

# On the importance of ice algae-based energy in a summerly Arctic Ocean

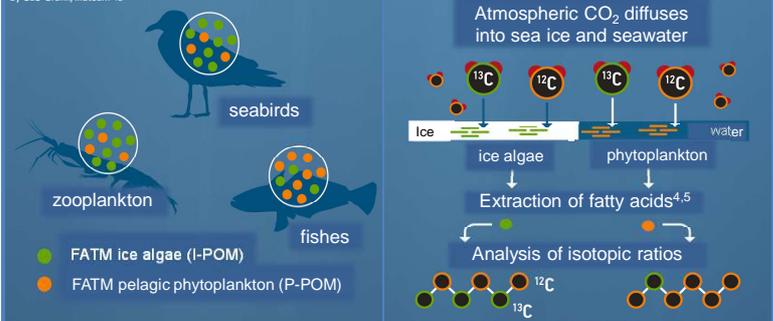
## Food web insights revealed by biomarker approaches

The underside of sea ice in polar regions represents a natural habitat for heterotrophic organisms, e.g. copepods and amphipods. These organisms constitute the under-ice community, which plays a key role in transferring ice algae-produced carbon into pelagic and benthic food webs of polar ecosystems. Animals at higher trophic levels show an indirect dependency on microalgae-produced biomass. In order to improve our understanding of the potential ecological consequences of a changing sea ice environment, we aim to quantify the extent to which ice algae-produced carbon is channelled into the under-ice community, and from there to pelagic food webs.

### Methods.

Trophic interactions of abundant under-ice zooplankton were studied using **bulk stable isotope analysis (BSIA)** of natural abundance carbon and nitrogen<sup>1</sup>, **fatty acid trophic marker (FATM) fingerprinting**, and **compound-specific SIA (CSIA)** of FATMs<sup>2</sup>. Sample collection was carried out during ARK XXVII-3 expedition of RV Polarstern (August-September 2012) within the Eastern Central Arctic Ocean north of 80°N. The under-ice habitat was sampled by the SUIT, the Surface and Under-Ice Trawl<sup>3</sup>.

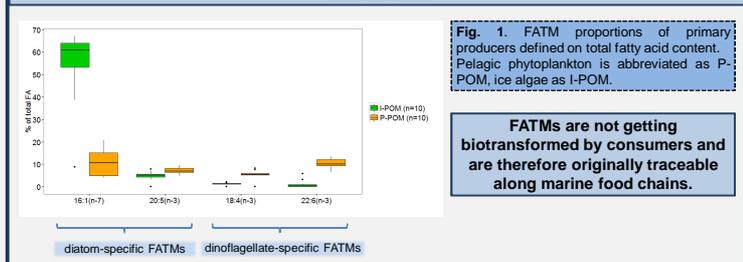
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### Results.

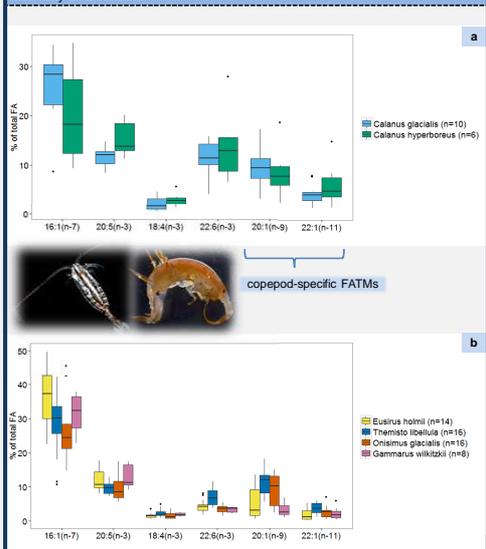
Based on fatty acid patterns, copepods *C. glacialis* and *C. hyperboreus* were feeding on both, ice algae and pelagic phytoplankton. Several amphipod species demonstrated high amounts of diatom-related fatty acids. Besides, *T. libellula* and *O. glacialis* indicated *Calanus*-integrated diets. Based on diatom-specific FA 20:5(n-3), FA material derived from ice algae accounted for averaged 56 % in zooplanktonic consumers (Table 1).

**Dinoflagellates are the main taxonomic group of P-POM, diatoms of I-POM. Taxonomic composition of I-POM and P-POM communities can be similar, e.g. diatoms can occur in both communities.**



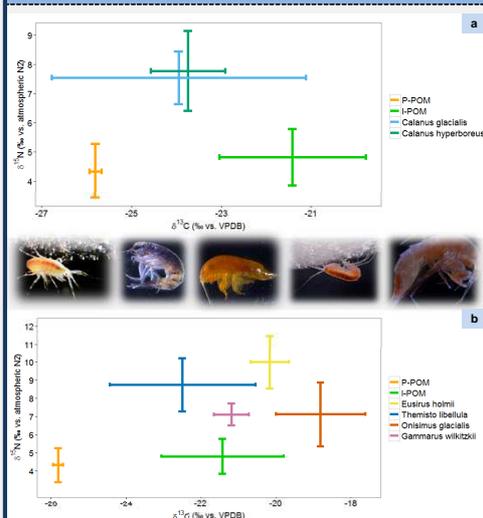
### FATM FINGERPRINTING

**Fig. 2.** FATM proportions of Arctic copepods (a) and amphipods (b) defined on total fatty acid content.



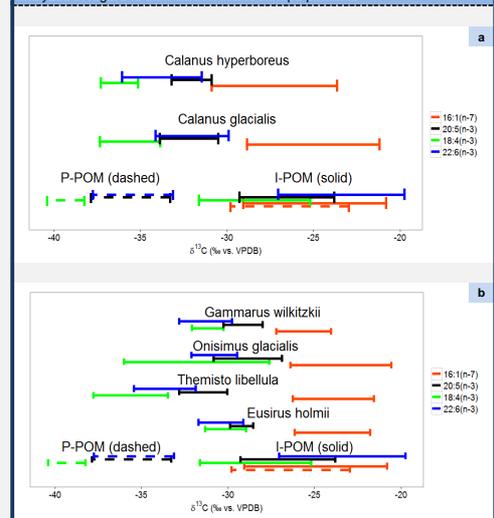
### BSIA

**Fig. 3.** Stable isotope compositions of bulk carbon and nitrogen compounds for Arctic copepods (a) and amphipods (b). Isotopic ratios are expressed as:  $\delta X = [(R_{Sample}/R_{Standard}) - 1] \times 1000$ , where X is  $\delta^{13}C$  or  $\delta^{15}N$  and  $R_{Sample}$  represents  $^{13}C/^{12}C$  or  $^{15}N/^{14}N$  relative to international standards.



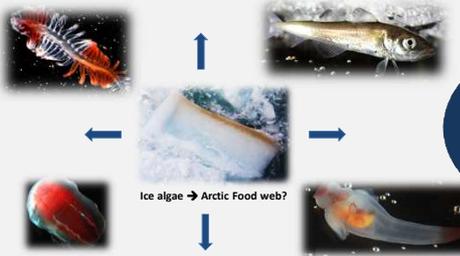
### CSIA

**Fig. 4.** Compound-specific stable isotopic compositions of FATMs for Arctic copepods (a) and amphipods (b). Extracted FATMs were separated by gas chromatography and individually analyzed in regard to their carbon stable isotope patterns.



**Differences in  $\delta^{13}C$  between primary producers allow quantification of proportional distribution of ice algae-derived carbon x in consumers based on certain FATMs (Two-end-member mixing model, [Table 1])<sup>2</sup>.**

Species	X <sub>20:5(n=3)</sub>	X <sub>22:6(n=3)</sub>
<i>Calanus glacialis</i>	37.3±18.6	28.4±17.6
<i>Calanus hyperboreus</i>	39.1±16.7	16.7±15.0
<i>Eusirus holmi</i>	70.5±7.5	41.8±10.8
<i>Themisto libellula</i>	45.8±15.5	14.7±14.9
<i>Onisimus glacialis</i>	74.3±21.9	38.6±11.0
<i>Gammarus wilkitzkii</i>	71.3±12.6	34.3±12.7



**Conclusions.**  
Fatty acid signatures reflect the potential carbon sources, which are supported by stable isotope values. Even in summer, ice algae-produced carbon plays an important role for the diet of Arctic ecological key species.

References  
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<sup>2</sup>Budge, S. M., Wooller, M. J., Springer, A. M., Iverson, S. J., McRoy, C. P., Drevick, G. J. Tracing carbon flow in an arctic marine food web using fatty acid-stable isotope analysis. *Oecol.*, 157, 117-129 (2008)  
<sup>3</sup>van Franeker, J. A., Flores, H., van Dorssen, M. The Surface and Under Ice Trawl (SUIT). Frozen Desert Alive: The role of sea ice for pelagic macrofauna and its predators. PhD thesis, University of Groningen, 181-188 (2009)  
<sup>4</sup>Edlich, J., Lees, M., Stanley, G.H.S. A simple method for the isolation and purification of total lipids from animal tissues. *J. Biol. Chem.*, 226, 497-509 (1957)  
<sup>5</sup>Bligh, E. G., Dyer, W. J. A rapid method of total lipid extraction and purification. *Can. J. Biochem. Physiol.*, 37, 911-917 (1959)