

A new approach on the oxygen isotope micro analysis of diatom silica with a laser-fluorination based mass spectrometry unit

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Introduction

The analysis of oxygen isotopes from diatom silica in sediment cores has reached importance for reconstructing the paleoclimate and is especially valuable in non-carbonate lakes of cold regions, where no other bioindicators such as ostracods and foraminifera are available. A new approach for samples in sub-mg range has been developed to provide a better chronological resolution and to expand the method to periods where less biogenic silica is available. Sample material from Lake El'gygytyn (Center: N 67°30', E 172°5', core LZ 1024) will be analysed and a $\delta^{18}\text{O}$ curve of the last 280.000 years will be generated to add a strong climate proxy to the various analysis performed so far. The Lake lies inside a meteorite impact crater formed app. 3.6 million years ago and hence offers a unique option to fill the spatial gap of locations in the Arctic where paleoclimate reconstructions are rare.

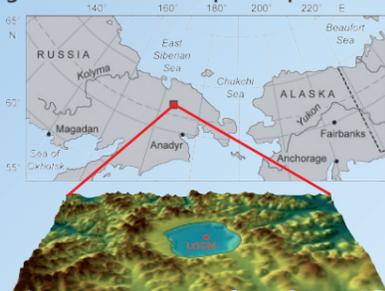


Fig. 1: Geography and Digital area model of the Surrounding area of Lake El'gygytyn (Courtesy: C. Kopsch).

Diatom extraction

The diatoms are extracted from sediment samples with various preparation steps. At first organic material and carbonates are removed with H_2O_2 and HCl . Second, after rinsing out the acid, the sample is dispersed with Na -hexametaphosphat. Third, sieves with mesh sizes of $5\mu\text{m}$, $10\mu\text{m}$, $20\mu\text{m}$, $32\mu\text{m}$ and $125\mu\text{m}$ are used to receive the different sized fractions. For Lake El'gygytyn the *cyclotella-ocellata* complex can be found mostly in the 5-10 μm fraction whereas the species of *Pliocenicus costatus* is predominant in the 10 μm to 20 μm fraction. Finally a heavy liquid separation is performed in a centrifuge by adding 2.1 sg Wolfram-polytungstate solution to the different fractions of the sample. A minimum of $\sim 700\mu\text{g}$ fine material from 5 g of wet sample is obtained after rinsing out the heavy liquid in another sieving process. A control slide is prepared to determine the degree of purity.

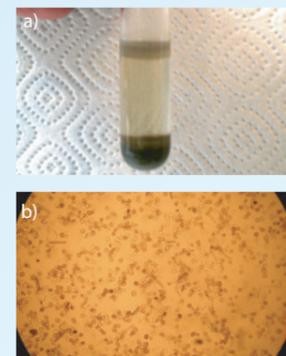


Fig. 2: Centrifuge tube after the heavy liquid separation: Diatoms float on top. (a) Control slide of the 10-20 μm fraction (b)

Melting powder to beads

Between 0.7 and 2 mg of standard (NBS 28, Campolungo) or diatom material is pressed into the holes of a Platinum plate. The prepared plate is arranged on a specially designed stand that fits into the conditioning chamber where the samples will be melted into beads. The pressure in the chamber has to be under 10^{-3}mbar before the CO_2 laser can be operated. It is adequate to direct the defocused beam over the sample at first with an increasing power of 0.5 W, 0.6 W and 0.7 W (Fig.3;1). At each stage the move should follow a spiral starting from the center point to

equally warm up the powder and evade parts spreading apart or a potential explosion of the sample. A light glow can be seen when 0.8 W are applied (Fig.3;2). Then, the laser power is increased fast to 5-6 W with this power stable for about 5 to 10 seconds (Fig.3;5). The power is then reduced to zero within 10 seconds. A bead is formed (Fig.3;8) with a calculated mass loss between 4% (NBS 28) lacking any water and 20% (Radiolarians/Diatoms) due to chemically-bound water in the sample.

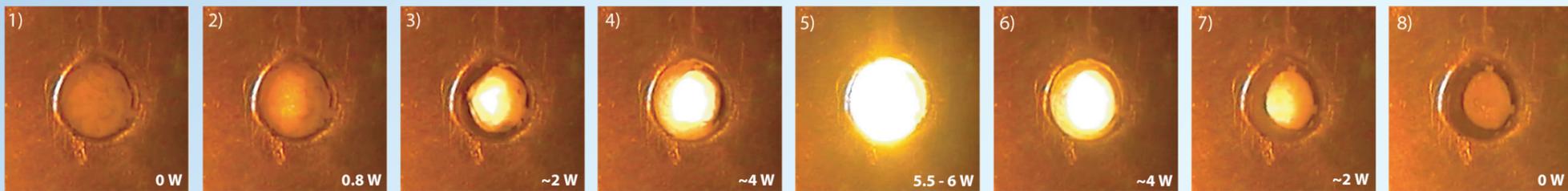


Fig. 3: The process of melting the powdered samples to beads takes about 5 to 10 seconds with a recommended laser power between 5.5W and 6W. Picture 1 to 8 show the different stages of the sample in this process: Picture 1 shows the powdered sample where as at Picture 8 the finished bead can be seen.

The process

The extracted diatom sample is melted into a bead (see above) and then heated with a CO_2 laser under BrF_5 atmosphere to release the O_2 . The non-oxygen gas components are trapped in a -150°C cold trap, whereas oxygen passes on to the molecular sieve cooled with liquid nitrogen. It is then transferred to the mass spectrometer and compared with reference O_2 calibrated with several standards. By measuring the isotope ratio difference between sample and reference, the $\delta^{18}\text{O}$ value of the sample vs. VSMOW is determined.

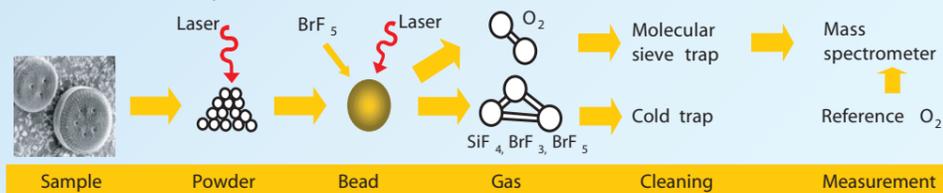


Fig. 4: Principle of the analysis from sample to measurement. After ground to powder and melted to a bead the sample is heated with a CO_2 laser under BrF_5 atmosphere to release the O_2 . The non-oxygen gas components are trapped in cold trap, Oxygen passes to the molecular sieve cooled with liquid nitrogen and is transferred to the mass spectrometer for isotope measurement.

Rooms & Safety

Due to high safety regulation principles for toxic gases such as BrF_5 the instrument is arranged in two rooms: The reaction room (Fig. 5a) and the control room (Fig. 5b). The fluorination line is installed in the reaction room under a hood containing conditioning chamber and reaction chamber, laser, gas bottles, reagent reservoirs, pumps and cold traps. Additionally the chambers and the laser are located in a specially designed safety box to assure maximum security. Mass spectrometer and control unit are connected through a void in the wall and set up in the control room.

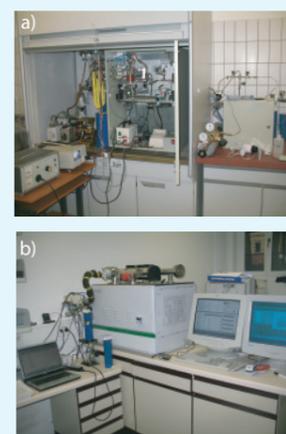


Fig. 5: The reaction room (a) is separated from the control room (b) and contains the fluorination line. The mass spectrometer and the work station are located in the control room.

Connections & Valve Chart

The whole system is assembled with 1/4 and 3/8 inch stainless steel tubing connected with fittings from Swagelok. Pneumatic valves can be switched on and off automatically (see „The Software“ left) where as for more critical spots (release of BrF_5 , etc.) manual valves are included. Pressure gauges are installed to secure an airtight system at all times. Temperature probes guarantee stable conditions in cold trap and molecular sieve.

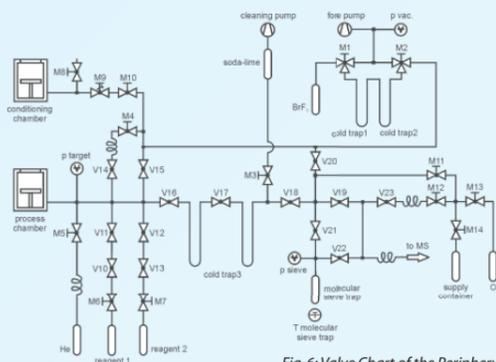


Fig. 6: Valve Chart of the Periphery

The Software

A Window-based software for a remotely-controlled fluorination line and mass spectrometer was developed. It allows the control of the mass spectrometer as well as of the pneumatic valves. A ‚Drag and drop‘ menu to compile a measurement procedure is integrated to measure the samples automatically. The measured data are transferred to an Excel file where the $\delta^{18}\text{O}$ values of the sample are determined. All parameters and the used measurement procedure are stored in the Excel file. A video camera is connected to a second computer to survey and record the process in the reaction chamber (see extracted images above).

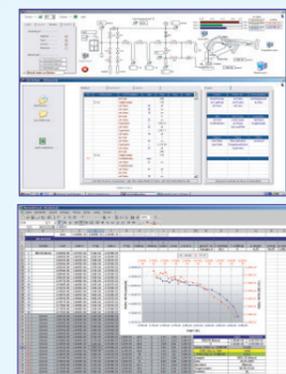


Fig. 7: Screenshots of the software windows (a) and the created Excel File (b). The software shows the clickable valve chart (for switching the valves on/off) and updated temperature and pressure information on top. On the bottom the measurement procedure window is opened.