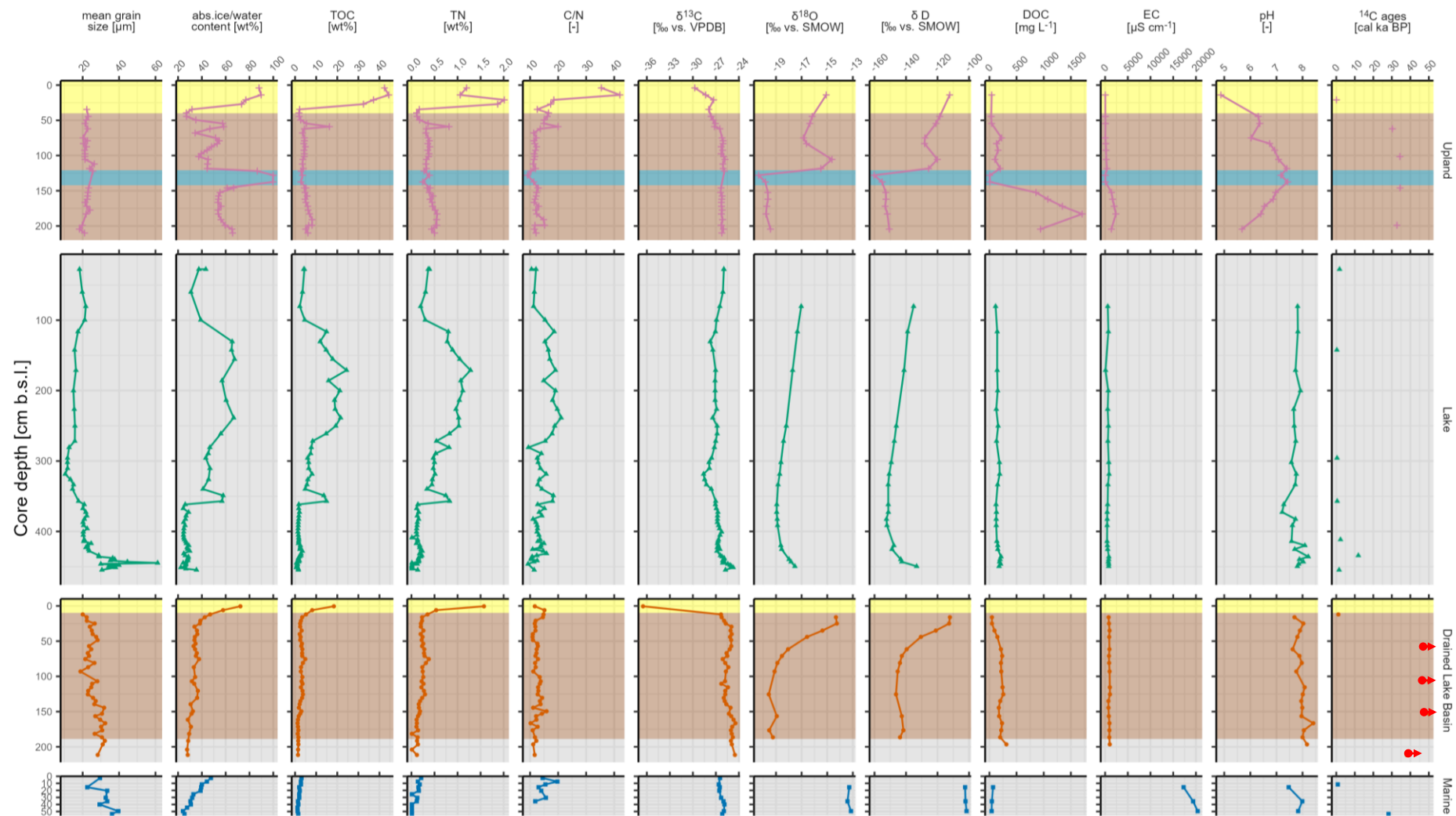


Figure 22. Bulked biogeochemical and hydrochemical parameters for the four cores against depth. The background colours indicate the active layer (yellow), perennial frozen ground (brown), the talik (grey) and the ice wedge (blue), according to the cryolithologically observation due the subsampling. Red arrows for the radiocarbon dating indicate infinite ages.

5.5 Biomarker

The visualization of the biomarker proxies for all cores is displayed in Figure 23.

Upland – BAL24-T0-UL

The total long chain alkane concentration is $1115.2 \mu\text{g g}^{-1}$ Sed and $3439.1 \mu\text{g g}^{-1}$ TOC in the top sample where after it drops to values below $12 \mu\text{g g}^{-1}$ Sed and $600 \mu\text{g g}^{-1}$ TOC. The same trend is true for CPI with 76.9 on the top and values below 13.6 beneath. P_{aq} is increasing in the upper part from 0.02 to values around 0.5, p_{wax} shows the opposite trend, as the ACL with values greater than 26.

Thermokarst Lake – BAL24-T0-L

The total long chain alkane concentration for the Thermokarst Lake varies strongly with depth without a trend between 1.6 and $160 \mu\text{g g}^{-1}$ Sed and 94.8 and $896.8 \mu\text{g g}^{-1}$ TOC. The CPI shows a similar pattern ranging around 18. P_{aq} is first decreasing and then increasing in depth between 0.05 and 0.56, vice versa for p_{wax} and ACL with values higher than 26.7.

Drained Lake Basin BAL24-T0-DLB

The concentration of the long chain alkanes of BAL24-T0-DLB varies slightly from 0.4 and 30.5 to $11 \mu\text{g g}^{-1}$ Sed and $201.6 \mu\text{g g}^{-1}$ TOC, with highest concentration at the top. P_{aq} is increasing with depth and slightly decreases in the bottom, opposite for p_{wax} . The CPI decreases with depth from 14.8 to 4.0, while the ACL stays around 27.6.

Marine – BAL24-T0-M

The marine core has a long chain alkane concentration between 3.1 and $8 \mu\text{g g}^{-1}$ Sed, and 146.2 and $412.4 \mu\text{g g}^{-1}$ TOC. The p_{aq} is slightly increasing from the top to the bottom to 0.5, while the CPI varies around 7.1 and the ACL decreases with a mean of 27.6.

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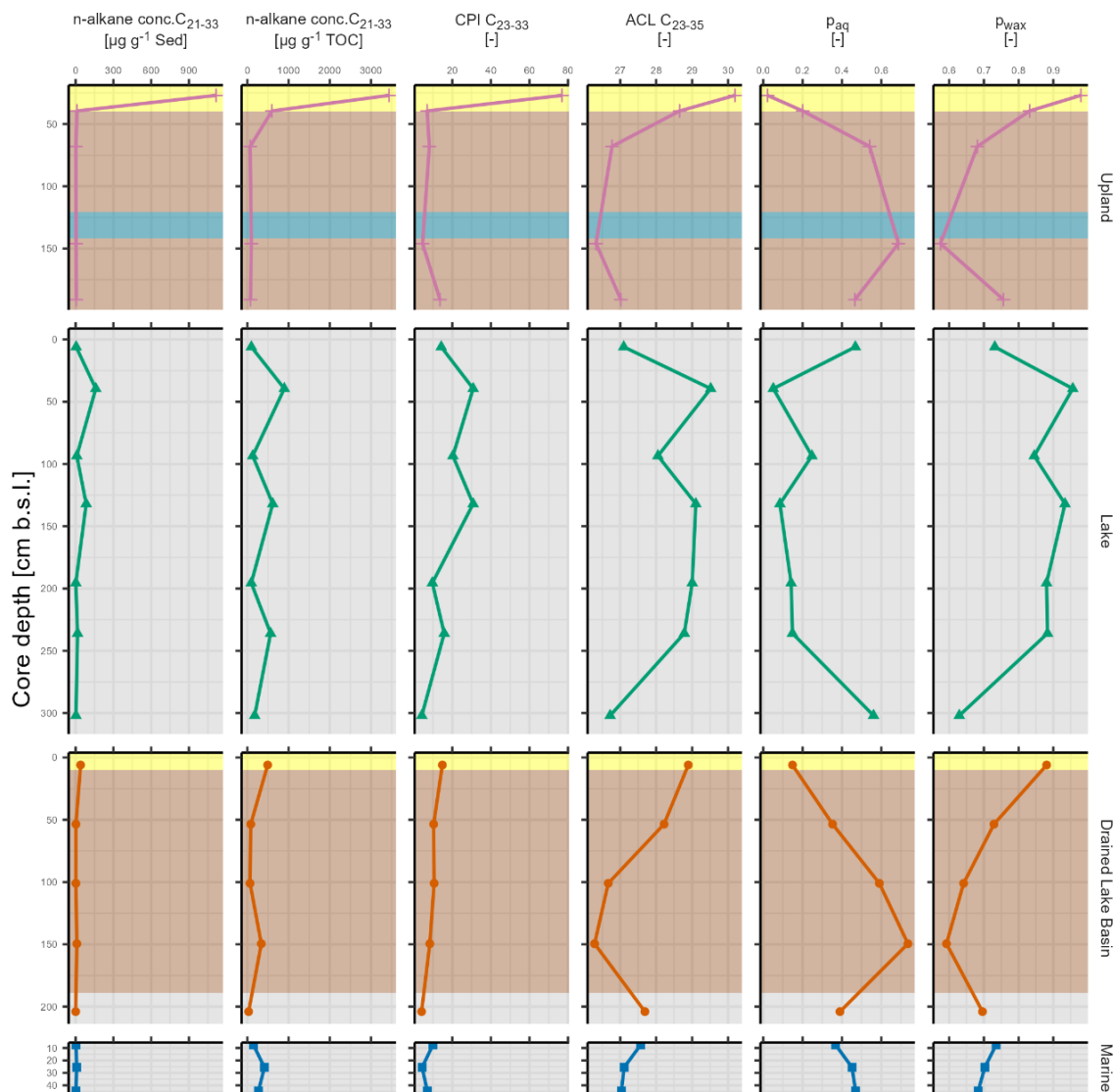


Figure 23. Biomarker parameters of the four cores against depth. The background colours indicate the active layer (yellow), perennial frozen ground (brown), the talik (grey) and the ice wedge (blue), according to the cryolithologically observation due the subsampling.

5.6 Statistical analysis

5.6.1 Correlations

The correlation matrix of all parameters can be seen in the Appendix Figure A. 7. Figure 24 shows the scatterplots of selected OM degradation markers and their correlation among each other. There is a clear correlation between TOC and C/N as well as ACL and C/N ($R = 0.67$ and $R = 0.54$) and a very strong correlation between TOC and CPI ($R = 0.97$). The correlation between C/N and CPI is moderate, whereby the p-value is under the significance level of 0.05 ($R = 0.4$).

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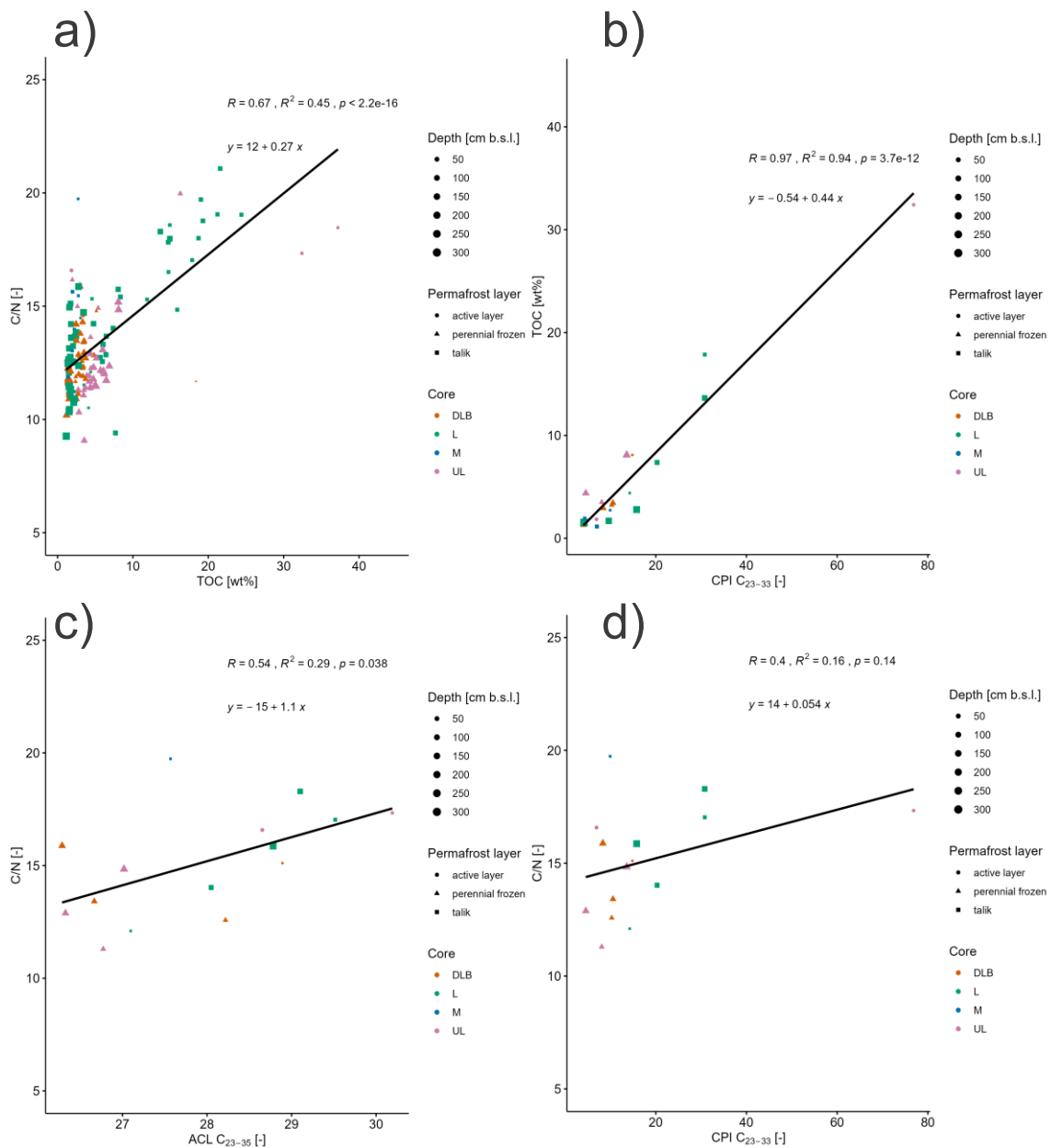


Figure 24. Scatter plots of selected OM degradation proxies. The regression equations, R , R^2 and the p -values are each shown at the top of the plots. The samples of AL are shown as circles, of PF at triangles and of T as squares.

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5.6.2 Significance testing

Difference-testing between the cores

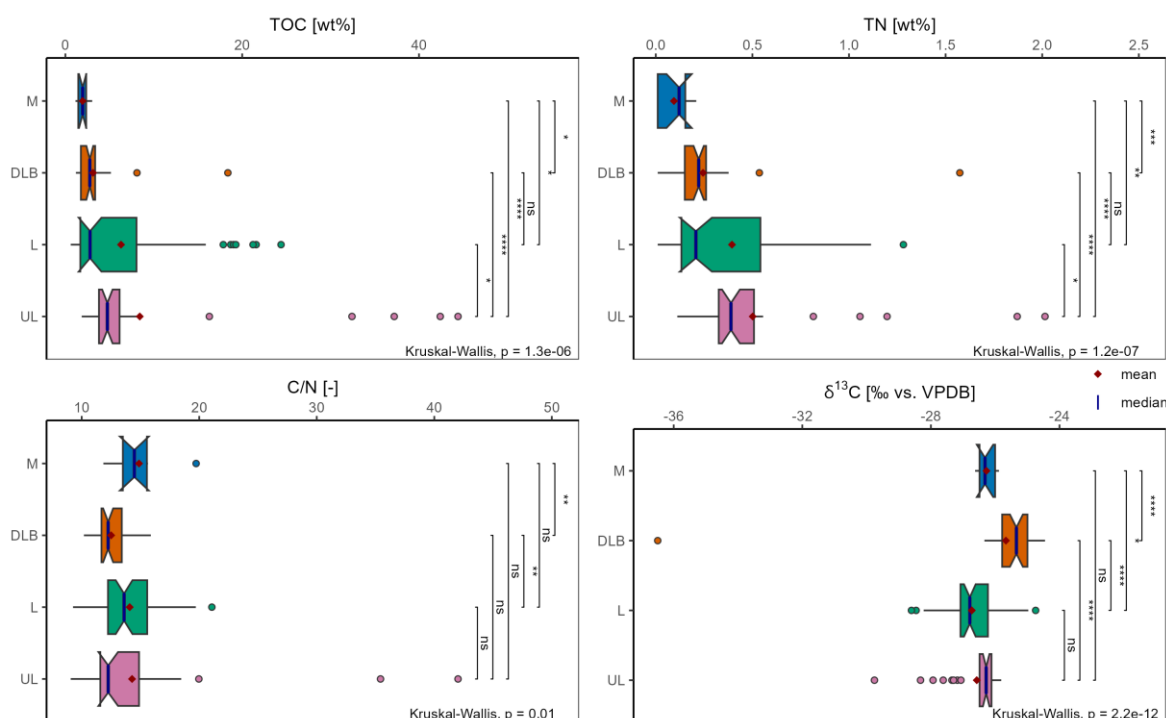


Figure 25. Boxplot of the biogeochemistry for the four sites. The length of the box is defined by the interquartile range (25-75%), the whiskers show the data range with outliers as points. The notches indicate the 95% confidence interval of the median (Overlapping between groups indicates no difference). The median is marked by the blue line, the mean by the red diamond. The statistical differences are shown by the bars on the right (Wilcoxon-test) and in the bottom-right corner (Kruskal-Wallis-test) with ns = not significant, * for $p < 0.05$, ** for $p < 0.01$, *** for $p < 0.001$, **** for $p < 0.0001$.

According to the biochemical parameters, the p-value of the Kruskal-Wallis-Test is under the significance level of 0.05 for all parameters (Figure 25). Therefore, significant differences between the cores exist. TOC is decreasing along the transect from UL to M, supported by the Wilcoxon test showing statistically significant differences between all cores except for L-DLB. The same applies for TN, though here TN is lower in median for L than for DLB. C/N varies more heterogenic across the transect with lower medians for UL and DLB, although UL has the highest values >40 . The differences are statistically significant for DLB-L and DLB-M. $\delta^{13}\text{C}$ is similar for UL, L and M in median (-26.8 to -26.27) with lowest values for UL, but higher in median for DLB. However, the pairwise differences are statistically significant for UL-DLB, L-M and DLB-M. As already seen in Figure 22, the range is the highest for all parameters in UL.

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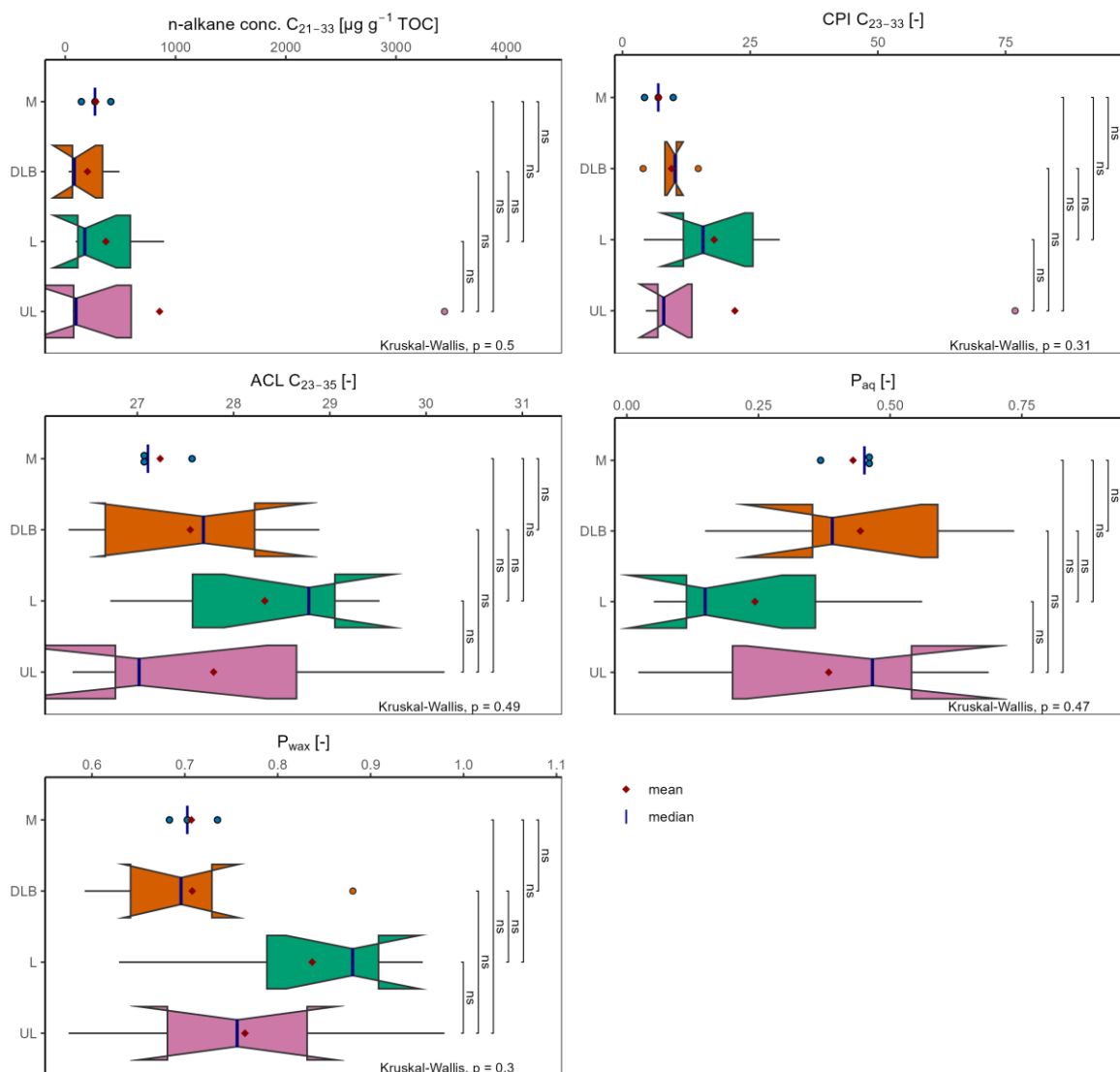


Figure 26. Boxplot of the biomarker proxies for the four sites. The length of the box is defined by the interquartile range (25-75%), the whiskers show the data range with outliers as points. The notches indicate the 95% confidence interval of the median (Overlapping between groups indicates no difference). The median is marked by the blue line, the mean by the red diamond. The statistical differences are shown by the bars on the right (Wilcoxon-test) and in the bottom-right corner (Kruskal-Wallis-test) with ns = not significant, * for $p < 0.05$, ** for $p < 0.01$, *** for $p < 0.001$, **** for $p < 0.0001$.

For the biomarker proxies, Figure 26, the Kruskal-Wallis-test doesn't show a statistically significant differences for the medians as the p-value is higher than the significance level of 0.05 for all parameters, yet the value range varies strongly between the cores. The same is true for the pairwise Wilcoxon-Test with p-values >0.05 . While UL has the highest values for the long-chain alkane concentration, followed by L and DLB, the cores are quite alike in median. This is the same for CPI, ACL, though L is highest in median, followed by DLB. P_{wax} (and vice versa p_{aq}) shows a similar pattern with highest range for UL, but highest median for L.

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Difference-testing between the permafrost layers

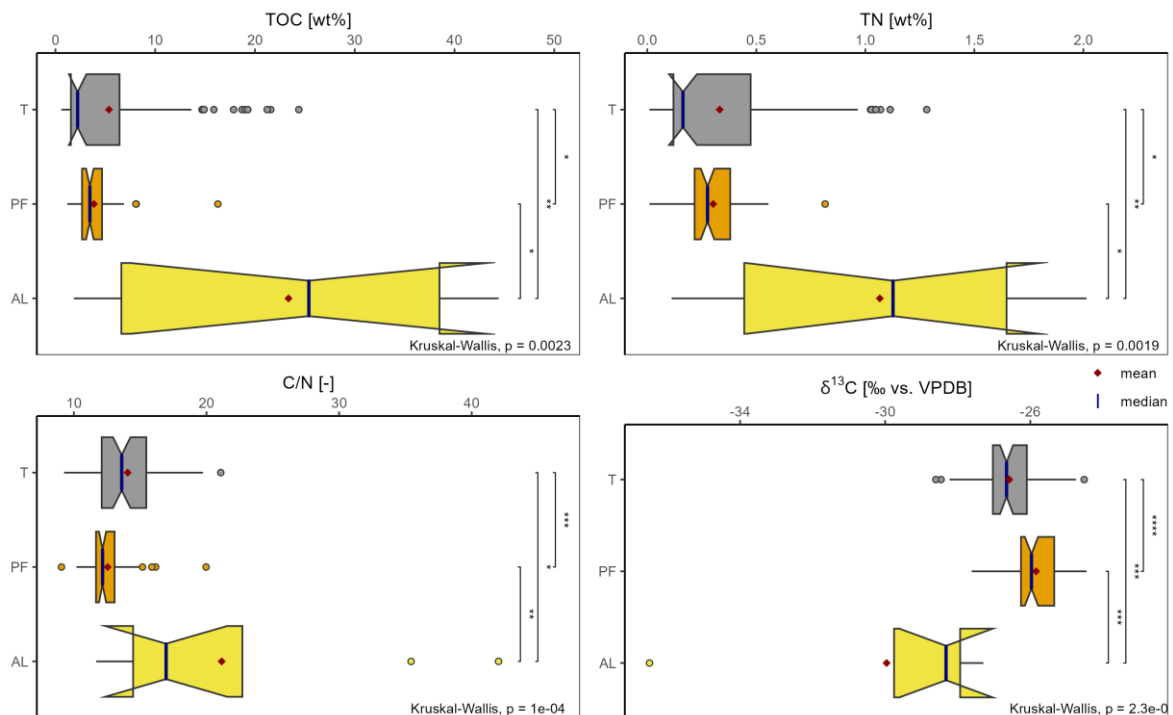


Figure 27. Boxplot biogeochemistry for the different permafrost layers (T = talik, PF = perennial frozen, AL = active layer). The length of the box is defined by the interquartile range (25-75%), the whiskers show the data range with outliers as points. The notches indicate the 95% confidence interval of the median (Overlapping between groups indicates no difference). The median is marked by the blue line, the mean by the red diamond. The statistical differences are shown by the bars on the right (Wilcoxon-test) and in the bottom-right corner (Kruskal-Wallis-test) with ns = not significant, * for $p < 0.05$, ** for $p < 0.01$, *** for $p < 0.001$, **** for $p < 0.0001$.

Regarding the biochemical parameters in Figure 27, TOC, TN and C/N have the highest range and values in median in the active layer (1.85 – 44.42 wt% with a mean of 23.36 wt% for TOC). The perennial frozen samples are lowest in average (3.45 wt% for TOC), the talik samples the lowest in median (2.22, with a mean of 5.37 wt% for TOC). The pattern for $\delta^{13}\text{C}$ is vice versa, with the highest range for the active layer, but the lowest mean and median (-29.96 and -28.32 ‰ vs. VDPB). According to the Kruskal-Wallis and Wilcoxon-Test, the differences for all parameters are statistically significant (As a whole and pairwise between the layers).

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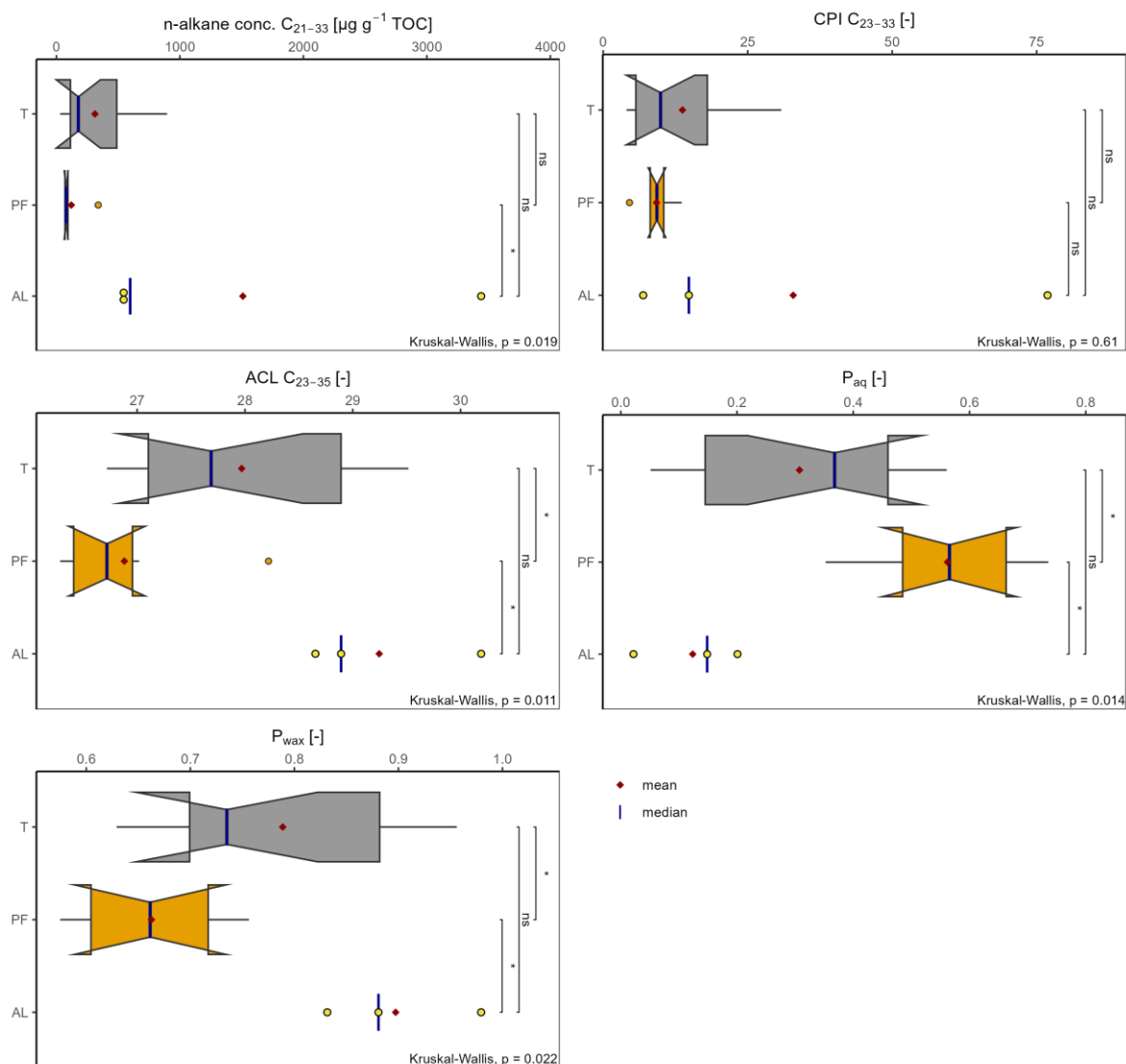


Figure 28. Boxplot of the biomarker proxies for the different permafrost layers (T = talik, PF = perennial frozen, AL = active layer). The length of the box is defined by the interquartile range (25-75%), the whiskers show the data range with outliers as points. The notches indicate the 95% confidence interval of the median (Overlapping between groups indicates no difference). The median is marked by the blue line, the mean by the red diamond. The statistical differences are shown by the bars on the right (Wilcoxon-test) and in the bottom-right corner (Kruskal-Wallis-test) with ns = not significant, * for $p < 0.05$, ** for $p < 0.01$, *** for $p < 0.001$, **** for $p < 0.0001$.

All biomarker proxies (despite p_{aq}) are in median and average the highest for the active layer and the lowest for the perennial frozen samples, as visualised in Figure 28. The range for the total long-chain n-alkane concentration and CPI are the highest in the active layer, for ACL, p_{wax} and p_{aq} in the talik. The p-values of the Kruskal-Wallis-test are <0.05 for all parameters, except for CPI ($p=0.61$), indicating statistically significant differences. The Wilcoxon-Coxney-test reveals a difference between the perennial frozen and the active layer samples. For ACL, p_{wax} and p_{aq} , the differences are statistically significant between the active layer and perennial frozen samples, as well as between the perennial frozen and talik samples, but not between the active layer and the talik samples. For CPI, there are no statistically significant differences.

5.6.3 Principal component analysis and hierarchical clustering

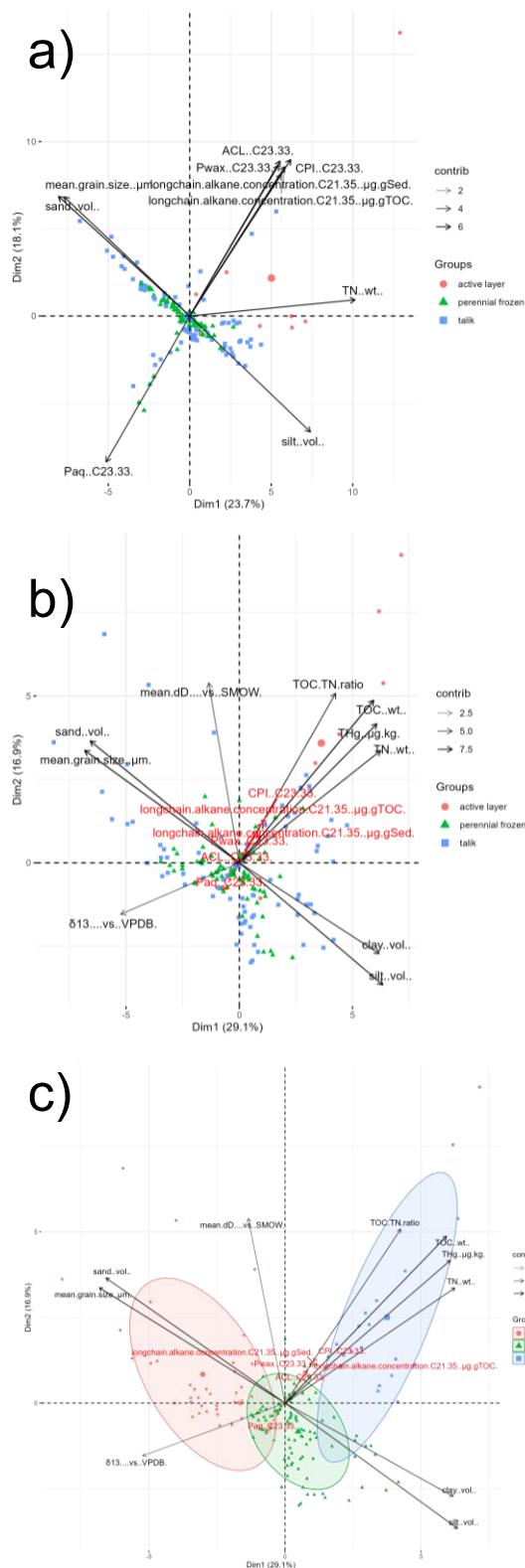


Figure 29. Biplots of the PCA. (a) PCA with all variables, (b) and (c) PCA with biomarker proxies as supplementary variables, while (c) additionally with clusters.

The diagram of the PCA with all variables (Figure 29a) explains 41.8% (23.7% + 18.1%) of the total data set variance. The variables mean grain size, CPI and ALC have the strongest contribution, followed by pwax, sand content and long-chain alkane concentration. For the PCA with die biomarker proxies as supplementary variables (Figure 29b), 46 % (29.1 % + 16.9 %) of the total data set variance is explained by the diagram. Mean grain size, sand content and TOC content have strongest contribution, followed by silt and TN. The biomarker proxies are consistent with TOC und TN content, despite paq. Both diagrams show no clear separation between the different thermal statuses. The PF samples are slightly more centred than the T samples, while the AL samples mostly cluster to the area of Dim1 and Dim2 > 1. The HCPC divides the data set mostly accordingly towards grain size (Cluster 1), the TOC content (Cluster 3) and the centred samples with a smaller grain size (Cluster 2) (Figure 29c). The distribution between the samples after the thermal status doesn't show a clear trend. While the third cluster is mainly dominated by the T samples, the first and second are dominated by the PF and T samples (Figure A. 9).

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Figure 30 shows the characteristics of the resulting cluster. The boxplots confirm the contribution of the variable's variance. Strong differences for the biomarker proxies between the second and the third cluster can be seen, as well as for the grain sizes and the radiocarbon age. Furthermore, is the third group characterized by the higher TOC and TN values.

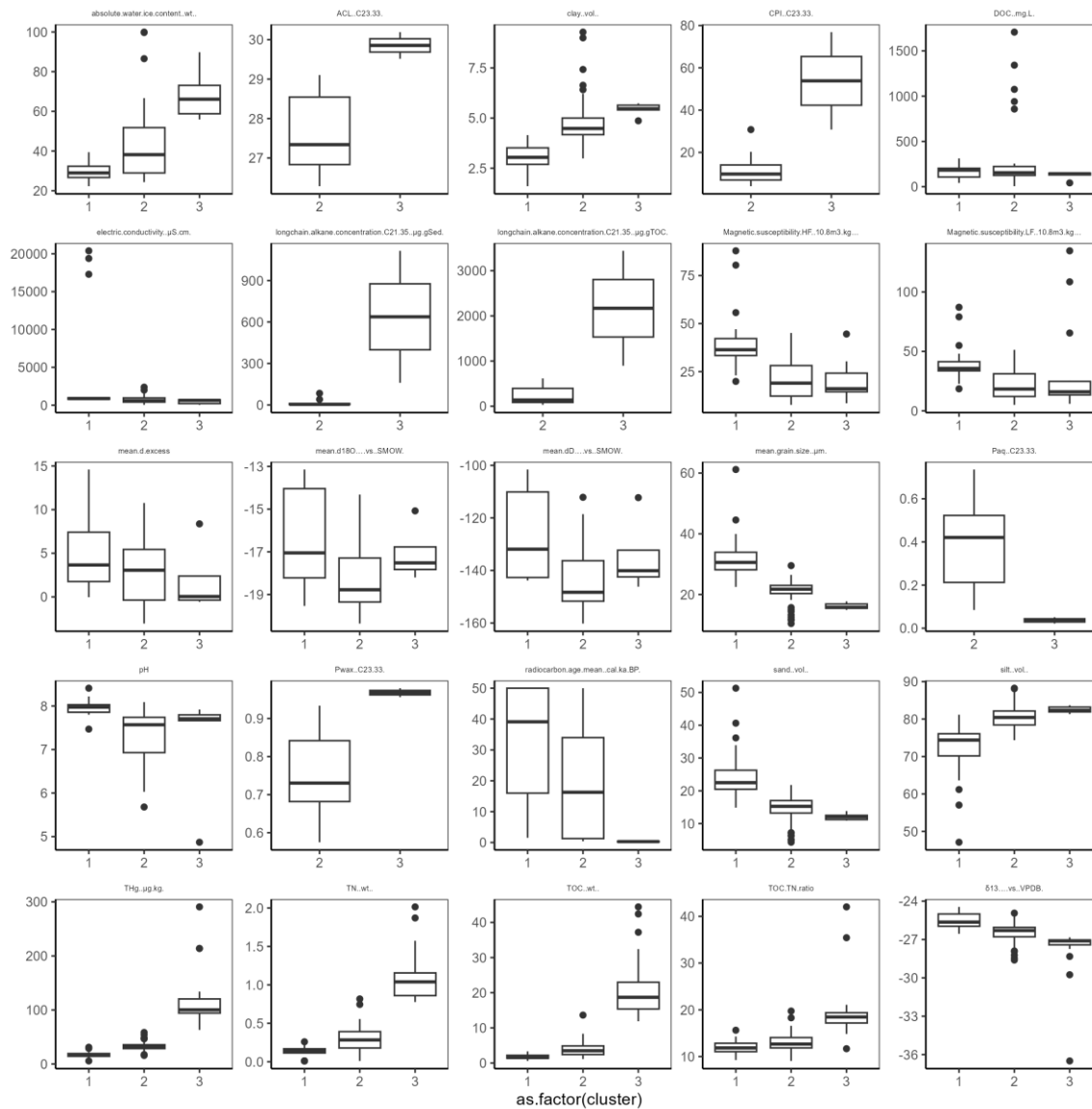


Figure 30. Boxplot of the parameters from the resulting cluster.

6. Discussion

To evaluate the obtained results, this section synthesises the findings regarding the research objectives (1) the reconstruction of the paleoenvironment and (2) the carbon characteristics. The discussion focuses on variations in OM quantity, source and quality and how these relate to permafrost dynamics and degradation processes.

6.1 Landscape history

6.1.1 Deposition mechanism at the study area

The grain size distribution shows a similar signal in all cores, indicating the same material origin at all sites. The sediments are mostly composed of poorly sorted coarse silt with a low clay content below 10 vol% and unimodal distribution, indicating aeolian processes as the dominating deposition mechanism (Strauss et al. 2012; Jongejans et al. 2018). However, the cores show a slightly bimodal distribution with a second peak at medium sand in some depths. This composition is in line with sediments from the west of the Baldwin Peninsula and indicates a slight influence of a second transport mechanism (Jongejans et al. 2018). While the sediments are stronger unimodal than found in Siberian Yedoma deposits, which showed polygenetic deposition processes with alluvial, fluvial and niveo-aeolian transport (Strauss et al. 2012; Strauss et al. 2015; Schirrmeister et al. 2020), similar patterns were found in Alaska (Jongejans et al. 2018; Schirrmeister et al. 2020). The grain size distribution with a slight shift towards finer material points towards loessal silts, which are according to Schirrmeister et al. (2025) primary aeolian deposits of coarse silt and fine sand, which were secondary redeposited. Additionally with the high massive ground ice contents up to 90 % and organic-rich layers, the syngenetic ice wedge, founded macro remains and radiocarbon ages of more than 50 cal. ka PB for DLB and 30 cal. ka BP for UL and the deeper marine samples are typically characteristics of late Pleistocene Yedoma (Schirrmeister et al. 2025).

6.1.2 Landscape evolution at the four study sites

BAL24-T0-UL

The radiocarbon dating of the upland indicates a constant deposition age of ~30 cal. ka BP with a modern top layer. The isotopic porewater composition (Figure 31) of the ice wedge and below shows a Pleistocene signal in the range of other late Pleistocene ice wedges in

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Alaska (CREEL Fox tunnel and the Chatanika River valley near Fairbanks (Lachniet et al. 2012; Schirrmeister et al. 2016) and the Barrow permafrost tunnel in Barrow ((Meyer et al. 2010)) and snow in this region (Meyer et al. unpublished data), implying a syngenetic formation. In contrast, the isotopes above the ice wedge show a Holocene and mixed signal of rain and snow (Meyer et al. 2010). The decrease of the isotopes together with the relatively strongly increasing DOC and EC concentration below the ice wedge indicate former thaw- and freezing cycles from both site due a mitigation of ions towards the freezing front, indicating earlier thermokarst processes (Lenz et al. 2016b). The isotopic composition of the active layer sample is in the same range as active layer data from the Seward Peninsula, indicating a summer rain signal (Lenz et al. 2016b). A slightly peat layer with higher TOC and TN contents is located between 57 and 61 cm b.s.l. Strauss et al. (2012) described similar layers in Yedoma deposits in Northeast Siberia as “paleocryosol”, as patches with plant and wood enrichment. These patches can be caused by more humid and plant growth favouring conditions in the Middle Weichselian 50 – 30 ka BP or in ponding water or boggy environments with increased bioproductivity and low OM decomposition (Schirrmeister et al. 2002; Wetterich et al. 2009; Schirrmeister et al. 2011a; Strauss et al. 2012). This corresponds to the pattern of the stable water isotopes and the DOC and EC concentrations, suggesting former thermokarst processes in the upland (e.g. relict talik), which also explains the slight aquatic influence visible in the biomarker proxies (Section 6.2.2). Grosse et al. (2007) also found fluvial and lacustrine sediments beneath Yedoma in Siberia). Additionally, the lenticular and horizontal structures, as well as ice veins, as seen in Figure A. 1, indicate a degraded permafrost influences by cryoturbation, frost heave and ice wedge growth. This is in line with the OM proxies showing a decrease in C/N and corresponding increase in $\delta^{13}\text{C}$ with depth and general quite low values for CPI and ACL compared to the other sites (Section 6.2.3).

BAL24-T0-L

The thermokarst lake shows a strong change in the sedimentology and biochemistry, with high water, TOC and TN content, finer grain sizes and stronger lamination above 360 cm b.s.l. and lower water, TOC and TN contents with increasing grain size below. This indicates a shifting input, similar like more aquatic signals in the biomarker proxies (Section 6.2.2), with the end of the lake sediments. This paleo active layer also explains the decrease and subsequent increase of the stable water isotopes. The ages with 0.3 – 2 cal. ka BP point towards the young history of the lake in the Holocene, not older than 0.5 ka as the first

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sample above the paleo active layer. Only the sample in 434 cm depth has an age at the border to the Pleistocene with 12 cal. ka BP. As all other depth above and below are much younger, this is more reasonable relocated or reworked material during the lake phase as reworking is unusual in syngenetic sedimentation, but more likely in taliks due volume loss (Jongeans et al. 2018; Jongeans et al. 2020; Farquharson et al. 2022). Along with this is it usual for thermokarst sediments to contain due thaw- and freezing processes radiocarbon ages outside the stratigraphic order (Grosse et al. 2007; Wetterich et al. 2009; Jongeans et al. 2018; Jenrich et al. 2021). Also, the biochemical and biomarker parameters portend to a young lake phase with fast deposition. Their strong variations point to a mixing input during the lake phase. While the partly high C/N, $\delta^{13}\text{C}$ and p_{aq} , as well as the laminations above the paleo active layer indicate an emergent to terrestrial input due shoreline erosion and thus input of mostly top layers with higher and more recent OM content, there are also layers with more aquatic signals with higher p_{aq} and lower C/N and $\delta^{13}\text{C}$ values, resulting from bio productivity in the lake (Anthony et al. 2014; Jongeans et al. 2018). The much coarser material at the bottom is a sign of fluvial influence or ancient thermokarst processes like in the upland, also found in Siberian Yedoma (Grosse et al. 2007; Schirrmeister et al. 2011b; Jongeans et al. 2018; Jenrich et al. 2021) and could represent the glaciomarine and glaciafluvial environment documented in Huston et al. (1990).

BAL24-T0-DLB

The ages of the DLB are with 50 ka BP the oldest, as deep layers could be reached due the subsidence, with a recent Holocene top layer. Even though DTLBs provide favourable conditions for fast peat accumulation (Jones et al. 2022), the modern peat-layer of DLB with high terrestrial OM ($p_{\text{wax}} \sim 9$) is with 10 – 15 cm typical low, as it drained just a few years ago (Section 3.2). Similar to UL, DLB shows signs of thaw- and freeze cycles and permafrost degradation as frost-heave and cryoturbation, especially in the active layer with frost cracking and frost desiccation holes. This is in line with the observations from Jones et al. (2023) that the basin was covered with a microtopography indicating melting ice wedges and degrading polygon centres up to 1 m. The unfrozen part on the bottom is the relict of the sub-aquatic talik during the lake phase, which is not refrozen yet. The massive structure without ice structures in the talik indicate that the whole core column was unfrozen during the lake phase, while the ice veins and lenses indicate refreezing and ice wedge growth after drainage. The shift in the water isotopes from heavier to lighter isotopes confirm a refreezing from top to bottom (Lenz et al. 2016b). The core shows very few signs of the lacustrine phase with

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few laminations (Figure A. 3), low OM content and mostly terrestrial inputs ($p_{\text{wax}} > 0.6$, section 6.2.2), which fits to the findings of Jones et al. (2023) that the lake in 1951 still consisted of two small lakes, which coalesced in 1974. Although a talik of more than 2 m is quite large for this short lake phase, it coincides with fast sub-aquatic talik formation, especially at the beginning of the lake phase, with depths of around 10 m after 50 a modelled by Ling and Zhang (2003) and (Kessler et al. (2012)). Further floating ice conditions of the former lake could have allowed a year-round talik growth. Only the layer between 13 and 27 cm depth implies a short lacustrine phase, with more macro remains and a horizontal and laminated structure (Figure A. 3). As the depth of 12 cm still has an age of >1 cal. ka BP, this is more likely eroded material from the lake margins. While the stable water isotopes in the bottom half point to a snow, and Pleistocene input, the upper half has a mixed signal between rain and snow with heavier, Holocene signals due the ion mitigation towards the freezing front under the active layer (Figure 31) (Meyer et al. 2010; Lenz et al. 2016b).

BAL24-T0-M

The top of the marine layer has with the turbated cryostructure visible signs of wave action (Figure A. 4). The high C/N and low $\delta^{13}\text{C}$ (14.9 and -26.3 ‰ vs. VPDB in average) in all depths show a clear terrestrial input and thus a deposition under terrestrial conditions. P_{wax} is slightly decreasing with depth, but always >0.68 and thus indicates a terrestrial – palustrine environment more in a slump or flat pond rather than marine, confirmed by the brackish signal of $\delta^{18}\text{O}$ and δD with mean values of -18.3 and -146.1 ‰ vs. SMOW, respectively (Figure 31). This suggests the sediments were deposited under terrestrial conditions and afterwards overprinted by the ocean due the marine transgression or shoreline erosion, as the Arctic coast has an erosion rate of 0.5 m/a in average and the core shows lamination between 15 and 22 cm b.s.l. This is also confirmed by the EC of the porewater, indicating water of secondary marine infiltration, not during the deposition. The marine core was taken beneath the erosion gully of the DLB, therefore the eroded sediments, also indicated by the laminations, could originated from the drainage event, as the sedimentological and biochemical values are similar to DLB. The large pebbles at the bottom of the core are therefore more likely resulting from ancient fluvial or thermokarst processes as also found in UL and TKL (Grosse et al. 2007; Schirrmeister et al. 2011b; Jongejans et al. 2018; Jenrich et al. 2021), than “original” marine sediments due to wave action or tidal forces.

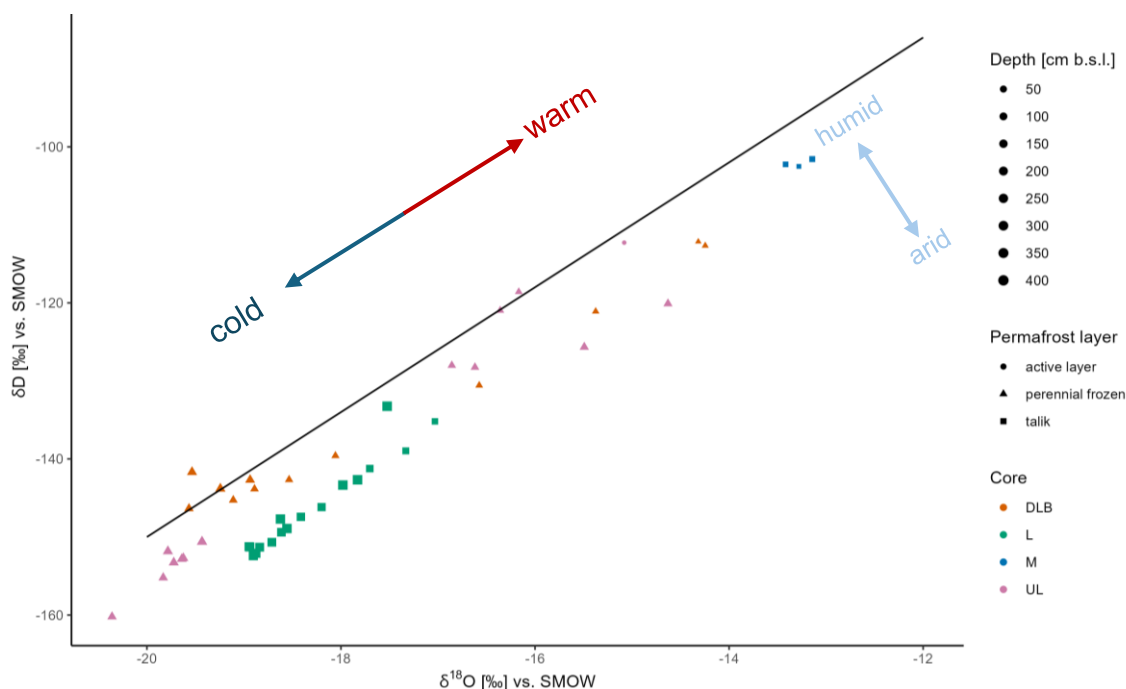


Figure 31. The $\delta^{18}\text{O} - \delta\text{D}$ diagram of the four cores compared with the global mean water line (GMWL) after Craig, 1961.

6.2 Organic carbon

The quantity and quality of the OM strongly influences its future degradability (Knoblauch et al. 2013; Strauss et al. 2013; Stapel et al. 2016; Jongejans et al. 2018). Therefore, it is essential not just to know how much carbon is stored, but also the source and quality of it. While the TOC content provides information about the carbon amount, the source can be indicated by C/N and $\delta^{13}\text{C}$ as well as the distribution of the n-alkanes by the proxies ACL, p_{aq} and p_{wax} , as C/N and $\delta^{13}\text{C}$ can be affected by mineralization and degradation (Meyers 1994). Since ACL in this study is also slightly impacted by degradation (Figure 32), this proxy should also be viewed with caution.

6.2.1 Organic carbon quantity

As seen in Figure 22. Bulk biogeochemical and hydrochemical parameters for the four cores against depth. The background colours indicate the active layer (yellow), perennial frozen ground (brown), the talik (grey) and the ice wedge (blue), according to the cryolithologically observation due the subsampling. Red arrows for the radiocarbon dating indicate infinite ages. In section 5.4, the TOC is highest in the terrestrial upland core, with the highest measurement > 40 wt% in the active layer, and decreasing along the transect, with statisti-

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cally significant differences between the cores, except DLB and L. The upland has the generally highest TOC values in the active layer and thus the top 100 cm due the active input of recent plants, but even in the bottom half the values are still higher compared to the same depth of the other cores, with a mean of 5.1 ± 1.53 below 100 cm. While the upper half is with 12.28 ± 15.23 wt% in the range of the upland from a study from Fuchs et al. (2019) in North Alaska (29.1 ± 9.7 wt%), the lower half is much lower (28.8 ± 2.2 wt%). Considering the whole core of UL, the mean is with 8.42 wt% much higher than in comparable studies in Alaska (1.95 and 4.66 wt% (Strauss et al. 2013; Giest et al. 2025) and Siberia (3.2, 2.4 and 1.9 wt% (Strauss et al. 2013; Jongejans et al. 2018)), resulting from the high TOC content in the active layer. Regarding therefore the median with 4.75 wt%, the TOC content is similar or slightly higher to other Yedoma sites. The mean TOC of the lake core is with 6.35 wt% in the range of other lake sediment studies in Alaska from Lenz et al. (2016a) (1.6, 7.0 and 5.6 wt% in depth 0 – 100cm), (Giest et al. 2025) 5.37 wt%) and (Strauss et al. 2013). The much higher content in the lake sediments (11.32 wt% in upper 352 cm) are likely due the erosion of organic rich active layers in the lake, a higher bioproductivity during the lake phase as well as slow decomposition rates due the cold lake conditions (Strauss et al. 2015; Jongejans et al. 2018) and coincide to the TOC in the upper 30 cm lake sediments from Jongejans et

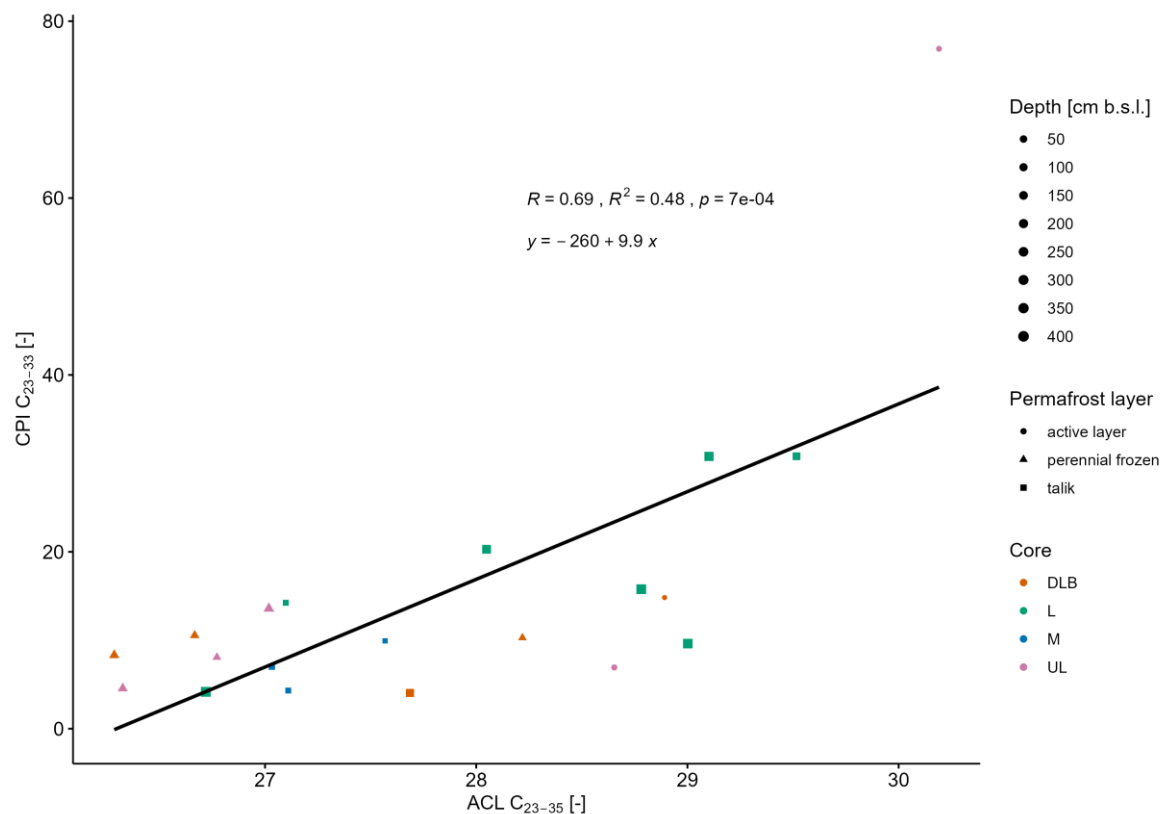


Figure 32. Scatter plot CPI against ACL. The regression equation, R , R^2 and the p -values are shown at the top. The samples of AL are shown as circles, of PF at triangles and of T as squares.

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al. (2018) with a mean of 14.4 wt%. The DLB samples have with 3.12 wt% compared to other studies in North Alaska with 9.75 wt% (Fuchs et al. 2019) and 7.65 wt% (Giest et al. 2025) low TOC contents. The upper 100 cm in particular has significantly lower values with 4.16 ± 3.60 wt% against 11.8 wt% (Giest et al. 2025) and 16 ± 12 wt% (Fuchs et al. 2019). This can be explained by the recent drainage and thus short time for peat accumulation, as Jones et al. (2012) investigated that revegetation starts 5 – 10 years and peat accumulation only 10 – 20 years after drainage. Consequently, the peat layer of our DLB is quite thin with ~10 cm, while Fuchs et al. (2019) measured peat layers of up to 42 cm in old basins (against 12.4 cm in ancient basins) and Giest et al. (2025) of 23 cm in the drained lake basin. The marine core has the lowest TOC concentration with a mean of 1.95 wt% and agrees to the marine sample of Giest et al. (2025) with 1.3 wt%. Furthermore, Jenrich et al. (2021) measured similar concentration in the top 200 cm in a semi-closed lagoon in Northeast Siberia (1.9 wt%). Comparing the different permafrost layers, TOC is in average with 23.35 wt% highest in the active layer with a large range of 1.85 – 44.42 wt%. In contrast, PF shows the lowest mean TOC content (3.85 wt%), while the mean TOC of T is with 5.7 wt% slightly higher. But even if the values for T and PF are much lower than for AL, they are still in the range of other studies for thermokarst and Yedoma sediments (5.10 wt% for taliks in Alaska (Giest et al. 2025), 6.48 wt% for thermokarst deposits, 3.02 wt% for Yedoma deposits (Strauss et al. 2013)).

6.2.2 Organic carbon source

According to Meyers (1994, 1997), our samples are with C/N values from 9.07 to 14.28 between the range of vascular land plants (> 17) and algae (3-9), as seen in Figure 33, while the AL samples from UL are with a C/N > 35 strong terrestrial. General, the samples are less terrestrial the deeper in the soil. This is likely because of degradation then the source (Strauss 13). Our samples fall outside the clear areas of Meyers (1994, 1997), but the same pattern was found in other permafrost studies in Siberia and Alaska (Lenz et al. 2016a; Jongejans et al. 2018; Bischoff 2023; Giest et al. 2025). While the C/N for L and DLB are on median in the range of the ones from Giest et al. (2025) with 13.61 and 12.25 against 14.28 and 13.95, the values for UL and M are much smaller (12.25 vs. 17.57 and 14.48 vs. 25.93). Still the mean values of 14.28, 14.08 and 12.50 for UL, L and DLB are slightly smaller than them from Giest et al. (2025) with 18.45, 14.39 and 17.5. Taking also $\delta^{13}\text{C}$ in account with a mean value of -26.36 ‰ vs. VPDB, all samples are in order of land plants (Meyers 1994, 1997)

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and similar to the upland and thermokarst lake samples from Giest et al. (2025) with mean values of -27.75 vs. -26.58 (UL) and -28.22 vs. -26.78 ‰ vs. VPDB (L), however the DLB samples of this study have smaller values (25.4 vs. 27.52 ‰ vs. VDPB). Figure 33 exemplifies how the DLB samples are more distributed above (and thus more marine) and the L samples below (more lacustrine) the regression line, while the UL, apart from the two outliers, and M samples are clustered in the middle. However, this variations among the cores could indicate less different OM sources and changing influences than a stronger degradation of the OM (Section 6.2.3), as it was discussed before in section 6.1 that the sites have the same sediment origin. Therefore the OM source will be further discussed based on the biomarker indices ACL, p_{aq} and p_{wax} , which are more robust against alteration due degradation (Meyers 1997).

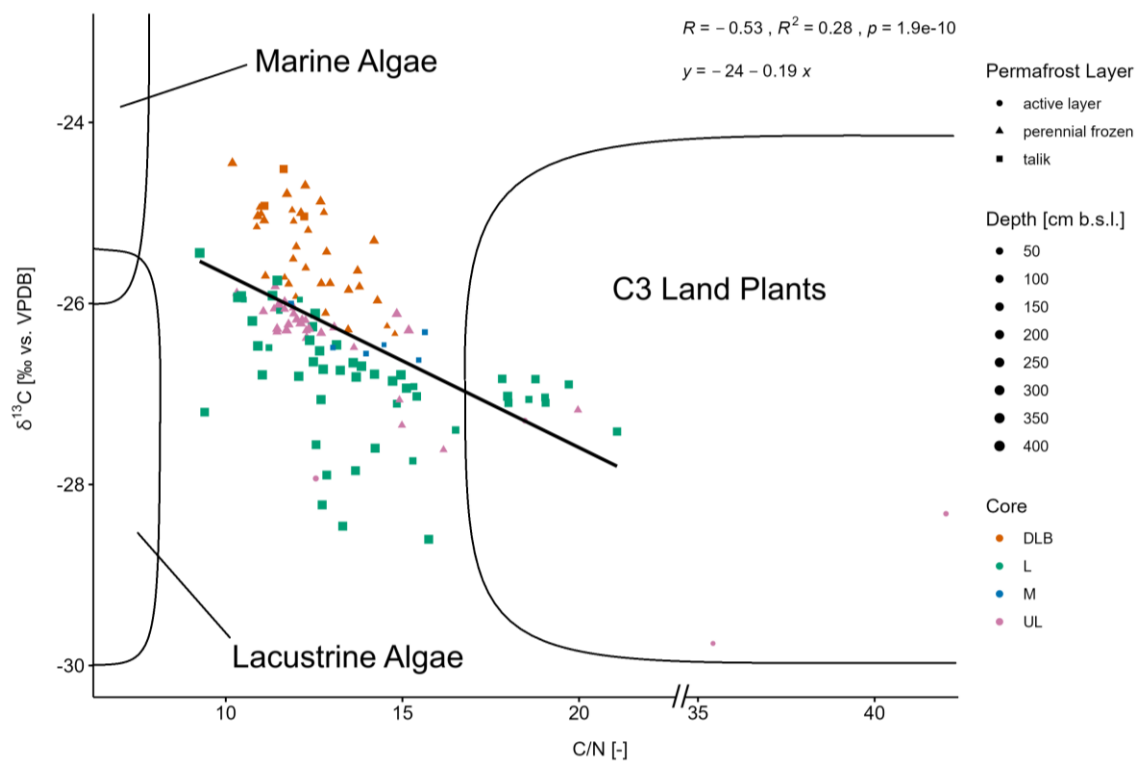


Figure 33. Scatter plot of $\delta^{13}C$ against C/N. The regression equation, R , R^2 and the p -values are shown at the top. The samples of AL are shown as circles, of PF at triangles and of T as squares, the colours represent the cores. Attention for the x-axis break. Regression without AL samples from UL. After Meyer et al. 1997, but identifiers converted with $1.167 * \text{atomic C/N} \approx \text{wt\% C/N}$. DLB sample at 0.5 cm b.s.l. is for clarify reasons not included ($\delta^{13}C = -36.50$ ‰ vs. VPDB and $C/N = 11.68$).

Concerning the biomarker proxies for the OM source, illustrates that the odd-and long-chained n-alkanes are dominating, indicating higher land Figure 34 plants, mostly trees and shrubs (27 and 29) and grasses and herbes (31 and 33) for AL, but also some of the L samples, suggesting a tundra vegetation (Poynter and Eglinton 1990; Ficken et al. 1998; Killops and Killops 2009; Zech et al. 2010; Zech et al. 2013). This is confirmed by the ACL which varies between 26 and 30, with the lowest mean value for M and highest mean value for L.

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The samples of M, UL and DLB below 100 cm have a similar ACL, while it fluctuates for L between 27 and 31, indicating the different OM inputs in the lake and potential shoreline erosion of fresher, more terrestrial soil with ACL in the range of the top layers of UL and DLB. The slightly increase at the bottom of the DLB and UL cores could be a pointer of the changing hydrological conditions, but also multiple thermokarst generations as discussed in section 6.1. As it could also indicate a vegetational succession from grasses to shrubs, this is unlikely here, as it would be much faster than the sedimentation. The further source proxies p_{aq} and p_{wax} confirm this pattern, p_{aq} vice versa to ACL and p_{wax} . The strong correlation between p_{wax} and ACL is shown in Figure 35 and much stronger than between CPI and ACL (Figure 32), indicating ACL here is more a indicator for the OM source than its degeneration. The AL samples as well as most of the deeper L samples are located in the more terrestrial top-right border with high ACL and p_{wax} values, while the deeper UL and DLB and the M

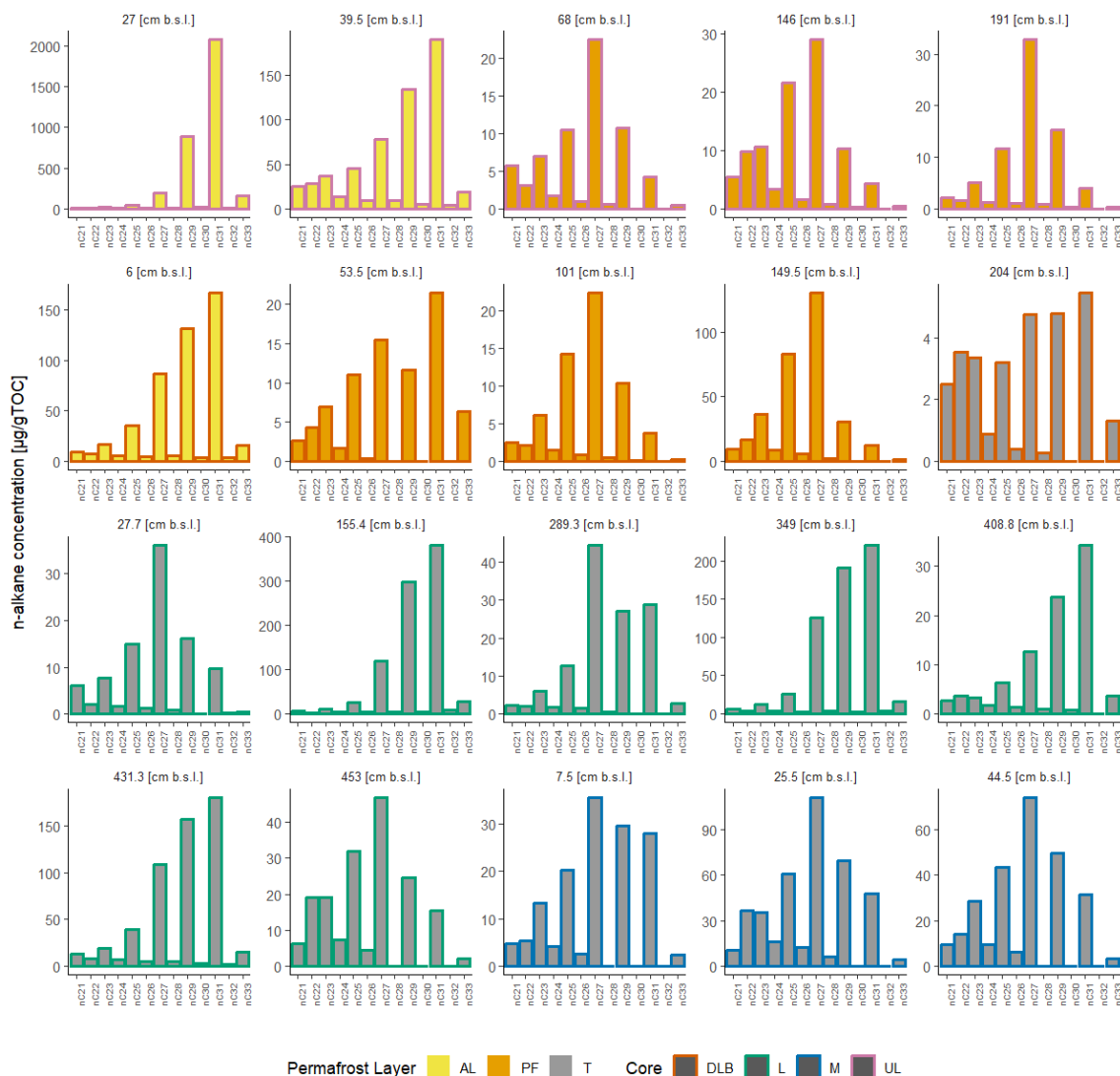


Figure 34. Chain-length distributions of *n*-alkanes. The envelope colour displays the related core, the filling colour the related permafrost layer. Note the x-axis scales are independent from each other for better visualization.

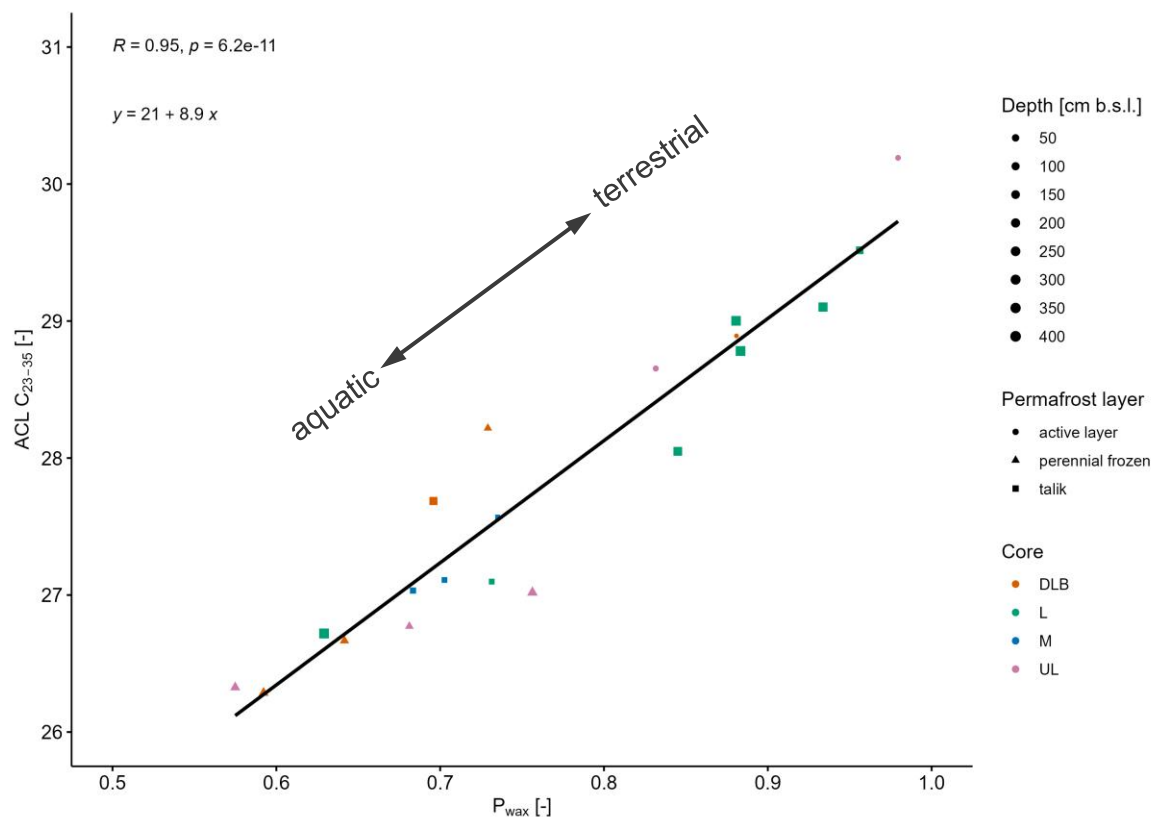


Figure 35. Scatter plot of ACL against p_{wax} . The regression equation, R , R^2 and the p -values are shown at the top. The samples of AL are shown as circles, of PF at triangles and of T as squares, the colours represent the cores.

and the bottom L sample are scattered towards the more aquatic influence with low ACL and p_{wax} values. However, even the values vary between the cores, the differences are not statistically significant for ACL, p_{wax} and p_{aq} , neither between all groups as a whole, nor in pairs between the individual groups. This corresponds to the assumptions from section 6.1 assuming the same parent material for all cores. In contrast, the differences between the three permafrost layers in total as well as paired between PF and both the AL and T are statistically significant (Figure 28). The AL samples have the highest terrestrial input (ACL in average 29.25 and p_{wax} in average 0.9), the ones of PF the lowest (26.88 and 0.66). The samples of T are in between with the highest range (26.72 – 29.52 for ACL and 0.63 – 0.96 for p_{wax}), but even the PF samples are in the range of terrestrial plants, with a slight influence of emergent macrophytes (Meyers 1997).

6.2.3 Organic carbon quality

As discussed in section 6.2.2, C/N and $\delta^{13}C$ are near the range of other studies, but different between the sites. According to the biomarker source proxies, these variances in C/N and $\delta^{13}C$ are more likely because of the OM degradation. OM is selectively degraded, as carbon is more lost as CO_2 while nitrogen tends to remain in the sediment, therefore the C/N alters

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with decomposition, with higher values suggesting better preservation and lower ratios higher degradation (Strauss et al. 2015). $\delta^{13}\text{C}$ becomes less negative with increasing decomposition, as the lighter carbon isotope ^{12}C is favoured for microbial decomposition, leading to an enrichment of the heavier isotope ^{13}C in the OM. Thus, higher $\delta^{13}\text{C}$ values indicate more strongly degraded OM, while lower (more negative) reflects better preserved, less degraded OM (Strauss et al. 2015; Heyer et al. 1976; Skrzypek et al. 2007).

The differences of C/N and $\delta^{13}\text{C}$ along all sites were statistically significant but not paired for UL-DLB and L-M (C/N) and UL-M ($\delta^{13}\text{C}$). While UL, L and M are on average quite alike (14.08 – 14.88) and higher than for DLB with 12.5, UL has a much lower median with 12.26, and thus even lower than DLB (12.25). This is consistent with the variability for $\delta^{13}\text{C}$, where DLB is on median and average higher and statistically significant different from the others. Correspondently to this results, Strauss et al. (2015) also found higher C/N and lower $\delta^{13}\text{C}$ mean and median and thus higher quality for thermokarst deposits than for Yedoma.

Additional, Schirrmeister et al. (2011b) states that low TOC and C/N and high $\delta^{13}\text{C}$ values are common for Yedoma deposits, as they were deposited in glacial or stadial periods with low bioproductivity (Jongejans et al. 2018).

Regarding the biomarker proxies, the CPI is quite similar for all cores between 8 and 16 in median, while again the lake core shores the best quality. However, the active layer sample in the upland has with 3 442 $\mu\text{g gTOC}^{-1}$ and 76.88 an extremely high n-alkane concentration and CPI compared to other studies. Even though it was taken from the peat layer with high organic content, Ficken et al. (1998) found CPI values only up to 20 for peat soils and fresh bog plants. Jongejans et al. (2018) also found n-alkane concentrations in the range of this sample (1 800 and 16 000 $\mu\text{g gTOC}^{-1}$), but with significantly lower CPI between 5 and 15. Thus, it is possible that the sample could not be completely degreased during extraction because of the very high organic content. This is also supported by the fact that the sample was still very greasy after extraction.

Between the permafrost layers, C/N and $\delta^{13}\text{C}$ are statistically significant different, with the highest C/N and lowest $\delta^{13}\text{C}$ in the active layer, where the OM is the most fresh and undegraded, as AL is composed of young OM due high input by recent plants and with contemporary peat accumulation and DTLBs favour fast peat accumulation and C sequestration (Jones et al. 2012). The PF shows the strongest degradation with values of 12.55 for C/N and -25.84 ‰ for $\delta^{13}\text{C}$ in average, indicating old, degraded OM. The higher quality of the talik compared to the perennial frozen samples may be due (1) shoreline erosion and thus input of fresh material and (2) the consistence of young material, as also shown by the young

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¹⁴C ages. This is confirmed by the CPI, which shows the same pattern, even though without significant differences. The CPI for the perennial frozen samples is in median 9.3, with values between 4.56 and 13.59, while it is 14.83 (32.88 ± 38.31 on average) and 9.93 (13.73 on average) for the active layer and talik, respectively. And as the PCA reveals, physical properties as grain size and grain size distribution has the strongest impact on variances in the data, it was expected that the differences wouldn't be that high, as the cores have the same origin and thus sedimentology with no clear separation in the PCAs along the cores and permafrost layers, which is also confirmed by the PCAs, where no really separation along the cores and permafrost layers. The slighter higher quality of thermokarst deposits could be explained due a compensation of the degradation processes by input of fresh OM during the Holocene (Strauss et al. 2015). But even though the OM quality proxies differ among the permafrost layers, they are all in median and average higher than 9 in the range of high quality OM (Brassell et al. 1978; Strauss et al. 2015). However, despite that, it should not be neglected that the soil, especially those of the L and M, have due the currently still undecomposed OM a high GHG release potential under further climate warming due the talik formation, especially as they thereby reach the deep, undegraded layers of the permafrost (Strauss et al. 2015).

7. Conclusion

Four this study, four cores from the primary permafrost units along a degradation transect were investigated to draw conclusions about the paleoenvironment and the evolution of this thermokarst affected landscape as well as the characterisation of the stored organic matter to access its vulnerability facing a changing Arctic.

The multidisciplinary analysis of permafrost cores from an upland to the coast on the Baldwin Peninsula reveals a consistent sedimentological signal indicating a common sediment origin and a predominance of aeolian deposition. Grain size distributions across all cores are largely unimodal and dominated by poorly sorted coarse silt with slightly secondary peaks in medium sand, confirming loessal origin with minor influences from ancient fluvial or reworked thermokarst processes. The presence of massive ground ice, organic-rich layers, and radiocarbon ages beyond 30–50 ka BP in the deeper cores aligns with characteristics of late Pleistocene Yedoma deposits.

The upland site represents a stable accumulation zone of Pleistocene deposition around 30 cal. ka BP with syngenetic ice-wedge formation and evidence of former thermokarst activity. Layers enriched in peat and TOC between 57–61 cm b.s.l. are similar to Yedoma paleocryosols reported for mid-Weichselian ponding conditions. Cryoturbation features and stable isotope shifts indicate past thaw–freeze cycles and aquatic influences. The thermokarst lake shows a clear transition between the lake and former-lake phase, with finer, laminated sediments with high TOC, TN and aquatic biomarker signals pointing to a young Holocene Lake (0.3–2 ka BP). The anomalously old 12 cal. ka BP sample at 434 cm likely reflects reworked material. Post-lake sedimentation includes terrestrial input through shoreline erosion as well as a high bioproductivity during the lake phase. The drained lake basin allows access to deep Yedoma layers due to thaw subsidence and therefore shows the oldest sediments with ages >50 cal. ka BP but has only a shallow modern peat layer due to its recent drainage. Its soil structure shows frost cracking and cryoturbation. The lower core retains an unfrozen talik as a relic from the former lake, while upper ice veins show renewed permafrost.

The marine core reflects terrestrial deposition later altered by coastal erosion and marine overprint, yet its biomarker and isotopic signatures still point to predominantly terrestrial organic matter. Large basal pebbles likely reflect ancient fluvial or thermokarst processes rather than original marine deposition.

The organic carbon content varies significant between the sites and permafrost layers, with

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the highest values in the active layer of the upland and lowest in the marine core. Across permafrost layers, TOC is highest in the active layer and lowest in perennial permafrost. Despite differences in carbon quantity, the origin of organic matter remains dominantly terrestrial, supported by C/N ratios, $\delta^{13}\text{C}$ values, and biomarker indices (ACL, p_{aq} , p_{wax}). Degradation proxies show thermokarst layers generally preserve slightly higher quality OM than cold-stored Yedoma sediments, likely due to high Holocene terrestrial input compensating microbial loss. Nevertheless, all sites show a high-quality OM. The variations between the cores likely reflect differing degradation states rather than fundamentally different OM sources. The Biomarkers support the assumption that all cores derive from a common parent material, consistent with physical sedimentology.

This study enriches the understanding of OM characteristics in thermokarst affected permafrost landscapes by combining sedimentary processes with biochemical indicators. It underlines the vulnerability of former frozen carbon pools in a warming Arctic and supports the need for further approaches to evaluate future carbon feedback as incubation experience or in-situ gas measurements. Especially when regarding the predicted shift from a lake to drained lake landscape in the Arctic, it is key to investigate carbon release in drained lake basins to estimate future GHG emissions.

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Appendix

Table A. 1. Raw data of the sedimentology.

Prefix	mean depth [cm b.s.l.]	abs. ice/water content [wt %]	mean grain size [µm]	clay [vol %]	silt [vol %]	silt acc. [vol %]	sand [vol %]	sand acc. [vol %]	sediment name
BAL24-T0-UL-A_0-8	4	87.98498	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_8-20	14	89.78495	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_20-22	21	76.92308	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_22-32	27	73.33916	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_32-37	34.5	31.6726	22.23978	4.089086	81.76628	85.85537	14.14463	100	Coarse Silt
BAL24-T0-UL-A_37-42	39.5	26.95716	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_42-47	44.5	27.06983	23.05513	3.917614	80.42891	84.34652	15.65348	100	Coarse Silt
BAL24-T0-UL-A_47-52	49.5	34.94898	21.94023	4.035608	82.07108	86.10669	13.89331	100	Coarse Silt
BAL24-T0-UL-A_52-57	54.5	57.46888	21.2729	4.755816	80.22266	84.97848	15.02152	100	Coarse Silt
BAL24-T0-UL-A_57-61	59	58.28066	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_61-63	62	46.72897	22.77758	4.486238	76.98021	81.46644	18.53356	100	Coarse Silt
BAL24-T0-UL-A_63-73	68	34.35074	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_73-77	75	51.46871	20.23842	5.765892	77.23882	83.00471	16.99529	100	Coarse Silt
BAL24-T0-UL-A_77-81	79	54.61957	22.61986	5.055724	75.24725	80.30298	19.69702	100	Coarse Silt
BAL24-T0-UL-A_81-85	83	52.80899	20.40054	5.462279	77.84958	83.31185	16.68815	100	Coarse Silt
BAL24-T0-UL-A_85-90	87.5	48.14815	22.09123	4.474893	75.67899	80.15389	19.84611	100	Coarse Silt
BAL24-T0-UL-A_90-95	92.5	44.30479	21.05137	4.78027	76.4869	81.26717	18.73283	100	Coarse Silt
BAL24-T0-UL-A_95-100	97.5	39.42857	21.29187	4.637566	76.51104	81.14861	18.85139	100	Coarse Silt
BAL24-T0-UL-A_100-103	101.5	37.3494	21.17054	4.529681	81.94168	86.47136	13.52864	100	Coarse Silt
BAL24-T0-UL-A_103-108	105.5	45.31371	21.27447	4.780184	81.88589	86.66607	13.05538	99.72145	Coarse Silt
BAL24-T0-UL-A_108-116	112	44.76285	26.38969	3.936395	74.35164	78.28803	21.71197	100	Coarse Silt
BAL24-T0-UL-A_116-121	118.5	44.44444	23.7659	4.537456	78.22674	82.7642	17.2358	100	Coarse Silt
BAL24-T0-UL-A_121-123	122	86.54354	25.64531	4.182551	77.06504	81.24759	18.75241	100	Coarse Silt
BAL24-T0-UL-A_123-133	128	99.9181	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_133-142	137.5	99.70646	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_142-150	146	66.66667	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_142-150	146	61.22449	23.24753	5.048397	77.01765	82.06605	17.93395	100	Coarse Silt
BAL24-T0-UL-A_150-155	152.5	55.24718	22.84674	4.403946	78.42086	82.82481	17.17519	100	Coarse Silt
BAL24-T0-UL-A_155-160	157.5	54.03304	22.68115	4.262241	80.12783	84.39007	15.60993	100	Coarse Silt
BAL24-T0-UL-A_160-164	162	53.90625	22.53607	5.007396	78.81736	83.82475	16.17525	100	Coarse Silt
BAL24-T0-UL-A_164-169	166.5	53.46535	21.29005	5.003849	77.89177	82.89562	17.10438	100	Coarse Silt
BAL24-T0-UL-A_169-175	172	56.18221	22.2587	4.800364	78.95244	83.7528	16.2472	100	Coarse Silt
BAL24-T0-UL-A_175-180	177.5	53.52941	24.039	3.795608	78.52672	82.32233	17.67767	100	Coarse Silt
BAL24-T0-UL-A_180-186	183	53.81264	21.85677	5.117247	78.31672	83.43396	16.56604	100	Coarse Silt
BAL24-T0-UL-A_186-196	191	56.05139	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_196-202	199	59.53488	19.75375	4.782722	79.95398	84.7367	15.2633	100	Coarse Silt
BAL24-T0-UL-A_196-202	199	59.53488	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-UL-A_202-207	204.5	65.32438	18.22156	5.536457	82.27574	87.81219	12.18781	100	Coarse Silt
BAL24-T0-UL-A_207-213	210	65.82827	21.00709	4.97112	78.4298	83.40092	16.59908	100	Coarse Silt
BAL24-T0-L-A-VIRBA_1-11	6	37.4695	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_1-11	6	43.29268	18.27359	5.375367	83.24763	88.62299	11.377008	100	Coarse Silt
BAL24-T0-L-A-VIRBA_11-16	13.5	30.79268	19.77444	5.022346	80.87861	85.90096	14.099042	100	Coarse Silt
BAL24-T0-L-A-VIRBA_16-21	18.5	NA	21.70398	4.367397	81.42866	85.79606	14.203944	100	Coarse Silt
BAL24-T0-L-A-VIRBA_21-26	23.5	39.05085	21.12154	4.703476	81.23996	85.94343	14.056566	100	Coarse Silt
BAL24-T0-L-A-VIRBA_26-30	28	NA	17.41997	5.468147	81.81921	87.28735	12.712645	100	Coarse Silt

Appendix

Prefix	mean depth [cm b.s.l.]	abs. ice/water content [wt %]	mean grain size [µm]	clay [vol %]	silt [vol %]	silt acc. [vol %]	sand [vol %]	sand acc. [vol %]	sediment name
BAL24-T0-L-A-VIRBA_30-34	32	65.48673	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_34-37	35.5	64.94253	15.57885	5.40229	83.69458	89.09687	10.903129	100	Medium Silt
BAL24-T0-L-A-VIRBA_37-42	39.5	67.48286	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_42-47	44.5	NA	16.31473	5.412329	83.26187	88.6742	11.325799	100	Coarse Silt
BAL24-T0-L-A-VIRBA_47-52	49.5	56.91188	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_52-57	54.5	NA	14.92544	5.736715	83.13001	88.86672	11.133277	100	Medium Silt
BAL24-T0-L-A-VIRBA_57-62	59.5	60.34913	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_62-67	64.5	NA	15.37606	5.588694	82.33388	87.92257	12.053578	99.97615	Medium Silt
BAL24-T0-L-A-VIRBA_67-72	69.5	66.72444	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_72-77	74.5	NA	15.77927	5.69667	82.00961	87.70628	12.293724	100	Coarse Silt
BAL24-T0-L-A-VIRBA_77-82	79.5	55.91739	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_82-87	84.5	NA	15.69652	4.902507	87.47016	92.37267	7.627333	100	Coarse Silt
BAL24-T0-L-A-VIRBA_87-91	89	46.68459	12.53332	6.206144	86.5607	92.76684	7.233158	100	Medium Silt
BAL24-T0-L-A-VIRBA_91-96	93.5	45.3411	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_96-98	97	43.27323	11.68426	6.635934	88.24236	94.8783	5.121703	100	Medium Silt
BAL24-T0-L-A-VIRBA_98-103	100.5	NA	11.63914	9.286964	84.41295	93.69991	6.300088	100	Medium Silt
BAL24-T0-L-A-VIRBA_103-108	105.5	46.81159	11.71753	7.424536	87.59867	95.02321	4.976794	100	Medium Silt
BAL24-T0-L-A-VIRBA_108-113	110.5	NA	10.43236	9.001512	86.692	95.69351	4.306489	100	Medium Silt
BAL24-T0-L-A-VIRBA_113-118	115.5	45.60944	13.27685	6.217235	85.98821	92.20545	7.794551	100	Medium Silt
BAL24-T0-L-A-VIRBA_118-123	120.5	NA	14.98052	5.540079	88.11079	93.65087	6.349128	100	Medium Silt
BAL24-T0-L-A-VIRBA_123-127	125	40.92284	14.41094	6.416653	86.42231	92.83896	7.161038	100	Medium Silt
BAL24-T0-L-A-VIRBA_127-137	132	57.89099	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_137-140	138.5	56.83453	17.73079	4.864729	81.30296	86.16769	13.83231	100	Coarse Silt
BAL24-T0-L-A-VIRBA_140-145	142.5	25.96917	20.87463	4.356131	82.56001	86.91614	13.08386	100	Coarse Silt
BAL24-T0-L-A-VIRBA_145-150	147.5	24.32821	20.13235	4.222679	83.72266	87.94534	12.054657	100	Coarse Silt
BAL24-T0-L-A-VIRBA_150-155	152.5	28.81699	21.71619	4.140065	81.78362	85.92368	14.076316	100	Coarse Silt
BAL24-T0-L-A-VIRBA_155-160	157.5	26.6858	22.25695	4.998728	78.97986	83.97859	16.02141	100	Coarse Silt
BAL24-T0-L-A-VIRBA_160-165	162.5	26.21359	20.93211	4.278689	82.5855	86.86419	13.135813	100	Coarse Silt
BAL24-T0-L-A-VIRBA_165-170	167.5	24.62936	20.16208	4.887149	82.83529	87.72243	12.277565	100	Coarse Silt
BAL24-T0-L-A-VIRBA_170-175	172.5	25.83222	20.50829	4.544479	82.14039	86.68487	13.31513	100	Coarse Silt
BAL24-T0-L-A-VIRBA_175-181	178	24.86441	22.45998	4.370409	81.31374	85.68414	14.315855	100	Coarse Silt
BAL24-T0-L-A-VIRBA_181-187	184	24.7451	20.32145	4.307433	82.72753	87.03496	12.965039	100	Coarse Silt
BAL24-T0-L-A-VIRBA_187-193	190	24.50903	20.32145	4.307433	82.72753	87.03496	12.965039	100	Coarse Silt
BAL24-T0-L-A-VIRBA_193-198	195.5	24.81442	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_198-201	199.5	25.29412	20.8681	4.449169	82.16566	86.61483	13.385167	100	Coarse Silt
BAL24-T0-L-A-VIRBA_201-206	203.5	26.40227	20.59522	4.324413	82.14642	86.47083	13.529168	100	Coarse Silt
BAL24-T0-L-A-VIRBA_206-211	208.5	26.86671	24.6684	3.261335	80.54684	83.80818	16.191825	100	Coarse Silt
BAL24-T0-L-A-VIRBA_211-216	213.5	29.0152	22.6263	4.128251	80.62312	84.75137	15.248625	100	Coarse Silt
BAL24-T0-L-A-VIRBA_216-221	218.5	27.53807	21.75626	4.019066	82.25556	86.27463	13.72537	100	Coarse Silt
BAL24-T0-L-A-VIRBA_221-226	223.5	28.00241	23.21871	4.0752	80.53469	84.60989	15.390106	100	Coarse Silt
BAL24-T0-L-A-VIRBA_226-231	228.5	29.8278	23.62217	4.726057	78.25681	82.98287	17.017129	100	Coarse Silt
BAL24-T0-L-A-VIRBA_231-241	236	24.8059	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_241-244	242.5	25.93718	28.85455	3.306867	74.42857	77.73544	22.264558	100	Coarse Silt
BAL24-T0-L-A-VIRBA_244-249	246.5	28.6399	28.95546	3.578458	72.40045	75.97891	24.021092	100	Coarse Silt
BAL24-T0-L-A-VIRBA_249-254	251.5	28.35561	36.49246	2.264748	71.76101	74.02576	25.97424	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_254-259	256.5	28	37.63426	2.861683	64.52693	67.38861	32.611391	100	Very Coarse Silt

Appendix

Prefix	mean depth [cm b.s.l.]	abs. ice/water content [wt %]	mean grain size [µm]	clay [vol %]	silt [vol %]	silt acc. [vol %]	sand [vol %]	sand acc. [vol %]	sediment name
BAL24-T0-L-A-VIRBA_259-264	261.5	28.11989	35.14786	3.090284	65.30714	68.39743	31.602575	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_264-269	266.5	26.67423	44.54215	2.343448	57.02873	59.37218	40.627818	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_269-274	271.5	24.26266	61.16878	1.604184	47.10108	48.70526	51.294738	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_274-279	276.5	26.65535	29.95883	3.783835	69.92848	73.71232	26.287684	100	Coarse Silt
BAL24-T0-L-A-VIRBA_279-285	282	26.18718	39.94387	2.785307	63.90733	66.69263	33.307365	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_285-291	288	25.50562	34.28687	2.927489	68.06095	70.98844	29.011562	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_291-297	294	22.29299	38.04889	2.470771	63.62063	66.0914	33.908596	100	Very Coarse Silt
BAL24-T0-L-A-VIRBA_297-307	302	26.51347	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-L-A-VIRBA_307-311	309	35.44715	30.57104	3.129967	74.38845	77.51841	22.481587	100	Coarse Silt
BAL24-T0-DLB-A_0-1	0.5	72.31405	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_1-11	6	57.8003	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_11-13	12	46.94737	20.04775	4.948802	82.00391	86.95271	13.04729	100	Coarse Silt
BAL24-T0-DLB-A_13-19	16	42.45333	22.20757	4.249615	78.86269	83.11231	16.88769	100	Coarse Silt
BAL24-T0-DLB-A_19-23	21	38.70095	22.28019	4.980125	78.19979	83.17991	16.82009	100	Coarse Silt
BAL24-T0-DLB-A_23-27	25	38.31169	26.60083	3.853828	76.62118	80.47501	19.52499	100	Coarse Silt
BAL24-T0-DLB-A_27-32	29.5	33.78747	23.94192	4.293517	78.80491	83.09842	16.90158	100	Coarse Silt
BAL24-T0-DLB-A_32-38	35	35.84432	24.98128	4.149495	78.10177	82.25126	17.74874	100	Coarse Silt
BAL24-T0-DLB-A_38-42	40	36.16037	25.50787	4.122558	78.59216	82.71472	17.28528	100	Coarse Silt
BAL24-T0-DLB-A_42-46	44	34.12395	27.41325	3.421792	77.67638	81.09817	18.90183	100	Coarse Silt
BAL24-T0-DLB-A_46-51	48.5	33.79265	28.21096	3.752363	76.05842	79.81079	20.18921	100	Coarse Silt
BAL24-T0-DLB-A_51-56	53.5	34.68635	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_56-58	57	33.00589	23.4947	4.042978	81.11152	85.1545	14.8455	100	Coarse Silt
BAL24-T0-DLB-A_58-65	61.5	34.30762	24.71895	4.104203	79.02736	83.13156	16.86844	100	Coarse Silt
BAL24-T0-DLB-A_65-69	67	35.76642	22.77109	4.318298	79.96863	84.28693	15.71307	100	Coarse Silt
BAL24-T0-DLB-A_69-73	71	34.95146	23.53615	4.163384	78.90748	83.07087	16.92913	100	Coarse Silt
BAL24-T0-DLB-A_73-78	75.5	37.63626	21.43625	4.481403	81.22553	85.70693	14.29307	100	Coarse Silt
BAL24-T0-DLB-A_78-84	81	NA	26.46283	3.450107	78.44084	81.89095	18.10905	100	Coarse Silt
BAL24-T0-DLB-A_84-90	87	33.21779	22.96875	4.211709	81.54565	85.75736	14.24264	100	Coarse Silt
BAL24-T0-DLB-A_90-96	93	NA	18.78735	6.069748	78.98664	85.05638	14.94362	100	Coarse Silt
BAL24-T0-DLB-A_96-106	101	34.36727	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_106-108	107	31.57895	28.08025	3.455184	76.67582	80.13101	19.86899	100	Coarse Silt
BAL24-T0-DLB-A_108-113	110.5	33.61823	25.18158	3.704584	78.04237	81.74696	18.25304	100	Coarse Silt
BAL24-T0-DLB-A_113-118	115.5	NA	24.67469	4.139495	78.70271	82.8422	17.1578	100	Coarse Silt
BAL24-T0-DLB-A_118-123	120.5	36.65615	23.02423	4.241721	80.2821	84.52382	15.47618	100	Coarse Silt
BAL24-T0-DLB-A_123-128	125.5	NA	22.84029	4.112543	78.81923	82.93178	17.06822	100	Coarse Silt
BAL24-T0-DLB-A_128-133	130.5	36.02151	25.71241	3.794521	76.31761	80.11213	19.88787	100	Coarse Silt
BAL24-T0-DLB-A_133-137	135	NA	27.17673	3.675412	75.53228	79.20769	20.79231	100	Coarse Silt
BAL24-T0-DLB-A_137-142	139.5	30.55787	26.40352	3.94648	75.09022	79.0367	20.9633	100	Coarse Silt
BAL24-T0-DLB-A_142-147	144.5	NA	31.78688	2.527226	75.6693	78.19652	21.80348	100	Very Coarse Silt
BAL24-T0-DLB-A_147-152	149.5	32.45455	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_152-154	153	31.33641	30.57534	3.060812	72.77409	75.8349	24.1651	100	Coarse Silt
BAL24-T0-DLB-A_154-159	156.5	NA	26.91889	3.608571	76.0787	79.68727	20.31273	100	Coarse Silt
BAL24-T0-DLB-A_159-164	161.5	28.20676	29.61542	3.04112	74.82129	77.86241	22.13759	100	Coarse Silt
BAL24-T0-DLB-A_164-169	166.5	NA	32.36259	2.480526	72.7557	75.23623	24.76377	100	Very Coarse Silt
BAL24-T0-DLB-A_169-174	171.5	30.8084	30.12251	3.402629	74.01819	77.42082	22.57918	100	Coarse Silt
BAL24-T0-DLB-A_174-179	176.5	NA	30.71379	2.699199	74.88156	77.58075	22.41925	100	Coarse Silt

Appendix

Prefix	mean depth [cm b.s.l.]	abs. ice/wa- ter content [wt %]	mean grain size [μm]	clay [vol %]	silt [vol %]	silt acc. [vol %]	sand [vol %]	sand acc. [vol %]	sediment name
BAL24-T0-DLB-A_179-184	181.5	29.29178	26.58622	3.29042	78.54995	81.84037	18.15963	100	Coarse Silt
BAL24-T0-DLB-A_184-189	186.5	NA	30.55884	2.825594	74.84361	77.6692	22.3308	100	Coarse Silt
BAL24-T0-DLB-A_189-194	191.5	28.51109	32.1898	2.608155	72.01442	74.62257	25.37743	100	Very Coarse Silt
BAL24-T0-DLB-A_194-199	196.5	NA	31.00712	2.668665	73.62478	76.29344	23.70656	100	Coarse Silt
BAL24-T0-DLB-A_199-209	204	27.59014	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-DLB-A_209-214	211.5	28.12331	28.26309	3.034177	76.80953	79.84371	20.15629	100	Coarse Silt
BAL24-T0-M1-PUSH_0-5	2.5	47.55102	29.51701	2.987537	75.23651	78.22404	21.77596	100	Coarse Silt
BAL24-T0-M1-PUSH_5-10	7.5	44.18819	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-M1-PUSH_10-13	11.5	39.97628	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-M1-PUSH_13-18	15.5	39.41134	22.5091	3.966835	81.16358	85.13041	14.86959	100	Coarse Silt
BAL24-T0-M1-PUSH_18-23	20.5	38.90034	33.48276	2.698098	71.84072	74.53882	25.46118	100	Very Coarse Silt
BAL24-T0-M1-PUSH_23-28	25.5	32.87995	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-M1-PUSH_28-33	30.5	31.84524	32.62821	2.585367	75.23277	77.81814	22.18186	100	Very Coarse Silt
BAL24-T0-M1-PUSH_33-38	35.5	30.36903	33.52656	3.348088	70.3949	73.74299	26.25701	100	Very Coarse Silt
BAL24-T0-M1-PUSH_38-42	40	30.75299	29.29419	2.855051	76.52568	79.38073	20.61927	100	Coarse Silt
BAL24-T0-M1-PUSH_42-47	44.5	27.57808	NA	NA	NA	NA	NA	NA	NA
BAL24-T0-M1-PUSH_47-52	49.5	24.10667	39.61364	2.685043	61.18244	63.86748	36.13252	100	Very Coarse Silt
BAL24-T0-M1-PUSH_52-55	53.5	25.36294	36.10735	2.921177	64.51046	67.43163	32.56837	100	Very Coarse Silt

Table A. 2. Raw data of the hydrochemistry.

Prefix	mean depth [cm b.s.l.]	DOC [mg L ⁻¹]	mean $\delta^{18}\text{O}$ [‰ vs. SMOW]	mean δD [‰ vs. SMOW]	mean d excess [‰]	EC [$\mu\text{S cm}^{-1}$]	pH [-]
BAL24-T0-UL-A_8-20	14	42.05	-15.08081	-112.2792	8.367266	44.62	4.87
BAL24-T0-UL-A_42-47	44.5	33.075	-16.16772	-118.5781	10.763627	49.51	6.3
BAL24-T0-UL-A_52-57	54.5	40.31	-16.35906	-120.9678	9.904634	88.08	6.36
BAL24-T0-UL-A_73-77	75	223.3	-16.8575	-128.0053	6.854633	48.89	6.03
BAL24-T0-UL-A_81-85	83	131.4	-16.61907	-128.2484	4.704199	156.2	6.74
BAL24-T0-UL-A_90-95	92.5	159.1	NA	NA	NA	205.9	6.93
BAL24-T0-UL-A_103-108	105.5	99.9	-14.63003	-120.0985	-3.05824	256.2	7.08
BAL24-T0-UL-A_116-121	118.5	199.8	-15.49239	-125.6627	-1.723543	409.3	7.4
BAL24-T0-UL-A_123-133	128	5.783	-20.35954	-160.2123	2.663984	43.41	7.17
BAL24-T0-UL-A_133-142	137.5	9.155	-19.83193	-155.21	3.445384	73.22	7.43
BAL24-T0-UL-A_150-155	152.5	857.6	-19.64046	-152.8184	4.305286	1405	6.97
BAL24-T0-UL-A_160-164	162	1075	-19.62421	-152.7071	4.286592	1656	6.87
BAL24-T0-UL-A_169-175	172	1342	-19.72329	-153.2688	4.517505	1998	6.54
BAL24-T0-UL-A_180-186	183	1707	-19.78382	-151.8303	6.440239	2379	6.4
BAL24-T0-UL-A_202-207	204.5	940	-19.43193	-150.6093	4.846156	1385	5.68
BAL24-T0-L-A-VIRBA_16-21	18.5	113.8	-17.03143	-135.1828	1.06863598	589.4	7.81
BAL24-T0-L-A-VIRBA_26-30	28	143.7	-17.33175	-138.9616	-0.30764745	721.3	7.82
BAL24-T0-L-A-VIRBA_42-47	44.5	142.9	-17.70276	-141.2318	0.39032678	91	7.73
BAL24-T0-L-A-VIRBA_52-57	54.5	151.7	NA	NA	NA	695.6	7.92
BAL24-T0-L-A-VIRBA_62-67	64.5	126.3	NA	NA	NA	575.7	7.66
BAL24-T0-L-A-VIRBA_72-77	74.5	160.35	-18.19971	-146.1684	-0.57065067	721.6	7.68
BAL24-T0-L-A-VIRBA_82-87	84.5	130.55	-18.41303	-147.4176	-0.11343559	618.8	7.74
BAL24-T0-L-A-VIRBA_98-103	100.5	183.85	-18.61363	-149.3889	-0.47986551	802.1	7.57
BAL24-T0-L-A-VIRBA_108-113	110.5	188.95	-18.71228	-150.6735	-0.97525971	862.6	7.76

Appendix

Prefix	mean depth [cm b.s.l.]	DOC [mg L ⁻¹]	mean δ ¹⁸ O [‰ vs. SMOW]	mean δD [‰ vs. SMOW]	mean d excess [-]	EC [μS cm ⁻¹]	pH [-]
BAL24-T0-L-A-VIRBA_118-123	120.5	151.4	-18.8376	-151.3072	-0.60640004	634	7.72
BAL24-T0-L-A-VIRBA_140-145	142.5	121.5	-18.93901	-151.2825	0.22961042	569.8	7.28
BAL24-T0-L-A-VIRBA_150-155	152.5	129.1	-18.94678	-151.2433	0.33095647	470.4	7.22
BAL24-T0-L-A-VIRBA_160-165	162.5	124.65	-18.90321	-152.3942	-1.16849969	471.4	7.73
BAL24-T0-L-A-VIRBA_170-175	172.5	125.75	-18.8772	-152.0884	-1.07083759	459.9	7.61
BAL24-T0-L-A-VIRBA_201-206	203.5	136.7	NA	NA	NA	438.9	7.58
BAL24-T0-L-A-VIRBA_211-216	213.5	153.7	-18.62386	-147.6998	1.29112189	520.3	8.09
BAL24-T0-L-A-VIRBA_221-226	223.5	154.15	-18.55659	-148.9103	-0.45764161	572.5	7.7
BAL24-T0-L-A-VIRBA_244-249	246.5	213.5	NA	NA	NA	821.1	8.22
BAL24-T0-L-A-VIRBA_254-259	256.5	195.7	-17.98037	-143.3454	0.49753938	783.3	7.88
BAL24-T0-L-A-VIRBA_264-269	266.5	186.8	-17.82924	-142.6691	-0.03516393	708	8.03
BAL24-T0-L-A-VIRBA_274-279	276.5	204.95	NA	NA	NA	799.4	7.85
BAL24-T0-L-A-VIRBA_285-291	288	177.75	-17.52408	-133.2424	6.95026549	784.3	7.8
BAL24-T0-DLB-A_13-19	16	46.63	-14.3174	-112.1498	2.389364	771.9	7.69
BAL24-T0-DLB-A_23-27	25	47.095	-14.24584	-112.6655	1.301212	853.3	8.03
BAL24-T0-DLB-A_32-38	35	93.55	-15.37501	-121.0903	1.909739	980.8	7.9
BAL24-T0-DLB-A_42-46	44	146.25	-16.57504	-130.5702	2.030058	899.4	7.8
BAL24-T0-DLB-A_58-65	61.5	211.25	-18.05706	-139.5861	4.870412	958.8	7.62
BAL24-T0-DLB-A_69-73	71	235.35	-18.5347	-142.6532	5.624325	850.7	7.88
BAL24-T0-DLB-A_78-84	81	211.5	-18.89099	-143.8412	7.286734	887.7	7.97
BAL24-T0-DLB-A_90-96	93	218.1	-19.11008	-145.2686	7.612041	987.9	7.76
BAL24-T0-DLB-A_113-118	115.5	241.8	NA	NA	NA	1048	8.08
BAL24-T0-DLB-A_123-128	125.5	255.25	-19.56639	-146.3475	10.183626	1071	8.01
BAL24-T0-DLB-A_133-137	135	198.35	NA	NA	NA	733.9	7.95
BAL24-T0-DLB-A_142-147	144.5	173.1	NA	NA	NA	742.9	7.99
BAL24-T0-DLB-A_154-159	156.5	179.1	-18.93635	-142.6799	8.810921	815.3	7.96
BAL24-T0-DLB-A_164-169	166.5	232.2	NA	NA	NA	961.4	8.41
BAL24-T0-DLB-A_174-179	176.5	217.6	-19.53497	-141.6816	14.598104	944.4	8.05
BAL24-T0-DLB-A_184-189	186.5	197.45	-19.24295	-143.8099	10.133716	923.7	8
BAL24-T0-DLB-A_194-199	196.5	312.25	NA	NA	NA	1029	8.17
BAL24-T0-M1-PUSH_13-18	15.5	66.71	-13.28085	-102.5079	3.738853	17290	7.47
BAL24-T0-M1-PUSH_33-38	35.5	45.18	-13.41767	-102.2469	5.094446	19380	7.99
BAL24-T0-M1-PUSH_47-52	49.5	41.375	-13.14296	-101.5741	3.569637	20390	7.83

Table A. 3. Raw data of the biogeochemistry.

Prefix	mean depth [cm b.s.l.]	TOC [wt %]	TN [wt %]	C/N [-]	δ ¹³ C [‰ vs. VPDB]
BAL24-T0-UL-A_0-8	4	42.404752	1.197	35.42586	-29.75592
BAL24-T0-UL-A_8-20	14	44.415247	1.057	42.0201	-28.32276
BAL24-T0-UL-A_20-22	21	37.199883	2.0145	18.46606	-27.29474
BAL24-T0-UL-A_22-32	27	32.420201	1.8705	17.33237	NA
BAL24-T0-UL-A_32-37	34.5	2.133366	0.17	12.54921	-27.93374
BAL24-T0-UL-A_37-42	39.5	1.848613	0.1115	16.57949	NA
BAL24-T0-UL-A_42-47	44.5	1.955831	0.121	16.16389	-27.61642
BAL24-T0-UL-A_47-52	49.5	2.645075	0.1765	14.98626	-27.34555
BAL24-T0-UL-A_52-57	54.5	5.392463	0.3615	14.91691	-27.06595
BAL24-T0-UL-A_57-61	59	16.284694	0.8155	19.96897	-27.17738

Appendix

Prefix	mean depth [cm b.s.l.]	TOC [wt %]	TN [wt %]	C/N [-]	$\delta^{13}\text{C}$ [‰ vs. VPDB]
BAL24-T0-UL-A_61-63	62	4.388157	0.322	13.62782	-26.48602
BAL24-T0-UL-A_63-73	68	3.51261	0.311	11.29457	NA
BAL24-T0-UL-A_73-77	75	4.431164	0.3615	12.25772	-26.18605
BAL24-T0-UL-A_77-81	79	4.421886	0.389	11.36732	-26.05933
BAL24-T0-UL-A_81-85	83	4.141357	0.355	11.66579	-26.0627
BAL24-T0-UL-A_85-90	87.5	4.875413	0.397	12.28064	-26.29928
BAL24-T0-UL-A_90-95	92.5	4.214175	0.3535	11.92129	-26.11445
BAL24-T0-UL-A_95-100	97.5	4.368179	0.3815	11.45001	-26.31385
BAL24-T0-UL-A_100-103	101.5	4.334399	0.3715	11.66729	-25.97824
BAL24-T0-UL-A_103-108	105.5	3.693207	0.324	11.39879	-25.81215
BAL24-T0-UL-A_108-116	112	3.457644	0.3125	11.06446	-26.08825
BAL24-T0-UL-A_116-121	118.5	3.904331	0.3275	11.92162	-26.04694
BAL24-T0-UL-A_121-123	122	2.836034	0.275	10.31285	-25.88572
BAL24-T0-UL-A_123-133	128	3.544638	0.391	9.06557	NA
BAL24-T0-UL-A_133-142	137.5	2.742171	0.243	11.28465	NA
BAL24-T0-UL-A_142-150	146	4.38217	0.34	12.88873	NA
BAL24-T0-UL-A_142-150	146	5.162768	0.4065	12.70054	-26.3262
BAL24-T0-UL-A_150-155	152.5	4.750294	0.3865	12.29054	-26.37964
BAL24-T0-UL-A_155-160	157.5	5.703895	0.469	12.16182	-26.18455
BAL24-T0-UL-A_160-164	162	4.752249	0.4035	11.77757	-26.23169
BAL24-T0-UL-A_164-169	166.5	5.142456	0.449	11.45313	-26.28077
BAL24-T0-UL-A_169-175	172	5.98625	0.458	13.07041	-26.2642
BAL24-T0-UL-A_175-180	177.5	6.186264	0.515	12.01216	-26.17713
BAL24-T0-UL-A_180-186	183	6.86349	0.5555	12.35552	-26.28104
BAL24-T0-UL-A_186-196	191	8.0954	0.5455	14.84033	-26.11563
BAL24-T0-UL-A_196-202	199	6.464248	0.5515	11.72121	-26.29726
BAL24-T0-UL-A_196-202	199	8.059861	0.531	15.17865	-26.29726
BAL24-T0-UL-A_202-207	204.5	5.015068	0.436	11.50245	-26.01277
BAL24-T0-UL-A_207-213	210	6.078353	0.5015	12.12034	-26.21934
BAL24-T0-L-A-VIRBA_1-11	6	4.3978818	0.3635	12.098712	-25.9575
BAL24-T0-L-A-VIRBA_1-11	6	4.1149664	0.3915	10.51077	-25.9575
BAL24-T0-L-A-VIRBA_11-16	13.5	3.5183134	0.3055	11.516574	-26.07906
BAL24-T0-L-A-VIRBA_16-21	18.5	2.2221847	0.198	11.223155	-26.48833
BAL24-T0-L-A-VIRBA_21-26	23.5	4.567094	0.298	15.325819	-26.91835
BAL24-T0-L-A-VIRBA_26-30	28	14.8916718	0.8015	18.579753	-27.06007
BAL24-T0-L-A-VIRBA_30-34	32	11.876732	0.7765	15.295212	-27.7389
BAL24-T0-L-A-VIRBA_34-37	35.5	14.7244447	0.892	16.507225	-27.39818
BAL24-T0-L-A-VIRBA_37-42	39.5	17.8605949	1.0485	17.034425	NA
BAL24-T0-L-A-VIRBA_42-47	44.5	24.3962746	1.2815	19.03728	-27.03801
BAL24-T0-L-A-VIRBA_47-52	49.5	15.8813932	1.07	14.842424	-27.10747
BAL24-T0-L-A-VIRBA_52-57	54.5	21.2254205	1.114	19.05334	-27.09753
BAL24-T0-L-A-VIRBA_57-62	59.5	18.7040005	1.039	18.001925	-27.09933
BAL24-T0-L-A-VIRBA_62-67	64.5	19.0191548	0.965	19.708969	-26.89464
BAL24-T0-L-A-VIRBA_67-72	69.5	21.5813095	1.024	21.075498	-27.41507
BAL24-T0-L-A-VIRBA_72-77	74.5	19.2826855	1.0275	18.766604	-26.83618
BAL24-T0-L-A-VIRBA_77-82	79.5	14.6873548	0.824	17.82446	-26.83288
BAL24-T0-L-A-VIRBA_82-87	84.5	8.3422914	0.5415	15.405894	-27.02691

Appendix

Prefix	mean depth [cm b.s.l.]	TOC [wt %]	TN [wt %]	C/N [-]	$\delta^{13}\text{C}$ [‰ vs. VPDB]
BAL24-T0-L-A-VIRBA_87-91	89	7.6817501	0.817	9.402387	-27.20079
BAL24-T0-L-A-VIRBA_91-96	93.5	7.3699013	0.5255	14.02455	NA
BAL24-T0-L-A-VIRBA_96-98	97	5.9515512	0.474	12.556015	-27.56028
BAL24-T0-L-A-VIRBA_98-103	100.5	6.4226206	0.4995	12.858099	-27.89548
BAL24-T0-L-A-VIRBA_103-108	105.5	6.4669803	0.473	13.672263	-27.84737
BAL24-T0-L-A-VIRBA_108-113	110.5	8.045323	0.511	15.744272	-28.6055
BAL24-T0-L-A-VIRBA_113-118	115.5	6.0295037	0.453	13.310163	-28.46013
BAL24-T0-L-A-VIRBA_118-123	120.5	5.6388233	0.443	12.728721	-28.22503
BAL24-T0-L-A-VIRBA_123-127	125	4.7733779	0.3355	14.227654	-27.59902
BAL24-T0-L-A-VIRBA_127-137	132	13.6370179	0.7455	18.292445	NA
BAL24-T0-L-A-VIRBA_137-140	138.5	14.8954943	0.8285	17.978871	-27.02562
BAL24-T0-L-A-VIRBA_140-145	142.5	1.7520784	0.138	12.696221	-27.06198
BAL24-T0-L-A-VIRBA_145-150	147.5	1.7155261	0.1135	15.114767	-26.93592
BAL24-T0-L-A-VIRBA_150-155	152.5	2.0590155	0.1555	13.241257	-26.73859
BAL24-T0-L-A-VIRBA_155-160	157.5	1.7832901	0.1255	14.209483	-26.78139
BAL24-T0-L-A-VIRBA_160-165	162.5	1.7157634	0.1555	11.033848	-26.78885
BAL24-T0-L-A-VIRBA_165-170	167.5	1.5202062	0.126	12.065129	-26.80448
BAL24-T0-L-A-VIRBA_170-175	172.5	1.5287158	0.1225	12.479313	-26.64466
BAL24-T0-L-A-VIRBA_175-181	178	1.4364977	0.1135	12.656368	-26.52386
BAL24-T0-L-VIRBA_181-187	184	1.3388378	0.1075	12.454305	-26.25675
BAL24-T0-L-A-VIRBA_187-193	190	1.5501021	0.118	13.136459	-26.45784
BAL24-T0-L-A-VIRBA_193-198	195.5	1.6936209	0.01	NA	NA
BAL24-T0-L-A-VIRBA_198-201	199.5	1.8165475	0.1335	13.607098	-26.65473
BAL24-T0-L-A-VIRBA_201-206	203.5	1.6903925	0.1325	12.75768	-26.72718
BAL24-T0-L-A-VIRBA_206-211	208.5	1.5854789	0.106	14.957349	-26.78911
BAL24-T0-L-A-VIRBA_211-216	213.5	2.4843497	0.1795	13.840388	-26.69328
BAL24-T0-L-A-VIRBA_216-221	218.5	2.4439894	0.1785	13.691817	-26.81237
BAL24-T0-L-A-VIRBA_221-226	223.5	2.2634744	0.2075	10.90831	-26.46927
BAL24-T0-L-A-VIRBA_226-231	228.5	3.4450036	0.234	14.722238	-26.85643
BAL24-T0-L-A-VIRBA_231-241	236	2.7923137	0.176	15.865419	NA
BAL24-T0-L-A-VIRBA_241-244	242.5	2.7969065	0.226	12.375693	-26.40654
BAL24-T0-L-A-VIRBA_244-249	246.5	2.145	0.1995	10.75188	-26.19399
BAL24-T0-L-A-VIRBA_249-254	251.5	1.5864755	0.152	10.437339	-25.91843
BAL24-T0-L-A-VIRBA_254-259	256.5	1.5307057	0.148	10.342606	-25.93311
BAL24-T0-L-A-VIRBA_259-264	261.5	2.0488653	0.1635	12.531286	-26.11144
BAL24-T0-L-A-VIRBA_264-269	266.5	1.721033	0.152	11.322586	-25.91325
BAL24-T0-L-A-VIRBA_269-274	271.5	0.6743496	0.01	NA	-26.11358
BAL24-T0-L-A-VIRBA_274-279	276.5	1.1531283	0.1245	9.262075	-25.44191
BAL24-T0-L-A-VIRBA_279-285	282	0.9227866	0.01	NA	-25.14901
BAL24-T0-L-A-VIRBA_285-291	288	0.9332479	0.01	NA	-24.97012
BAL24-T0-L-A-VIRBA_291-297	294	0.6033988	0.01	NA	-24.7424
BAL24-T0-L-A-VIRBA_297-307	302	1.5378102	0.01	NA	NA
BAL24-T0-L-A-VIRBA_307-311	309	1.5555746	0.13575	11.459113	-25.74635
BAL24-T0-DLB-A_0-1	0.5	18.380763	1.5735	11.68145	-36.49745
BAL24-T0-DLB-A_1-11	6	8.097071	0.536	15.10648	NA
BAL24-T0-DLB-A_11-13	12	5.153482	0.3485	14.78761	-26.33504
BAL24-T0-DLB-A_13-19	16	3.357859	0.2305	14.56772	-26.25033

Appendix

Prefix	mean depth [cm b.s.l.]	TOC [wt %]	TN [wt %]	C/N [-]	$\delta^{13}\text{C}$ [‰ vs. VPDB]
BAL24-T0-DLB-A_19-23	21	2.761938	0.2305	11.98238	-25.92509
BAL24-T0-DLB-A_23-27	25	2.385997	0.2045	11.66747	-25.71227
BAL24-T0-DLB-A_27-32	29.5	2.417581	0.2035	11.88001	-24.97002
BAL24-T0-DLB-A_32-38	35	3.09344	0.2595	11.92077	-25.09181
BAL24-T0-DLB-A_38-42	40	2.181106	0.199	10.96033	-24.93879
BAL24-T0-DLB-A_42-46	44	2.626967	0.2415	10.87771	-25.15381
BAL24-T0-DLB-A_46-51	48.5	2.442182	0.222	11.00082	-25.0079
BAL24-T0-DLB-A_51-56	53.5	3.232071	0.257	12.57615	NA
BAL24-T0-DLB-A_56-58	57	2.938902	0.23	12.77784	-24.99468
BAL24-T0-DLB-A_58-65	61.5	3.422192	0.2775	12.33222	-25.19139
BAL24-T0-DLB-A_65-69	67	3.341776	0.2725	12.2634	-25.60757
BAL24-T0-DLB-A_69-73	71	3.376273	0.2835	11.90925	-25.50874
BAL24-T0-DLB-A_73-78	75.5	4.835132	0.377	12.82528	-26.10669
BAL24-T0-DLB-A_78-84	81	3.705011	0.3145	11.78064	-25.78487
BAL24-T0-DLB-A_84-90	87	2.813204	0.2345	11.9966	-25.37337
BAL24-T0-DLB-A_90-96	93	2.722128	0.24475	11.12207	-25.69591
BAL24-T0-DLB-A_96-106	101	3.439864	0.2565	13.41077	NA
BAL24-T0-DLB-A_106-108	107	3.039286	0.2205	13.78361	-25.81563
BAL24-T0-DLB-A_108-113	110.5	3.5208	0.2615	13.46386	-26.29006
BAL24-T0-DLB-A_113-118	115.5	2.773378	0.21575	12.85459	-25.43005
BAL24-T0-DLB-A_118-123	120.5	3.503892	0.2705	12.95339	-25.77751
BAL24-T0-DLB-A_123-128	125.5	3.728514	0.2935	12.70363	-25.77952
BAL24-T0-DLB-A_128-133	130.5	3.323376	0.2325	14.29409	-25.96818
BAL24-T0-DLB-A_133-137	135	2.520167	0.187	13.47683	-25.85085
BAL24-T0-DLB-A_137-142	139.5	2.173149	0.15825	13.73238	-25.63851
BAL24-T0-DLB-A_142-147	144.5	1.796353	0.162	11.0886	-25.0832
BAL24-T0-DLB-A_147-152	149.5	2.953993	0.186	15.88168	NA
BAL24-T0-DLB-A_152-154	153	2.449105	0.1725	14.19771	-25.30685
BAL24-T0-DLB-A_154-159	156.5	1.716261	0.1415	12.12905	-25.00082
BAL24-T0-DLB-A_159-164	161.5	1.372018	0.112	12.25016	-24.69777
BAL24-T0-DLB-A_164-169	166.5	1.191931	0.117	10.18744	-24.45007
BAL24-T0-DLB-A_169-174	171.5	1.373012	0.10825	12.68372	-24.87394
BAL24-T0-DLB-A_174-179	176.5	1.548614	0.142	10.90574	-25.03603
BAL24-T0-DLB-A_179-184	181.5	1.244712	0.01	NA	-24.93435
BAL24-T0-DLB-A_184-189	186.5	1.577965	0.1345	11.73208	-24.79041
BAL24-T0-DLB-A_189-194	191.5	1.466503	0.12	12.22086	-25.03898
BAL24-T0-DLB-A_194-199	196.5	1.559061	0.1405	11.09652	-24.92143
BAL24-T0-DLB-A_199-209	204	1.416598	0.01	NA	NA
BAL24-T0-DLB-A_209-214	211.5	1.373285	0.118	11.638	-24.51508
BAL24-T0-M1-PUSH_0-5	2.5	3.033437	0.2095	14.47941	-26.45659
BAL24-T0-M1-PUSH_5-10	7.5	2.733394	0.1385	19.7357	NA
BAL24-T0-M1-PUSH_10-13	11.5	2.759908	0.1785	15.46167	-26.62513
BAL24-T0-M1-PUSH_13-18	15.5	1.961512	0.1505	13.0333	-26.48704
BAL24-T0-M1-PUSH_18-23	20.5	2.228241	0.1595	13.97016	-26.55382
BAL24-T0-M1-PUSH_23-28	25.5	1.942485	0.01	NA	NA
BAL24-T0-M1-PUSH_28-33	30.5	1.977977	0.1265	15.63619	-26.31708
BAL24-T0-M1-PUSH_33-38	35.5	1.362283	0.115	11.84594	-26.00289

Appendix

Prefix	mean depth [cm b.s.l.]	TOC [wt %]	TN [wt %]	C/N [-]	$\delta^{13}\text{C}$ [‰ vs. VPDB]
BAL24-T0-M1-PUSH_38-42	40	1.535652	0.01	NA	-25.88241
BAL24-T0-M1-PUSH_42-47	44.5	1.143047	0.01	NA	NA
BAL24-T0-M1-PUSH_47-52	49.5	1.180917	0.01	NA	-25.95536
BAL24-T0-M1-PUSH_52-55	53.5	1.490674	0.01	NA	-26.1697

Table A. 4. Raw data of the biomarker parameters.

Prefix	mean depth [cm b.s.l.]	alkane conc. C_{21-35} [$\mu\text{g gSed}^{-1}$]	alkane conc. C_{21-35} [$\mu\text{g gTOC}^{-1}$]	$\text{P}_{\text{aq}} \text{C}_{23-33}$ [-]	$\text{P}_{\text{wax}} \text{C}_{23-33}$ [-]	$\text{CPI } \text{C}_{23-33}$ [-]	$\text{ACL } \text{C}_{23-33}$ [-]
BAL24-T0-UL-A_22-32	27	1115.15277	3439.70626	0.02181736	0.9795025	76.878005	30.19112
BAL24-T0-UL-A_37-42	39.5	11.050796	597.66338	0.20080849	0.8316055	6.926095	28.65358
BAL24-T0-UL-A_63-73	68	2.36492	67.32657	0.54065125	0.6812005	8.078622	26.77215
BAL24-T0-UL-A_142-150	146	4.265215	97.33112	0.68729556	0.574889	4.559737	26.32625
BAL24-T0-UL-A_186-196	191	6.161974	76.12074	0.46638166	0.7563173	13.586647	27.01829
BAL24-T0-L-A-VIRBA_1-11	6	4.272079	97.13686	0.46839374	0.7314854	14.242161	27.09811
BAL24-T0-L-A-VIRBA_37-42	39.5	160.170444	896.76079	0.05124576	0.9561071	30.805571	29.51639
BAL24-T0-L-A-VIRBA_91-96	93.5	9.571823	129.87722	0.24752499	0.8450576	20.287753	28.04895
BAL24-T0-L-A-VIRBA_127-137	132	84.069858	616.48352	0.08480376	0.9336935	30.788197	29.10213
BAL24-T0-L-A-VIRBA_193-198	195.5	1.604963	94.76516	0.14178045	0.8806197	9.625946	29.00176
BAL24-T0-L-A-VIRBA_231-241	236	15.756902	564.29554	0.14874058	0.8833486	15.774501	28.7813
BAL24-T0-DLB-A_1-11	6	39.8535716	492.2017	0.148614	0.8808532	14.829638	28.8919
BAL24-T0-DLB-A_51-56	53.5	2.6537584	82.10706	0.3525095	0.7291223	10.289443	28.21858
BAL24-T0-DLB-A_96-106	101	2.2281591	64.77463	0.5904245	0.6415702	10.55633	26.66692
BAL24-T0-DLB-A_147-152	149.5	9.9973378	338.43475	0.7355769	0.592202	8.316762	26.28606
BAL24-T0-DLB-A_199-209	204	0.4320802	30.50125	0.3901185	0.6958903	4.046167	27.68644
BAL24-T0-M1-PUSH_5-10	7.5	3.995972	146.1908	0.3679285	0.7351091	9.934087	27.56853
BAL24-T0-M1-PUSH_23-28	25.5	8.030239	413.4004	0.4511478	0.7026432	4.315528	27.10998
BAL24-T0-M1-PUSH_42-47	44.5	3.084785	269.8739	0.4696408	0.6834221	7.009157	27.03258

Table A. 5. Raw data of the radiocarbon ages.

Prefix	mean depth [cm b.s.l.]	^{14}C age mean [cal. ka BP]	^{14}C age min. [cal. ka BP]	^{14}C age max. [cal. ka BP]	range [ka BP]
BAL24-T0-UL-A_20-22	21	0.086	0.032	0.14	0.108
BAL24-T0-UL-A_61-63	62	30.2915	30.175	30.408	0.233
BAL24-T0-UL-A_100-103	101.5	34.385	34.198	34.572	0.374
BAL24-T0-UL-A_142-150	146	34.4765	34.29	34.663	0.373
BAL24-T0-UL-A_196-202	199	32.7775	32.269	33.286	1.017
BAL24-T0-L-A-VIRBA_34-37	35.5	0.2975	0.287	0.308	0.021
BAL24-T0-L-A-VIRBA_96-98	97	0.3965	0.36	0.433	0.073
BAL24-T0-L-A-VIRBA_137-140	138.5	0.4805	0.458	0.503	0.045
BAL24-T0-L-A-VIRBA_198-201	199.5	2.27	2.24	2.3	0.06
BAL24-T0-L-A-VIRBA_241-244	242.5	11.9155	11.829	12.002	0.173
BAL24-T0-L-A-VIRBA_307-311	309	1.5335	1.517	1.55	0.033
BAL24-T0-DLB-A_11-13	12	1.1155	1.054	1.177	0.123
BAL24-T0-DLB-A_56-58	57	50	50	50	0
BAL24-T0-DLB-A_106-108	107	50	50	50	0
BAL24-T0-DLB-A_152-154	153	50	50	50	0

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Prefix	mean depth [cm b.s.l.]	¹⁴ C age mean [cal. ka BP]	¹⁴ C age min. [cal. ka BP]	¹⁴ C age max. [cal. ka BP]	range [ka BP]
BAL24-T0-DLB-A_209-214	211.5	50	50	50	0
BAL24-T0-M1-PUSH_10-13	11.5	0.8	0.684	0.916	0.232
BAL24-T0-M1-PUSH_52-55	53.5	28.221	27.678	28.764	1.086

Table A. 6. Biomarker statistics for the four cores.

Core	Parameter	n.valid	min	med	max	mean	sd
DLB	ACL C ₂₃₋₃₃ [-]	5	26.29	27.69	28.89	27.55	1.08
	CPI C ₂₃₋₃₃ [-]	5	4.05	10.29	14.83	9.61	3.91
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gSed ⁻¹]	5	0.43	2.65	39.85	11.03	16.52
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gTOC ⁻¹]	5	30.50	82.11	492.20	201.60	203.38
	P _{aq} C ₂₃₋₃₃ [-]	5	0.15	0.39	0.74	0.44	0.23
	P _{wax} C ₂₃₋₃₃ [-]	5	0.59	0.70	0.88	0.71	0.11
L	ACL C ₂₃₋₃₃ [-]	7	26.72	28.78	29.52	28.32	1.07
	CPI C ₂₃₋₃₃ [-]	7	4.17	15.77	30.81	17.96	10.11
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gSed ⁻¹]	7	1.60	9.57	160.17	39.74	60.60
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gTOC ⁻¹]	7	94.77	177.70	896.76	368.15	321.66
	P _{aq} C ₂₃₋₃₃ [-]	7	0.05	0.15	0.56	0.24	0.20
	P _{wax} C ₂₃₋₃₃ [-]	7	0.63	0.88	0.96	0.84	0.12
M	ACL C ₂₃₋₃₃ [-]	3	27.03	27.11	27.57	27.24	0.29
	CPI C ₂₃₋₃₃ [-]	3	4.32	7.01	9.93	7.09	2.81
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gSed ⁻¹]	3	3.08	4.00	8.03	5.04	2.63
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gTOC ⁻¹]	3	146.19	269.87	413.40	276.49	133.73
	P _{aq} C ₂₃₋₃₃ [-]	3	0.37	0.45	0.47	0.43	0.05
	P _{wax} C ₂₃₋₃₃ [-]	3	0.68	0.70	0.74	0.71	0.03
UL	ACL C ₂₃₋₃₃ [-]	5	26.33	27.02	30.19	27.79	1.60
	CPI C ₂₃₋₃₃ [-]	5	4.56	8.08	76.88	22.01	30.85
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gSed ⁻¹]	5	2.36	6.16	1115.15	227.80	496.06
	Long-chain alkane conc. C ₂₁₋₃₅ [µg gTOC ⁻¹]	5	67.33	97.33	3439.71	855.63	1461.85
	P _{aq} C ₂₃₋₃₃ [-]	5	0.02	0.47	0.69	0.38	0.27
	P _{wax} C ₂₃₋₃₃ [-]	5	0.57	0.76	0.98	0.76	0.15

Table A. 7. Sedimentological statistics for the three permafrost layers.

Permafrost layer	Parameter	n.valid	min	med	max	mean	sd
active layer	Absolute ice / water content [wt%]	8	26.96	72.83	89.78	64.60	23.95
	Clay [vol %]	1	4.09	4.09	4.09	4.09	NA
	Mean grain size [µm]	1	22.24	22.24	22.24	22.24	NA
	Sand acc. [vol %]	1	100.00	100.00	100.00	100.00	NA
	Sand [vol %]	1	14.14	14.14	14.14	14.14	NA
	Silt acc. [vol %]	1	85.86	85.86	85.86	85.86	NA
	Silt [vol %]	1	81.77	81.77	81.77	81.77	NA
perennial frozen	Absolute ice / water content [wt%]	60	27.07	40.94	99.92	46.01	15.84
	Clay [vol %]	60	2.48	4.20	6.07	4.21	0.77
	Mean grain size [µm]	60	18.22	23.15	32.36	24.21	3.33
	Sand acc. [vol %]	60	99.72	100.00	100.00	100.00	0.04
	Sand [vol %]	60	12.19	17.21	24.76	17.80	2.80
	Silt acc. [vol %]	60	75.24	82.79	87.81	82.19	2.79
	Silt [vol %]	60	72.76	78.27	82.28	77.98	2.33
talik	Absolute ice / water content [wt%]	66	22.29	28.58	67.48	35.16	12.61
	Clay [vol %]	60	1.60	4.29	9.29	4.32	1.55
	Mean grain size [µm]	60	10.43	21.74	61.17	24.52	9.69
	Sand acc. [vol %]	60	99.98	100.00	100.00	100.00	0.00
	Sand [vol %]	60	4.31	14.15	51.29	17.93	9.68
	Silt acc. [vol %]	60	48.71	85.85	95.69	82.07	9.68
	Silt [vol %]	60	47.10	81.27	88.24	77.75	8.46

Table A. 8. Biogeochemical statistics for the three permafrost layers.

Permafrost layer	Parameter	n.valid	min	med	max	mean	sd
active layer	TN [wt%]	8	0.11	1.13	2.01	1.07	0.74
	TOC [wt%]	8	1.85	25.40	44.42	23.36	17.93
	C/N [-]	8	11.68	16.96	42.02	21.15	11.23
	δ ¹³ C [‰ vs. VPDB]	5	-36.50	-28.32	-27.29	-29.96	3.76
perennial frozen	TN [wt%]	70	0.01	0.28	0.82	0.30	0.14
	TOC [wt%]	70	1.19	3.45	16.28	3.85	2.15
	C/N [-]	69	9.07	12.16	19.97	12.55	1.64

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Permafrost layer	Parameter	n.valid	min	med	max	mean	sd
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	63	-27.62	-25.97	-24.45	-25.84	0.65
talik	TN [wt%]	77	0.01	0.16	1.28	0.33	0.34
	TOC [wt%]	77	0.60	2.22	24.40	5.37	6.12
	C/N [-]	65	9.26	13.61	21.08	14.06	2.80
	$\delta^{13}\text{C}$ [‰ vs. VPDB]	67	-28.61	-26.65	-24.52	-26.58	0.83

Table A. 9. Hydrochemical statistics for the three permafrost layers.

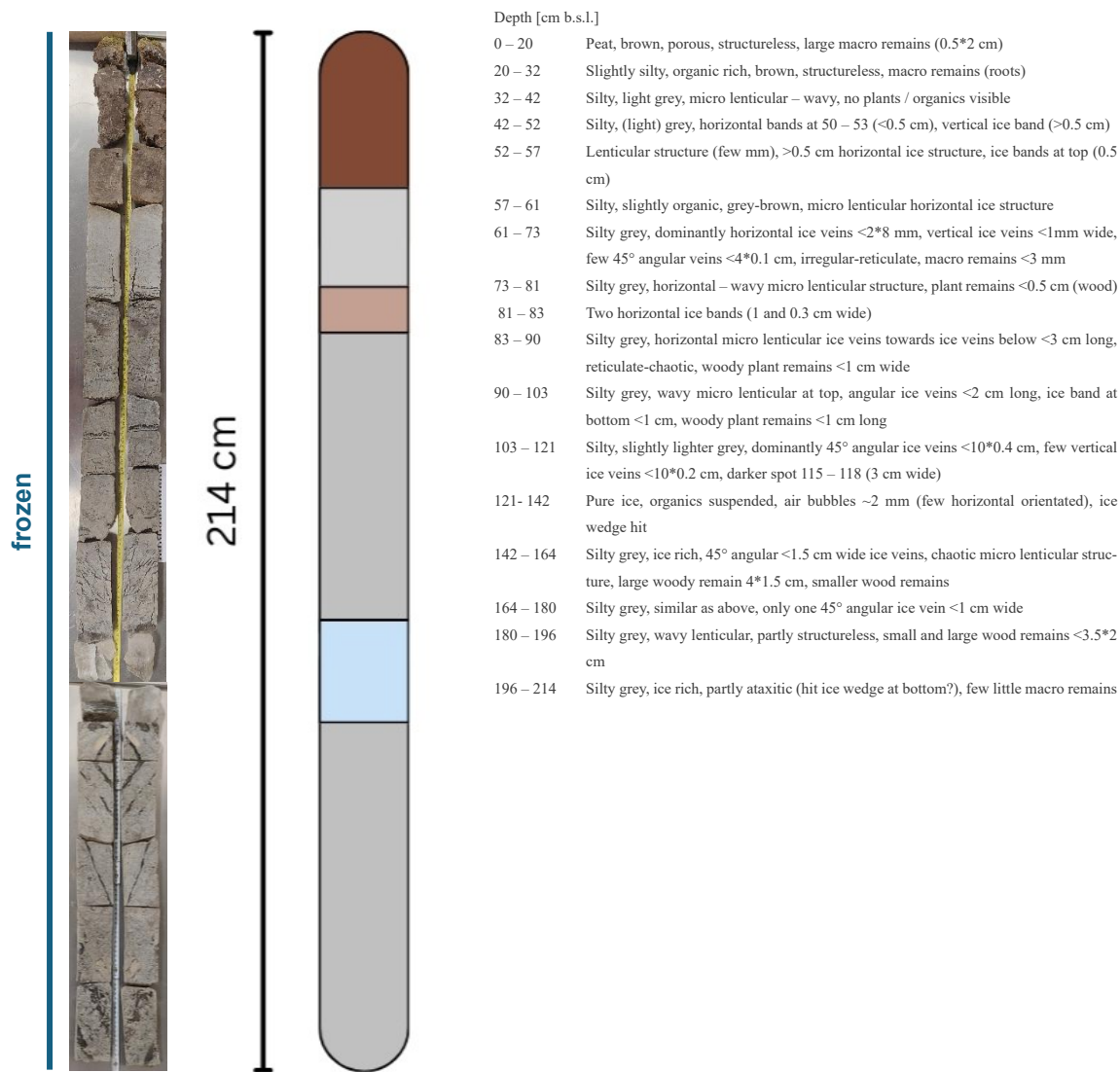
Permafrost layer	Parameter	n.valid	min	med	max	mean	sd
active layer	DOC [mg L ⁻¹]	1	42.05	42.05	42.05	42.05	NA
	EC [$\mu\text{S cm}^{-1}$]	1	44.62	44.62	44.62	44.62	NA
	Mean d excess [-]	1	8.37	8.37	8.37	8.37	NA
	Mean $\delta^{18}\text{O}$ [‰ vs. SMOW]	1	-15.08	-15.08	-15.08	-15.08	NA
	Mean δD [‰ vs. SMOW]	1	-112.28	-112.28	-112.28	-112.28	NA
	pH [-]	1	4.87	4.87	4.87	4.87	NA
perennial frozen	DOC [mg L ⁻¹]	30	5.78	199.08	1707.00	324.27	418.22
	EC [$\mu\text{S cm}^{-1}$]	30	43.41	870.50	2379.00	819.49	576.28
	Mean d excess [-]	25	-3.06	4.85	14.60	5.39	4.03
	Mean $\delta^{18}\text{O}$ [‰ vs. SMOW]	25	-20.36	-18.89	-14.25	-17.88	1.99
	Mean δD [‰ vs. SMOW]	25	-160.21	-142.65	-112.15	-137.62	14.49
	pH [-]	30	5.68	7.66	8.41	7.37	0.73
talik	DOC [mg L ⁻¹]	26	41.38	147.55	312.25	149.36	54.81
	EC [$\mu\text{S cm}^{-1}$]	26	91.00	701.80	20390.00	2761.55	6007.52
	Mean d excess [-]	20	-1.17	0.10	6.95	0.87	2.22
	Mean $\delta^{18}\text{O}$ [‰ vs. SMOW]	20	-18.95	-18.31	-13.14	-17.54	1.92
	Mean δD [‰ vs. SMOW]	20	-152.39	-146.79	-101.57	-139.48	17.02
	pH [-]	26	7.22	7.75	8.22	7.76	0.24

Table A. 10. Biomarker statistics for the three permafrost layers.

Permafrost layer	Parameter	n.valid	min	med	max	mean	sd
active layer	ACL C ₂₃₋₃₃ [-]	3	28.65	28.89	30.19	29.25	0.83
	CPI C ₂₃₋₃₃ [-]	3	6.93	14.83	76.88	32.88	38.31
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gSed}^{-1}$]	3	11.05	39.85	1115.15	388.69	629.30
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gTOC}^{-1}$]	3	492.20	597.66	3439.71	1509.86	1672.13
	P _{aq} C ₂₃₋₃₃ [-]	3	0.02	0.15	0.20	0.12	0.09
	P _{wax} C ₂₃₋₃₃ [-]	3	0.83	0.88	0.98	0.90	0.08
perennial frozen	ACL C ₂₃₋₃₃ [-]	6	26.29	26.72	28.22	26.88	0.71
	CPI C ₂₃₋₃₃ [-]	6	4.56	9.30	13.59	9.23	3.03
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gSed}^{-1}$]	6	2.23	3.46	10.00	4.61	3.04
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gTOC}^{-1}$]	6	64.77	79.11	338.43	121.02	107.15
	P _{aq} C ₂₃₋₃₃ [-]	6	0.35	0.57	0.74	0.56	0.14
	P _{wax} C ₂₃₋₃₃ [-]	6	0.57	0.66	0.76	0.66	0.07
talik	ACL C ₂₃₋₃₃ [-]	11	26.72	27.69	29.52	27.97	0.98
	CPI C ₂₃₋₃₃ [-]	11	4.05	9.93	30.81	13.73	9.90
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gSed}^{-1}$]	11	0.43	4.27	160.17	26.70	50.34
	Long-chain alkane conc. C ₂₁₋₃₅ [$\mu\text{g gTOC}^{-1}$]	11	30.50	177.70	896.76	312.45	275.98
	P _{aq} C ₂₃₋₃₃ [-]	11	0.05	0.37	0.56	0.31	0.18
	P _{wax} C ₂₃₋₃₃ [-]	11	0.63	0.74	0.96	0.79	0.11

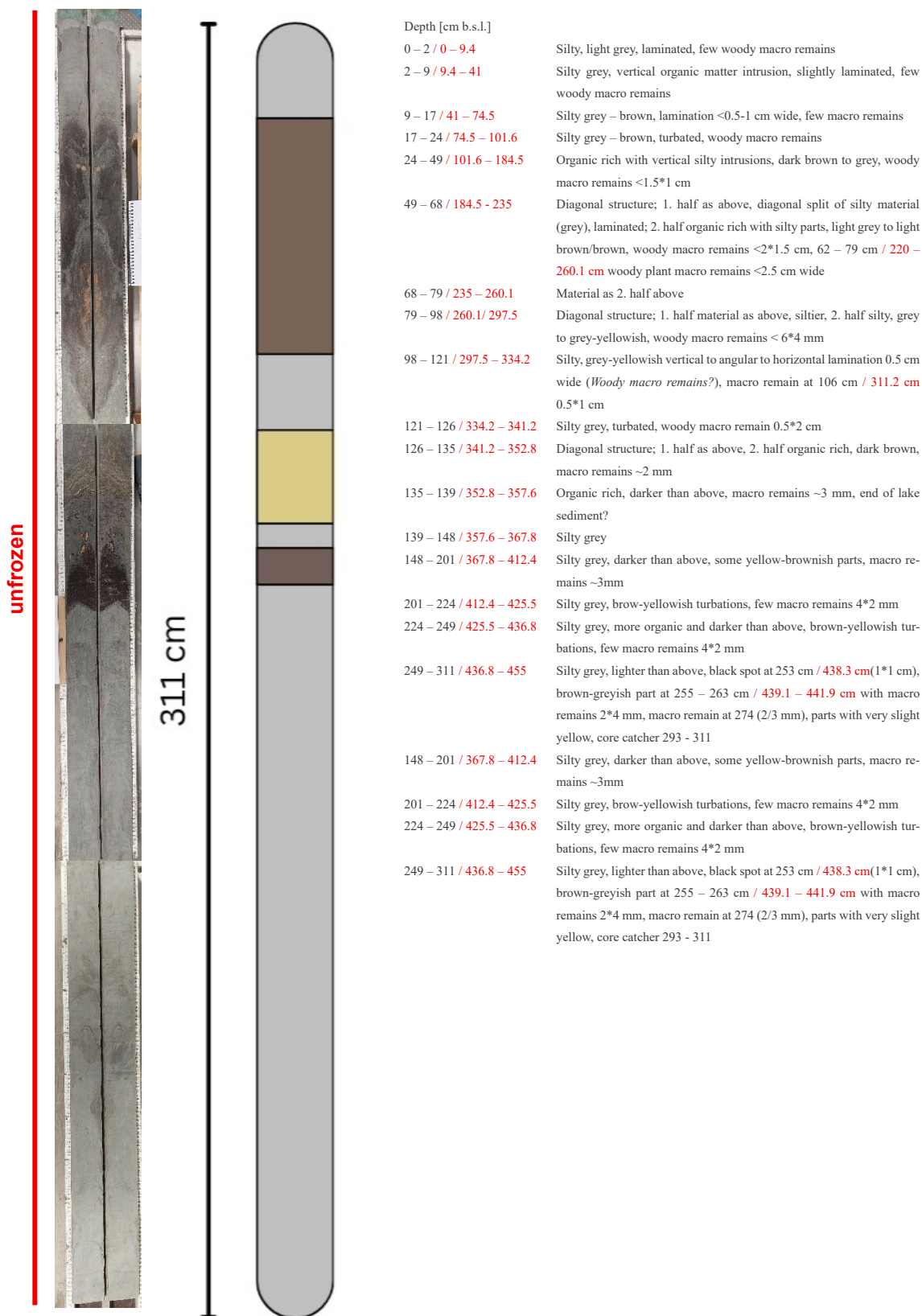
Appendix

Figure A. 1. Core BAL24-T0-UL with the sedimentological and cryolithological core description.



Appendix

Figure A. 2. Core BAL24-T0-L with the sedimentological and cryolithological core description. Corrected depths in red.



Appendix

Figure A. 3. Core BAL24-T0-DLB with the sedimentological and cryolithological core description.

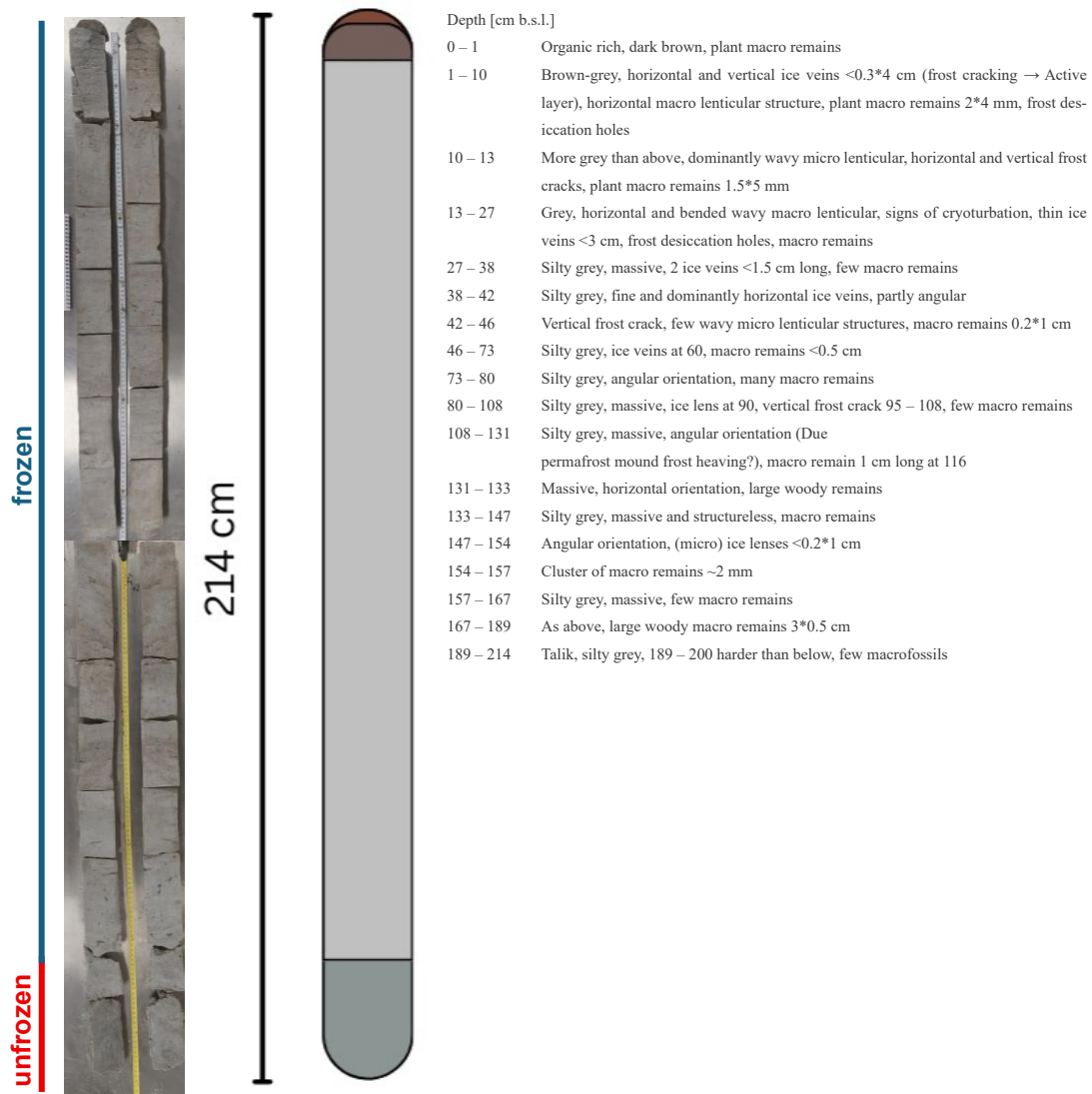
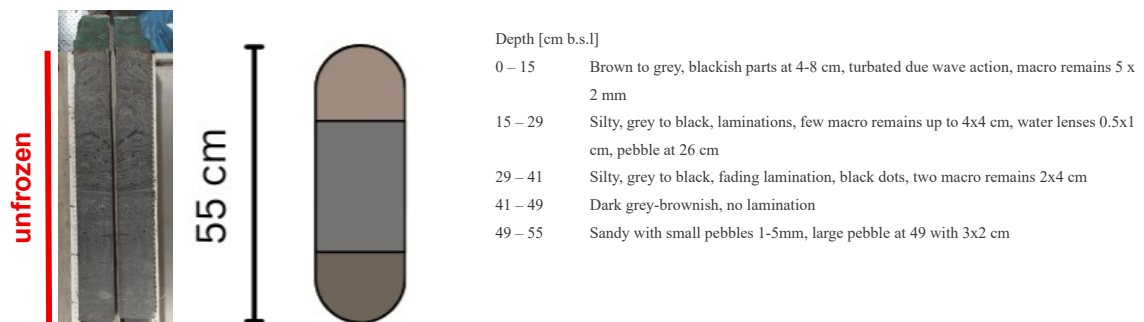


Figure A. 4. Core BAL24-T0-M with the sedimentological and cryolithological core description.



Appendix

Figure A. 5. Example of a gas chromatogram and mass spectra in Xcalibur (Thermo Fischer).

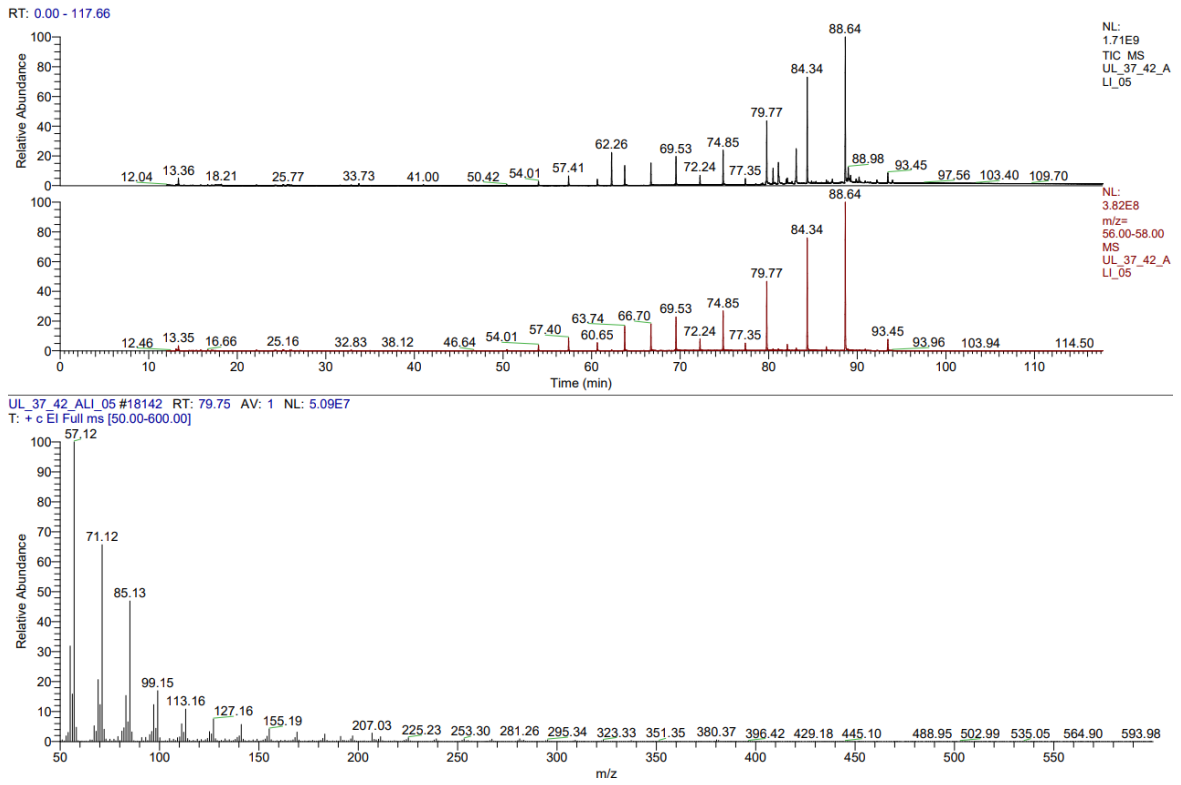
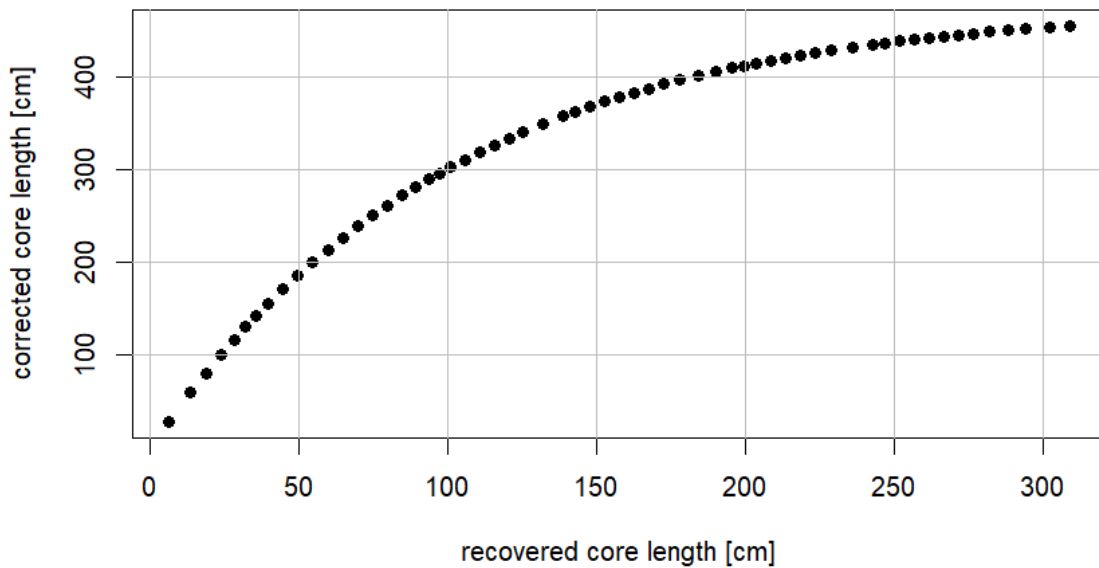
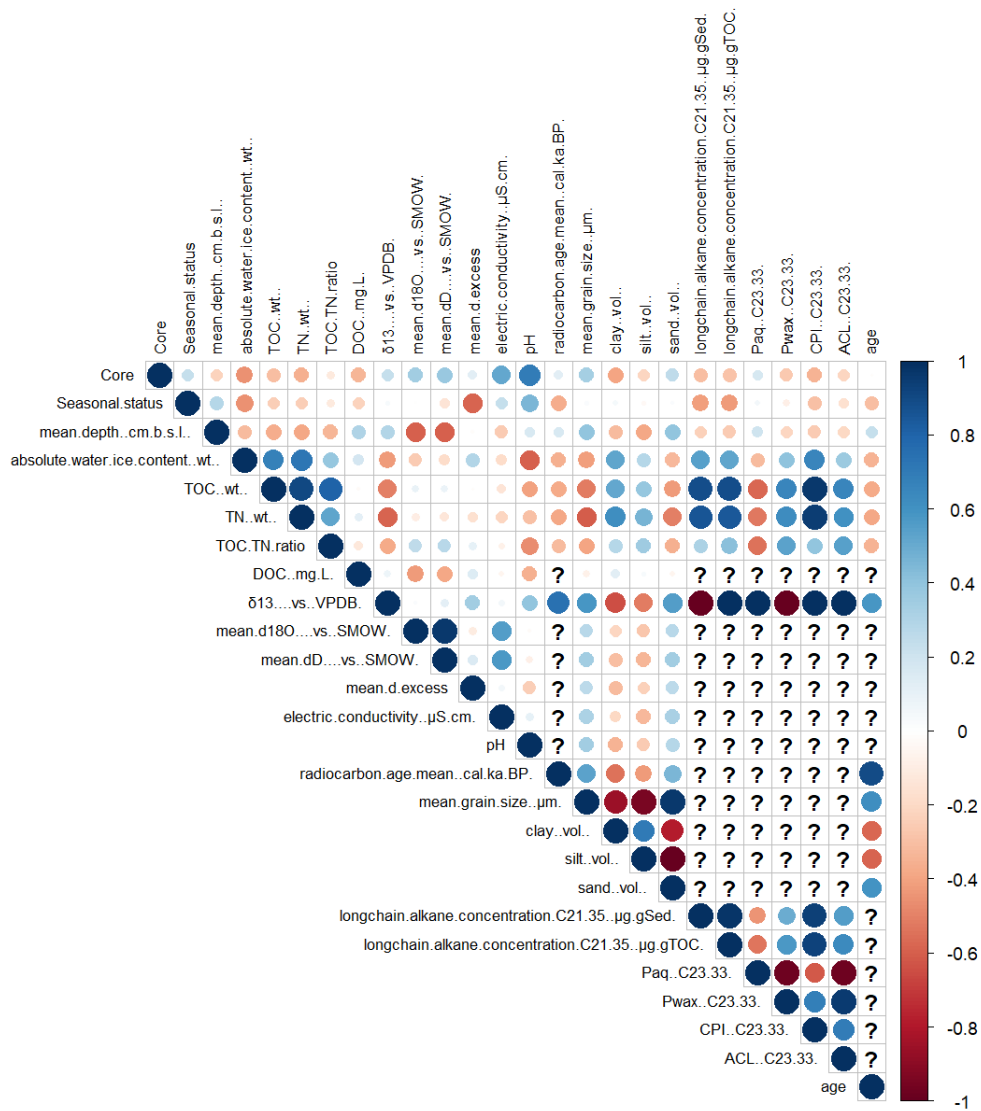


Figure A. 6. Result of the exponential compaction correction for the core length of BAL24-T0-L.



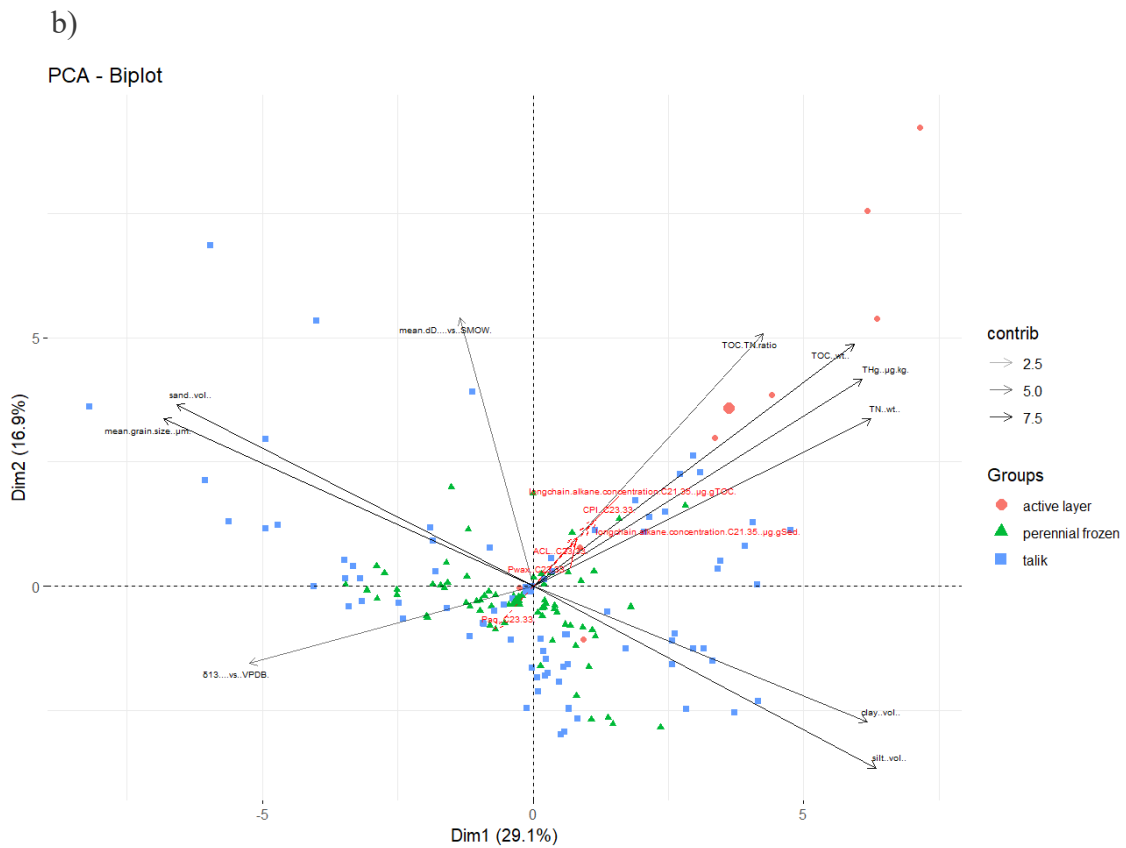
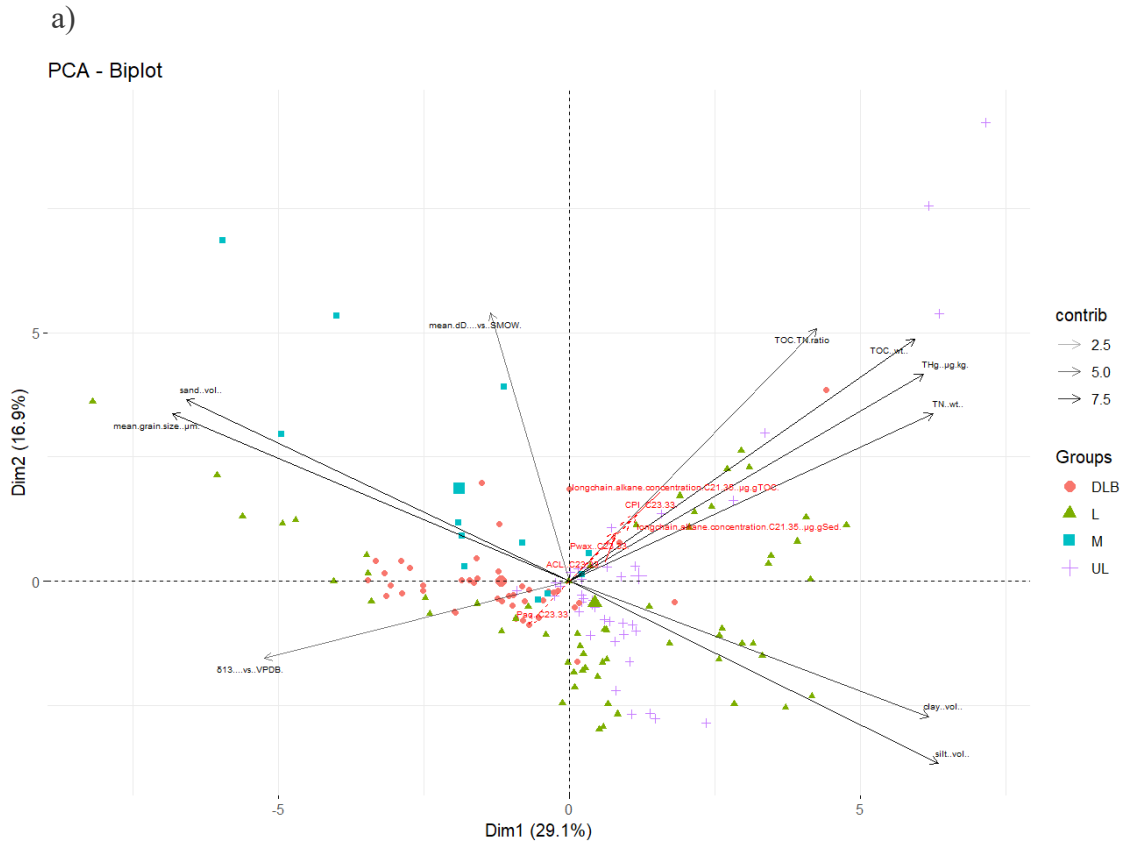
Appendix

Figure A. 7. Correlation matrix of all parameters.



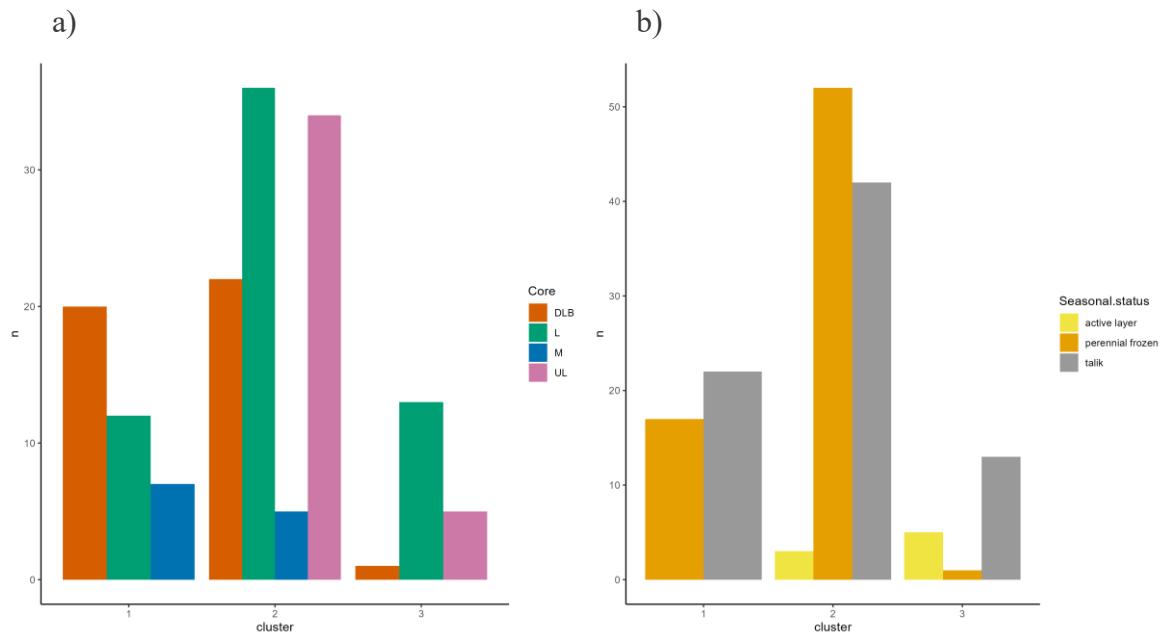
Appendix

Figure A. 8. Biplot of the PCA-Analysis with biomarker proxies as supplementary variables, colored after (a) cores and (b) permafrost layer.



Appendix

Figure A. 9. Contribution of (a) cores and (b) permafrost layers to the three different clusters resulting from the HCPC.



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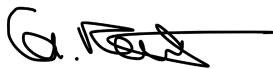
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Statement of Originality

Statement of Originality

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Signature (Carolin Luise Frauhammer)