

Background

In times of a rising plastic production the occurrence of microplastics (MPs, <5 mm in size) in the marine environment has been identified as an emerging topic of global concern. MPs are omnipresent in our environment, hardly degradable and are easily ingested by a wide range of organisms throughout all trophic levels. However, the extent of this MP pollution as well as the resulting impacts on the marine environment remains largely unknown. Therefore, standardized and reliable methods to securely detect MPs are urgently needed. The conclusive identification requires a successful extraction from different, complex environmental matrices.

Methods

Collection of surface water samples with 100 µm net (RV Heincke, 2014) mpli 24 stations, approx. 34000 L per station \rightarrow 1 L sample



Splitting into two size fractions with 500 µm stainless steel mesh:

>500 µm

Visual sorting with stereomicroscope and a Bogorov chamber

<500 µm

- Maceration of organic sample matrix in **MP-reactors** ^[1] using an **enzymatic**oxidative treatment ^[2]
- ZnCl₂ separation Density with $(\rho = 1.7 \text{ g cm}^{-3})$ to remove inorganic matrix

volume to cover measuring filter by using

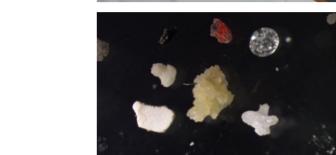
Of

appropriate



State-of-the-art methods were used and optimized to assess occurrence of MPs in the size range of $11 - 5000 \,\mu\text{m}$ in North Sea surface waters. Therefore plankton samples from 23 stations were analyzed to gain information on MP quantities as well as polymer composition and size distribution.

Results for MP particles >500 µm and <500 µm



Analyzed with ATR (Attenuated total reflectance) FTIR (Fourier transform infrared) spectroscopy



Analyzed with µFTIR spectroscopy and subsequent automated analysis ^[3] that compares spectra to a profound database

Determination

the FlowCam



sample

MPs in North Sea surface waters

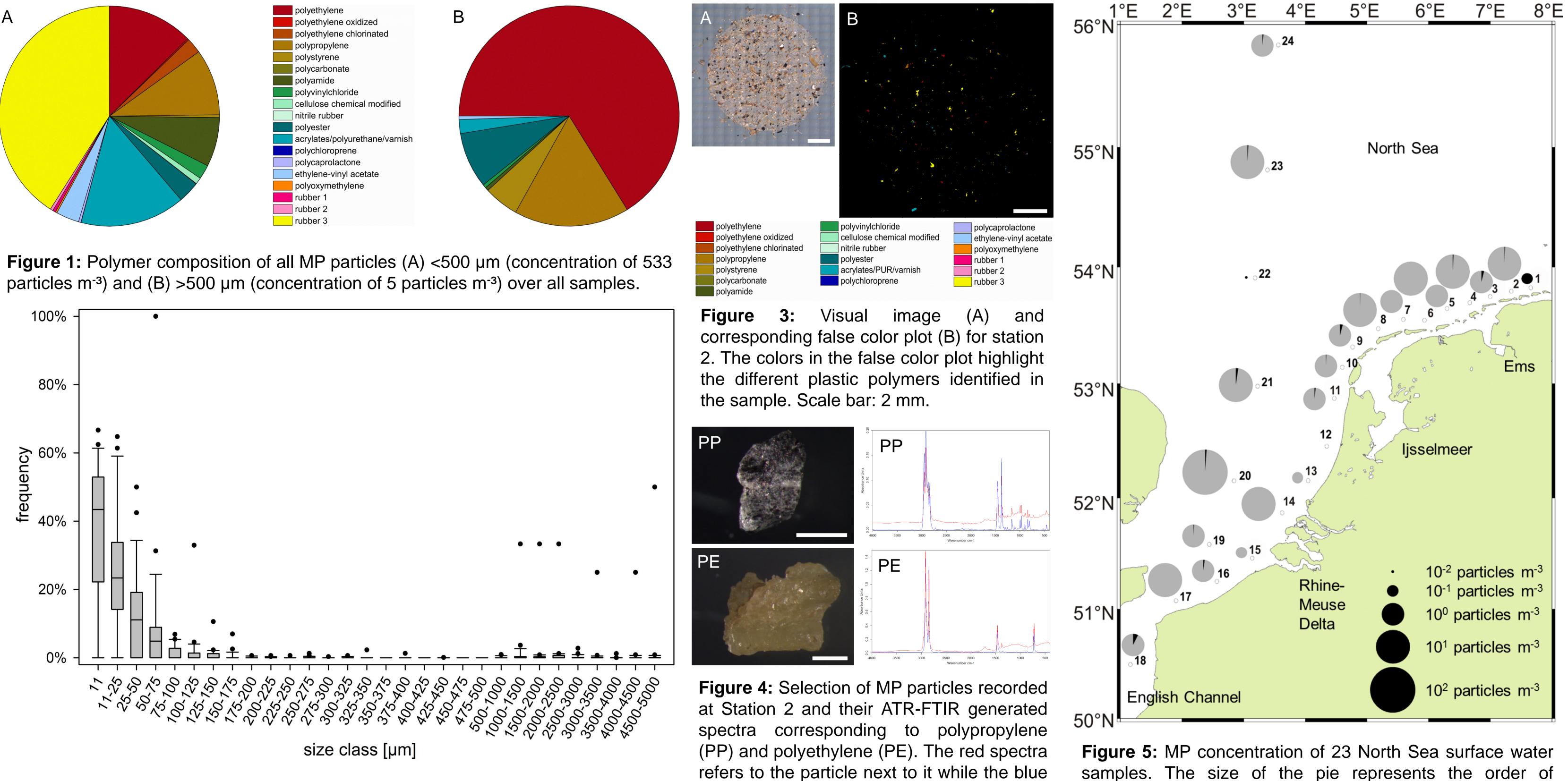


Figure 2: Proportional size distribution of all identified MPs over all stations ranging from 11 to 5000 µm.

samples. The size of the pie represents the order of magnitude. The black piece represents the proportion of particles $>500 \mu m$ and the gray piece of particles $<500 \mu m$.

Conclusions

The method proved successful, revealing MP occurrence in North Sea surface waters at all 23 analyzed stations.

 \rightarrow MP concentrations: 0 – 2.5 particles m⁻³ for MPs >500 µm vs. 0 – 211.4 particles m⁻³ for MP <500 µm

 \succ MP polymer composition: 8 different polymers for MP >500 μ m vs. 19 different polymers for MPs <500 μ m

 \succ Size distribution: 6 % of MPs >500 µm vs. 94 % of MPs <500 µm and almost 90 % of MPs being < 75 µm in length

The size distribution, differences in polymer composition as well as MP concentrations show that it is not sufficient to just analyze particles >500 µm because the results are not representative for the smaller MPs.

500 µm.



References:

[1] Gerdts G (2017) Reaktor zur enzymatischen Mazeration biogener Bestandteile einer Partikelprobe und Verwendung des Reaktors, DE102016123324 (B3) [2] Löder MGJ, Imhof HK, Ladehoff M, Löschel LA, Lorenz C, Mintenig S et al. (2017) Enzymatic Purification of Microplastics in Environmental Samples. Environ Sci Technol 51:14283-14292

[3] Primpke S, Lorenz C, Rascher-Friesenhausen R, Gerdts G (2017) An automated approach for microplastics analysis using focal plane array (FPA)

FTIR microscopy and image analysis. Anal Methods 9:1499-1511

spectrum serves as reference. Scale bar:

