

CRUISE REPORT: Repeat Hydrography on Line PR6:
WOCE Cruise No. 18DD9618/1

Chief Scientist: Frank Whitney
Ship: John P. Tully
Ports of Call: none
Cruise Dates: August 12 to September 6, 1996
Expedition Designation: 18DD9618/1

Cruise Narrative

Our repeat hydrography section was a joint program with Canadian JGOFS.

A full CTD survey along Line PR6 to a depth of 3000 m was completed, with CTD casts at 29 stations and full depth rosette/hydro casts at 5 stations. Water sampling in the upper 100 m was completed at an additional 8 stations. On the return leg, a CTD section along 51 N to 1000 m was supplemented by water sampling in the upper 100 m. Salinity, oxygen and nutrients (NO₃ & NO₂, PO₄ and Si) were analyzed onboard ship.

JGOFS participants collected samples at 5 stations for abundance and activities of bacteria, phytoplankton, micro- and meso-zooplankton and incubated water to measure growth and grazing rates of various groups of plankton. The natural abundance of stable isotopes (¹⁵N and ¹³C) in suspended particulate matter was also determined at these stations.

Cruise Summary Information

Cruise track

Line PR6 starts at the mouth of Juan de Fuca Strait on the west coast of Canada, and heads almost due west for 900 n mi. The terminal station is PRS1, formerly designated Ocean Weather Station Papa (50 N, 145 W). Our return leg headed north from PRS1 to 51 N and ran directly east to the coast.

Table of Stations by type

Sample type:

No. stations:

Max. depth (dbar):

CTD casts

46

3007 db

Rosette/Hydro casts

33

4319

Loop samples

72

5

Floats and Drifters deployed

A profiling Alace float was recovered near at station R16.
Free drifting sediment traps were deployed at PRS1 for 4 days.

Principal Investigators

Howard Freeland

Ocean circulation

IOS

C.S. Wong

Climate chemistry

IOS

Frank Whitney

WOCE coordinator

IOS

Philip Boyd

JGOFS coordinator

UBC

Goals Achieved

CTD survey of Line PR6 and along 51N.

Successful rosette casts at 5 stations on Line P.

Completion of JGOFS sampling for plankton and productivity measurements.

Problems and Goals not Achieved

none

Cruise Participants & Affiliations

Frank Whitney

Chief scientist, nutrients

IOS

Philip Boyd

phytoplankton

UBC

Edmand Fok

Watch

IOS

Tim Soutar

watch

IOS

Wendy Richardson

Oxygen, watch

IOS

Ron Bellegay

Watch, carbonates

IOS

Doug Anderson

Watch,

IOS

Michael Bentley

Watch, seabirds

contractor

Robert Goldblatt

Zooplankton biomass

UBC

Hugh Maclean

Watch, plankton sampling

UBC

Maureen Soon

particulate 13C & 15N

UBC

Nelson Sherry

bacteria enumeration

UBC

Todd Mudge

MULVFS pump

U.Vic

Robert Schultz

MULVFS pump

U.Vic

Delphine Thibault

Zooplankton excretion

Rimouski U.

Ken Crocker

Zooplankton grazing

Memorial U.

Paul Matthews

Micro-zooplankton

Memorial U.

Julie Granger
Fe uptake
McGill U.
Maite Maldonado
bacteria respiration & production
McGill U.

Measurement Techniques and Calibrations

CTD profiles

A Guildline 8705 CTD (SN 58483) coupled with a transmissometer was lowered to a maximum of 3000 m on casts where our rosette was not used.

Water sampling

A rosette holding a Guildline 8737 CTD and 23-10 L polycarbonate Niskin bottles was used for hydro casts. Go-Flo bottles clamped on Kevlar hydro line were used to collect clean water for plankton studies.

Each rosette/hydro station consisted of two casts - a down cast and an up cast. The CTD profile is taken from the down cast. Water samplers were tripped on the up cast and CTD pressure (dbar) and CTD temperature (uncorrected) recorded from this upcast.

At each station, samples for surface chlorophyll, salinity and nutrients were collected from the ship's sea water loop which pumps water from about 5 m continuously into the laboratory.

Salinity

Samples were collected in glass bottles and analyzed onboard ship using a Guildline Model 8410 Portasal. The Portasal was standardized daily with IAPSO standard sea water (batch xxx).

Oxygen

An automated titration system (Brinkman Dosimat and Fiber Optic Probe Colorimeter) using the micro-Winkler method (Carpenter, 1965), titrated samples to the iodine end-point. Standards were prepared as outlined in WOCE Report 73/91.

Nutrients

Samples from hydro casts were collected in polystyrene tubes and refrigerated for a maximum of 12 h before being analyzed. Loop samples (USW) were stored up to 2 days at 4°C before being analyzed. NO₃+NO₂, PO₄ and Si were analyzed using a Technicon Autoanalyzer.

NO₃+NO₂ samples were reduced with Cd/Cu, then complexed with sulfanilamide and N-Naphthylethylene-diamine to form an azo dye (Technicon Method No. 158-71W/B). PO₄ produces a molybdenum blue complex in presence of acidic molybdate and ascorbic acid (Technicon Method No. 155-71W). Dissolved Si also forms a molybdenum blue complex and oxalic acid removes PO₄ interference (Technicon Method 186-72W).

Concentrated standards were freshly prepared the week before the cruise from oven dried reagents. Working standards were made every 1 to 2 days by diluting 1 to 6 mL of various stock solutions to 250 mL with 3.2% NaCl (w/v in double run Milli-Q water).

Table. Laboratory temperatures for nutrients.

Date
Temp (C)
Date
Temp (C)
Aug 14
30.3 to 30.9
Aug 24
29.1 to 29.3
Aug 16

29.7 to 30.4
Aug 26
27.2 to 28.9
Aug 18
29.3 to 29.6
Aug 27
26.3 to 27.0
Aug 19
29.7 to 30.0
Aug 29
27.0 to 28.5
Aug 21
30.1 to 31.1
Aug 30
27.6 to 28.7
Aug 22
29.3 to 30.1
Sep 1
29.1
Aug 23
28.3 to 30.0
Sep 2
26.2 to 28.2

Sep 3
26.5 to 27.6

TCO₂, 13C and Alkalinity - samples were collected at stations P4, P12, P16 and PRS1, fixed with HgCl₂ and refrigerated.

O18/O16 - samples were collected in 60 mL polyethylene bottles and refrigerated.

JGOFS sampling - Go-flo bottles were used to collect water for POC/N, DOC/N, chlorophyll, nano- and micro-plankton and incubation experiments. Deck incubations were conducted to measure growth rates of bacteria, phytoplankton and micro-zooplankton.

References

Carpenter, J.H. 1965. The Chesapeake Bay Institute technique for the Winkler dissolved oxygen method. *Limnol. Oceanogr.*, 10: 141-143.

Technicon Industrial Method No. 155-71W. 1973. Orthophosphate in water and seawater.

Technicon Industrial Method No. 158-71W/A. 1977. Nitrate and nitrite in water and seawater.

Technicon Industrial Method No. 186-72W/B. 1977. Silicates in water and seawater.

WOCE Report 73/91. 1991. A comparison of methods for the determination of dissolved oxygen in seawater.