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Global Rates of Marine Sulfate Reduction and Implications for Sub–Sea-Floor Metabolic Activities

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Sulfate reduction is a globally important yet poorly quantified redox process in marine sediments. We developed an artificial neural network trained with 199 sulfate profiles, constrained with geomorphological and geochemical maps to estimate global sulfate reduction rate distributions. Globally, 11.3 Tmol sulfate are reduced yearly, ~15% of previous estimates, accounting for the oxidation of 12-29% of the organic carbon flux to the sea floor. Combined with global cell distributions in marine sediments, these results indicate a strong contrast in sub-sea-floor prokaryote habitats: in continental margins global cell numbers in sulfate-depleted sediment exceed those in the overlying sulfate-bearing sediment by an order of magnitude, whereas in the abyss most life occurs in oxic and/or sulfate-reducing sediments.

Sulfate reduction is a ubiquitous microbial process in oceanic sediments and an important pathway for carbon oxidation and redox cycling. Nevertheless, the currently estimated global sulfate reduction rate (SRR) of, 75 Tmol sulfate $y^{-1}(l)$ is based on coarse spatial averaging and is not consistent with the up-to-date assessment of the global organic matter flux to marine sediments (2) of 79 to 192 Tmol C y^{-1} (3, 4). Net sulfate follows a 2-C-to-1-S stoichiometric reduction ratio $C_6H_{12}O_6 + 3SO_4^{2-} \rightarrow 3S^{2-} + 6CO_2 + 6H_2O$, e.g., (5)].

Therefore, the current estimates for the rate of subsurface sulfate reduction and for the organic carbon flux to the sediment suggest that either insufficient organic carbon reaches the sediment to account for sulfate reduction or that most (78%) organic matter is channeled toward sulfate reduction. Nevertheless, the organic carbon reaching the sediment must also foment other prominent redox reactions such as carbon respiration (4, 6), and moreover, a sizable portion of sedimentary organic matter successfully survives early diagenesis and is buried (7). This discrepancy in global geochemical cycles gives impetus for an amended global view on sulfate reduction which can be merged with recently revised global prokaryotic abundances to properly assess the activity of sulfate reducing microorganisms at a global scale (5, 8-11).

We used currently available sulfate concentration profiles from multiple scientific ocean drilling programs (12) to estimate global net SRRs (Fig. 1A). These profiles were best described by assuming that sulfate concentrations exponentially decrease with depth. A total of 199 sulfate profiles (Fig. 1B) with a mean error square value $< 4 \text{ mM}^2$ based on a least-squares regression were selected for the global SRR analysis. We then used depth-decay constants (b) extracted from these profiles to

train an artificial neural network (ANN) using high-resolution (1x1 degree) satellite observations and water column chemistry maps (e.g., surface water chlorophyll A, particulate organic carbon, and bottom water O_2) (13, 14) (table S1) (12).

The ANN predicts depth-decay constants ranging 16 orders of magnitude from 5.8 \times 10^{-13} to 3.2 \times 10^{2} m^{-1} while depth-integrated SRRs calculated using a steady state diffusion, advection, and reaction function (eq. S8) (12) ranged from 5.8×10^{-12} to 8.2 mmol cm⁻² y⁻¹. The highest SRRs were predicted in shelf environments and the lowest SRRs were calculated in the nutrient-poor oceanic gyres (Fig. 1A and Table 1). These general trends are corroborated by a previous prediction of global, depth-integrated SRRs derived from, and mainly reflecting, the distributions of primary productivity. They are furthermore consistent with observations in previous studies of global sulfate profiles from oceanic deep drilling programs (5), as well as

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global compilations of radiotracer gross SRR measurements (15). Alt-hough the ANN is trained and validated solely with deep sea drilling data (Fig. 1B) (12), it replicates exponential sulfate depth-decay coeffi-cients from several published short cores (<15 m) (fig. S4) taken below the shelf break and captures deep-sea locations characterized by high regional rates of SR coupled to methane oxidation [e.g., 0.05 - 0.4 mmol $cm^{-2} y^{-1}$ in the Arabian Sea (16)]. From an environment-specific perspective, the ANN predicts a fourth of the previously estimated, area-weighted depth-integrated SRR for shelf sites: 0.097 mmol cm⁻² y⁻¹ (15) vs. 0.0247 mmol cm⁻² y⁻¹ (Ta-ble 1). All average depth-integrated rates determined here are consideraglobal compilations of radiotracer gross SRR measurements (15). Alt-

ble 1). All average depth-integrated rates determined here are considerably lower than previously reported (15), but appear to agree better with regionally averaged sulfate penetration depths. For example, assuming the previous upper continental slope average areal rate of 7.4×10^{-2} mmol cm⁻² y⁻¹ (15), a tortuosity-corrected diffusion coefficient of 120 cm² y⁻¹ and a porosity of 0.8, the average sediment depth where sulfate depletes to 0.3 mM in sediments would be only 35 cm or 1.74 m assuming a linear or an exponential profile, respectively. The calculations here yield an average depth integrated rate for the upper slope of 1.7×10^{-2} mmol $cm^{-2} y^{-1}$. This rate would produce 0.3 mM sulfate concentrations at a depth of 1.2 m or 20 m below sea floor (mbsf) for these same cases, respectively. The main reason for this difference is that the previously compiled global SRRs did not account for large sea-floor surface areas (e.g., ~70% of the continental shelf) consisting of organic-poor relict sands (17), and thus were averaged with a bias toward high-activity, organic-rich sites [cf. (8, 18)]. This and other geochemical and depositional heterogeneities observed in coastal sediments may also explain some of the larger deviations between fitted and ANN-predicted b values for some shallow-water cores (Fig. 1B). For instance, order-ofmagnitude variations in SRRs have been estimated within a single muddy basin [e.g., within 3000 km² in Arkona Basin (*19*)], and likewise for small gassy basins [e.g., 8 km² in Aarhus Bay (*20*)].

The ANN-based global net SRR estimate (11.3 Tmol S y⁻¹) (Table 1) is roughly 15% of previous estimates for gross SRR (*1*). Although calculating global net as opposed to gross SRR could explain this divergence, it is highly unlikely that gross SRRs account for more than 78% of the global organic carbon flux to the sea floor. In contrast, the 11.3 Tmol S y⁻¹ predicted by the ANN is equivalent to 22.6 Tmol C y⁻¹ [assuming a 2-C-to-1-S stoichiometry (*5*, *12*)] or a more realistic 11-29% of the estimated global organic carbon flux to the sea floor.

This substantial diminution in global SRRs inherently affects previous conceptions of global microbial process distributions in sub–seafloor sediments. Subsurface microorganisms largely depend on harvesting energy from the organic matter reaching the sea floor. This amount is minor in comparison to the carbon supplied to seawater prokaryotes via photosynthesis (4.3 Pmol C y^{-1}) (21). In spite of this sharp contrast in carbon availability the marine subsurface total prokaryotic biomass is approximately equal to that of seawater (8).

Coupling the ANN-derived global SRR maps to global sub-sea-floor biomass maps (8) allows for the calculation of potential cell-specific rates, which can further elucidate the activities of sulfate reducing microorganisms across various global sedimentary environments. These microorganisms thrive in anoxic surficial sediments where sulfate and labile organic substrates coincide. Within inner shelf sediments (<50 m water depth), which typically receive the highest inputs of labile organic matter, area-weighted SRR averages (Fig. 2, A and B) exhibited the highest rates (1.5 nmol cm^{-3} d⁻¹), leading to sub-micromolar sulfate concentrations by 6 mbsf. Furthermore, inner shelf sediments comprise the highest prokaryotic cell abundances (Fig. 2C) and cell specific rates (Fig. 2D) around 0.1 fmol $cell^{-1} d^{-1}$. These data are in strong contrast to deep-water environments, which receive considerably less organic carbon. Peak SRRs in abyss sediments (>3500 m water depth) are a fraction of the shallow-water counterparts, at 0.03 pmol cm⁻³ d⁻¹. Furthermore, cell abundances are in general lower, with the cell-specific rates reaching a maximum of 9×10^{-4} fmol cell⁻¹ d⁻¹.

Simultaneous measurements of SRRs and sulfate reducing microorganism abundances are rare but the existing data are consistent with our model (22-26). The majority of these data exist for relatively shallowwater, high-productivity sites (e.g., Aarhus Bay), with surficial cellspecific SRRs around 0.1 fmol cell⁻¹d⁻¹ and reaching 1.0×10^{-3} fmol $cell^{-1}d^{-1}$ by about 1 mbsf (11). These data are within the range of our areal weighted average for 0.1 mbsf of 0.1 fmol cell⁻¹d⁻¹ (<50 m water depth, Fig. 2). In our modeled shallow-water environments (i.e., inner and outer shelves), high SRRs lead to peak cell-specific rate values near the sediment-water interface. These cell-specific rates taper quickly to zero as sulfate becomes exhausted. Our results show that peak values for the slope also occur near the sediment-water interface (with a slight increase with sediment depth) and gradually decrease as sulfate approaches zero. The abyss, however, does not reach a peak cell-specific rate within the top 80 mbsf, and values remain an order of magnitude lower than the peak values for the other environments. Notably, for the assumed fractions of sulfate reducers within the total microbial community (1 to 30%), the general trends for of cell-specific SRR in different environments persist (Fig. 2D).

Results of the global survey show a distinct trend between environments in the continental margin and the abyss. The abyss (>3500 m water depth) is typically characterized by organic-poor sediments that allow for deep sulfate penetration. This prevalence of sulfate at great sedimentary depths indicates that most cells within the habitable deep sedimentary biosphere (down to 4000 mbsf or the specific basement depth) are found in either oxic or sulfate-reducing settings (Table 2). Nevertheless, within the other environments on the continental margins, sulfate is removed at comparatively shallow sediment depths (<100 m) (Table 2). The sulfate-methane transition zone (SMTZ) is a distinct geochemical horizon that represents an important transition from sulfate reducing (above) to methanogenic sediments (below) (1, 19, 20). Limited data from deep sea cores at these sites suggest that acetate and hydrogen can be abundant and thus serve as substrates for a vast methanogenic subsurface (27–29).

Collectively, these observations indicate that, although the lack of reduced substrate limits sulfate reduction in deep-sea sediments, the continental margins harbor an expansive biosphere below the SMTZ where traditional, energy-rich electron acceptors are exhausted, and thus this fraction of the microbial biosphere is largely fermentative and meth-anogenic (Table 2 and figs. S5 and S6). Roughly estimating the SMTZ at the depth where sulfate depletion reaches 0.1 mM sulfate, habitable sediments located below the SMTZ would comprise a total global subsurface volume of 10^8 km³ (32% of total), hosting approximately 50% of the sub-sea-floor biomass (*12*). However, ~90% of cells in the subsurface at the continental margins (<3500 m water depth) would be situated below the SMTZ (*12*).

References and Notes

- B. B. Jørgensen, S. Kasten, "Sulfur cycling and methane oxidation," in *Marine Geochemistry*, M. Zabel, H. Schulz, Eds. (Springer Press, Berlin, 2006), pp. 271-309.
- D. E. Canfield, Sulfate reduction in deep-sea sediments. Am. J. Sci. 291, 177– 188 (1991). <u>Medline doi:10.2475/ajs.291.2.177</u>
- P. Regnier, P. Friedlingstein, P. Ciais, F. T. Mackenzie, N. Gruber, I. A. Janssens, G. G. Laruelle, R. Lauerwald, S. Luyssaert, A. J. Andersson, S. Arndt, C. Arnosti, A. V. Borges, A. W. Dale, A. Gallego-Sala, Y. Goddéris, N. Goossens, J. Hartmann, C. Heinze, T. Ilyina, F. Joos, D. E. LaRowe, J. Leifeld, F. J. R. Meysman, G. Munhoven, P. A. Raymond, R. Spahni, P. Suntharalingam, M. Thullner, Anthropogenic perturbation of the carbon fluxes from land to ocean. *Nat. Geosci.* 6, 597–607 (2013). doi:10.1038/ngeo1830
- J. P. Dunne, J. L. Sarmiento, A. Gnanadesikan, A synthesis of global particle export from the surface ocean and cycling through the ocean interior and on the seafloor. *Global Biogeochem. Cycles* 21, GB4006 (2007). doi:10.1029/2006GB002907
- S. D'Hondt, S. Rutherford, A. J. Spivack, Metabolic activity of subsurface life in deep-sea sediments. *Science* 295, 2067–2070 (2002). <u>Medline</u> doi:10.1126/science.1064878
- H. E. Hartnett, R. G. Keil, J. I. Hedges, A. H. Devol, Influence of oxygen exposure time on organic carbon preservation in continental margin sediments. *Nature* 391, 572–575 (1998). <u>doi:10.1038/35351</u>
- K. Wallmann, E. Pinero, E. Burwicz, M. Haeckel, C. Hensen, A. Dale, L. Ruepke, The global inventory of methane hydrate in marine sediments: A theoretical approach. *Energies* 5, 2449–2498 (2012). doi:10.3390/en5072449
- J. Kallmeyer, R. Pockalny, R. R. Adhikari, D. C. Smith, S. D'Hondt, Global distribution of microbial abundance and biomass in subseafloor sediment. *Proc. Natl. Acad. Sci. U.S.A.* **109**, 16213–16216 (2012). <u>Medline</u> doi:10.1073/pnas.1203849109
- R. J. Parkes, G. Webster, B. A. Cragg, A. J. Weightman, C. J. Newberry, T. G. Ferdelman, J. Kallmeyer, B. B. Jørgensen, I. W. Aiello, J. C. Fry, Deep sub-

seafloor prokaryotes stimulated at interfaces over geological time. *Nature* **436**, 390–394 (2005). <u>Medline doi:10.1038/nature03796</u>

- B. B. Jørgensen, A. Boetius, Feast and famine—microbial life in the deep-sea bed. *Nat. Rev. Microbiol.* 5, 770–781 (2007). <u>Medline</u> doi:10.1038/nrmicro1745
- T. M. Hoehler, B. B. Jørgensen, Microbial life under extreme energy limitation. *Nat. Rev. Microbiol.* 11, 83–94 (2013). <u>Medline</u> doi:10.1038/nrmicro2939
- 12. See supplementary materials on Science Online.
- G. C. Feldman, C. R. McClain, *Ocean Color Web*, AquaMODros. Inf. Serv., N. Kuring, S. W. Bailey, Eds. (NASA Goddard Space Flight Center, 2013); <u>http://oceancolor.gsfc.nasa.gov/cgi/l3</u>
- S. Levitus, T. P. Boyer, World Ocean Atlas 1994: Volume 4: Temperature. NOAA Atlas NESDros. Inf. Serv. 4. (U.S. Government Printing Office, Washington, DC, (1994).
- D. E. Canfield, E. Kristensen, B. Thamdrup, *Aquatic Geomicrobiology* (Academic Press, Amsterdam, 2005).
- D. Fischer, J. M. Mogollón, M. Strasser, T. Pape, G. Bohrmann, N. Fekete, V. Spiess, S. Kasten, Subduction zone earthquake as potential trigger of submarine hydrocarbon seepage. *Nat. Geosci.* 6, 647–651 (2013). doi:10.1038/ngeo1886
- K. O. Emery, Relict sediments on continental shelves of the world. AAPG Bull. 52, 445–464 (1968).
- J. M. Mogollón, A. W. Dale, J. B. Jensen, M. Schlüter, P. Regnier, A method for the calculation of anaerobic oxidation of methane rates across regional scales: An example from the Belt Seas and The Sound (North Sea–Baltic Sea transition). *Geo-Mar. Lett.* 33, 299–310 (2013). <u>doi:10.1007/s00367-013-0329-z</u>
- J. M. Mogollón, A. W. Dale, H. Fossing, P. Regnier, Timescales for the development of methanogenesis and free gas layers in recently-deposited sediments of Arkona Basin (Baltic Sea). *Biogeosciences* 9, 1915–1933 (2012). doi:10.5194/bg-9-1915-2012
- A. W. Dale, P. Regnier, P. Van Cappellen, H. Fossing, J. B. Jensen, B. B. Jorgensen, Remote quantification of methane fluxes in gassy marine sediments through seismic survey. *Geology* 37, 235–238 (2009). doi:10.1130/G25323A.1
- G. A. Knauer, "Productivity and new production of the oceanic system," in Interactions of C, N, P and S Biogeochemical Cycles and Global Change, R. Wollast, F.T. Mackenzie, L. Chou, Eds. (Springer, Berlin, NATO ASI Series I 4, 211-231, 1993).
- K. Sahm, B. J. MacGregor, B. B. Jørgensen, D. A. Stahl, Sulphate reduction and vertical distribution of sulphate-reducing bacteria quantified by rRNA slot-blot hybridization in a coastal marine sediment. *Environ. Microbiol.* 1, 65–74 (1999). Medline doi:10.1046/j.1462-2920.1999.00007.x
- K. Ravenschlag, K. Sahm, C. Knoblauch, B. B. Jørgensen, R. Amann, Community structure, cellular rRNA content, and activity of sulfate-reducing bacteria in marine arctic sediments. *Appl. Environ. Microbiol.* 66, 3592–3602 (2000). <u>Medline doi:10.1128/AEM.66.8.3592-3602.2000</u>
- 24. J. Leloup, A. Loy, N. J. Knab, C. Borowski, M. Wagner, B. B. Jørgensen, Diversity and abundance of sulfate-reducing microorganisms in the sulfate and methane zones of a marine sediment, Black Sea. *Environ. Microbiol.* 9, 131–142 (2007). <u>Medline doi:10.1111/j.1462-2920.2006.01122.x</u>
- 25. J. Leloup, H. Fossing, K. Kohls, L. Holmkvist, C. Borowski, B. B. Jørgensen, Sulfate-reducing bacteria in marine sediment (Aarhus Bay, Denmark): Abundance and diversity related to geochemical zonation. *Environ. Microbiol.* 11, 1278–1291 (2009). <u>Medline</u> <u>doi:10.1111/j.1462-2920.2008.01855.x</u>
- L. Holmkvist, T. G. Ferdelman, B. B. Jørgensen, A cryptic sulfur cycle driven by iron in the methane zone of marine sediment (Aarhus Bay, Denmark). *Geochim. Cosmochim. Acta* **75**, 3581–3599 (2011). doi:10.1016/j.gca.2011.03.033
- V. B. Heuer, J. W. Pohlman, M. E. Torres, M. Elvert, K.-U. Hinrichs, The stable carbon isotope biogeochemistry of acetate and other dissolved carbon species in deep subseafloor sediments at the northern Cascadia Margin. *Geochim. Cosmochim. Acta* 73, 3323–3336 (2009). doi:10.1016/j.gca.2009.03.001
- S. L. D'Hondt *et al.*, *Proc. ODP*, *Init. Repts.*, 201 (Ocean Drilling Program, College Station, TX, 2003).
- 29. Y.-S. Lin, V. B. Heuer, T. Goldhammer, M. Y. Kellermann, M. Zabel, K.-U.

Hinrichs, Towards constraining H_2 concentration in subseafloor sediment: A proposal for combined analysis by two distinct approaches. *Geochim. Cosmochim. Acta* **77**, 186–201 (2012). doi:10.1016/j.gca.2011.11.008

- 30. R. Mithal, D. G. Becker, "The Janus Database: Providing worldwide access to ODP and IODP data." in *New Techniques in Sediment Core Analysis*, R. G. Rothwell, Ed. (Geological Society of London, London, 2006), vol. 267, pp. 253–259.
- National Geophysical Data Center, National Oceanic and Atmospheric Administration, ETOPO2v2 Global Gridded 2-minute Database, U.S. Dept. of Commerce, <u>www.ngdc.noaa.gov/mgg/global/etopo2.html</u>.
- 32. G. Laske, G. Masters, A global digital map of sediment thickness. *Eos Trans. AGU* **78**, F483 (1997).
- P. Wessel, W. H. F. Smith, a global self-consistent, hierarchical, highresolution shoreline database. J. Geophys. Res. 101, 8741–8743 (1996). doi:10.1029/96JB00104
- 34. R. D. M. Müller, C. Sdrolias, C. Gaina, W. R. Roest, Age, spreading rates, and spreading asymmetry of the world's ocean crust. *Geochem. Geophys. Geosyst.* 9, Q04006 (2008). doi:10.1029/2007GC001743
- B. B. Boudreau, Diagenetic Models and Their Implementations (Springer, 1997).
- 36. T. J. Müller, "Determination of salinity." in *Methods of Seawater Analysis*, K. Grasshoff, K. Kremling, M. Ehrhardt, Eds. (Wiley, Weinheim, Germany, 1999), pp. 41–74.
- N. B. Weston, W. P. Porubsky, V. A. Samarkin, M. Erickson, S. E. Macavoy, S. B. Joye, Porewater stoichiometry of terminal metabolic products, sulfate, and dissolved organic carbon and nitrogen in estuarine intertidal creek-bank sediments. *Biogeochemistry* 77, 375–408 (2006). <u>doi:10.1007/s10533-005-1640-1</u>
- P. J. Müller, E. Suess, Productivity, sedimentation rate, and sedimentary organic matter in the oceans-I. Organic carbon preservation. *Deep-Sea Res.* 26A, 1347–1362 (1979). doi:10.1016/0198-0149(79)90003-7
- J. Milucka, T. G. Ferdelman, L. Polerecky, D. Franzke, G. Wegener, M. Schmid, I. Lieberwirth, M. Wagner, F. Widdel, M. M. Kuypers, Zero-valent sulphur is a key intermediate in marine methane oxidation. *Nature* 491, 541–546 (2012). <u>Medline doi:10.1038/nature11656</u>
- C. Pallud, P. Van Cappellen, Kinetics of microbial sulfate reduction in estuarine sediments. *Geochim. Cosmochim. Acta* 70, 1148–1162 (2006). doi:10.1016/j.gca.2005.11.002
- S. Contreras, P. Meister, B. Liu, X. Prieto-Mollar, K. U. Hinrichs, A. Khalili, T. G. Ferdelman, M. M. Kuypers, B. B. Jørgensen, Cyclic 100-ka (glacialinterglacial) migration of subseafloor redox zonation on the Peruvian shelf. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 18098–18103 (2013). <u>Medline</u> doi:10.1073/pnas.1305981110
- S. Arndt, J.-H. Brumsack, K. W. Wirtz, Cretaceous black shales as active bioreactors: A biogeochemical model for the deep biosphere encountered during ODP Leg 207 (Demerara Rise). *Geochim. Cosmochim. Acta* 77, 186– 201 (2006).
- D. E. Canfield, Sulfate reduction and oxic respiration in marine sediments: Implications for organic carbon preservation in euxinic environments. *Deep-Sea Res.* 36, 121–138 (1989). <u>Medline doi:10.1016/0198-0149(89)90022-8</u>
- L. A. Levin, Oxygen minimum zone benthos: Adaptation and community response to hypoxia. *Oceanogr. Mar. Biol.* 41, 1–45 (2003).
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Supplementary Materials

www.sciencemag.org/cgi/content/full/science.1249213/DC1 Materials and Methods Supplementary Text Figs. S1 to S6 Table S1 to S4 References (30–44)

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Fig. 1. Global sulfate reduction rates in marine sediments. (**A**) Global distribution of depth integrated sulfate reduction rates (mmol $\text{cm}^{-2} \text{ y}^{-1}$) based on predictions of the exponential depth-decay constant, b, from an artificial neural network (ANN) in a 1° x 1° resolution. Black point symbols represent 199 DSDP/ODP/IODP sites with sulfate profiles described by an exponential fit ultimately used to train, validate, and test the ANN. (**B**) The correspondence of predicted b from the ANN and the actual fit values of b for all profiles (R = 0.88).



Fig. 2. Subsurface profiles of area-weighted parameters in various oceanic depth zones. (A) sulfate profiles (mM), (B) sulfate reduction rates (fmol cm⁻³ d⁻¹), (C) cellular abundances (cells cm⁻³) (δ), (D) cell specific rates (fmol cell⁻¹ d⁻¹) with bold lines representing a 10% sulfate reducing microorganism contribution to the total population while the shaded region for each line represents a 1% and 30% contribution of sulfate reducer to the total population.

Table 1. Weighted average depth-integrated sulfate reduction rate for different water depths. The total area covered here is around 349×10^6 km² or about 97% of the total ocean.

Region	Weighted average depth-integrated SRR (x 10 ⁻³ mmol cm ⁻² y ⁻¹)	Total SRR (Tmol y ⁻¹)	% Total SRR (Tmol y ⁻¹)	Area (x 10 ⁶ km ²)
Inner shelf (0-50 m)	39.3	2.9	26	7.6
Outer shelf (50-200 m)	16.2	2.1	19	13
Slope (200-2000 m)	11.2	3.3	29	29
Rise (2000-3500 m)	4.4	2.7	24	63
Abyss (>3500 m)	0.09	0.2	2	237
Global total	3.2	11.3	100	349

Table 2. Global analysis of sulfate reduction rates with respect to published carbon fluxes to the sea floor for various water depth environments. NR = concentration not reached. The global values in bold represent totals, whereas italics represent area weighted averages.

Region	Depth to 0.1 mM sulfate (m)	% Prokary- otes above 0.1 mM sulfate	Carbon flux (<i>3</i>) (Tmol y ⁻¹)	% Carbon re- mineralized by SR	Carbon flux (4) (Tmol y ⁻¹)	% Carbon re- mineralized by SR
Inner shelf (0-50 m)	3.8	0.25	63	16	92	6.3
Outer shelf (50-200 m)	9.9	0.86			43	9.7
Slope (200- 2000 m)	17	5.6			30	22
Rise (2000- 3500 m)	26	31	17	74	26	22
Abyss (>3500 m)	NR	100				
Global total	47	45	80	28	191	12