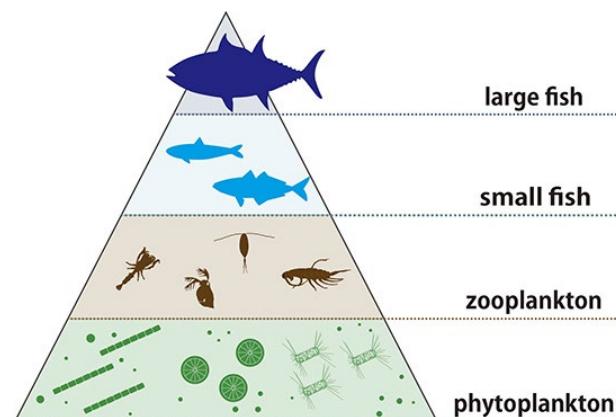


# 20 Years of HAB Research with the Help of Mass Spectrometry

# Introduction

- Marine toxins = Phycotoxins are toxic chemicals produced by photosynthetic plankton-species
- Dinoflagellates are the principle producers of phycotoxins
  - Also toxigenic diatoms or cyanobacteria amongst others
- Accumulate in a variety of filter feeding bivalves or shellfish and can reach high concentrations during algal blooms
  - “Harmful algal blooms” = HABs



# Introduction

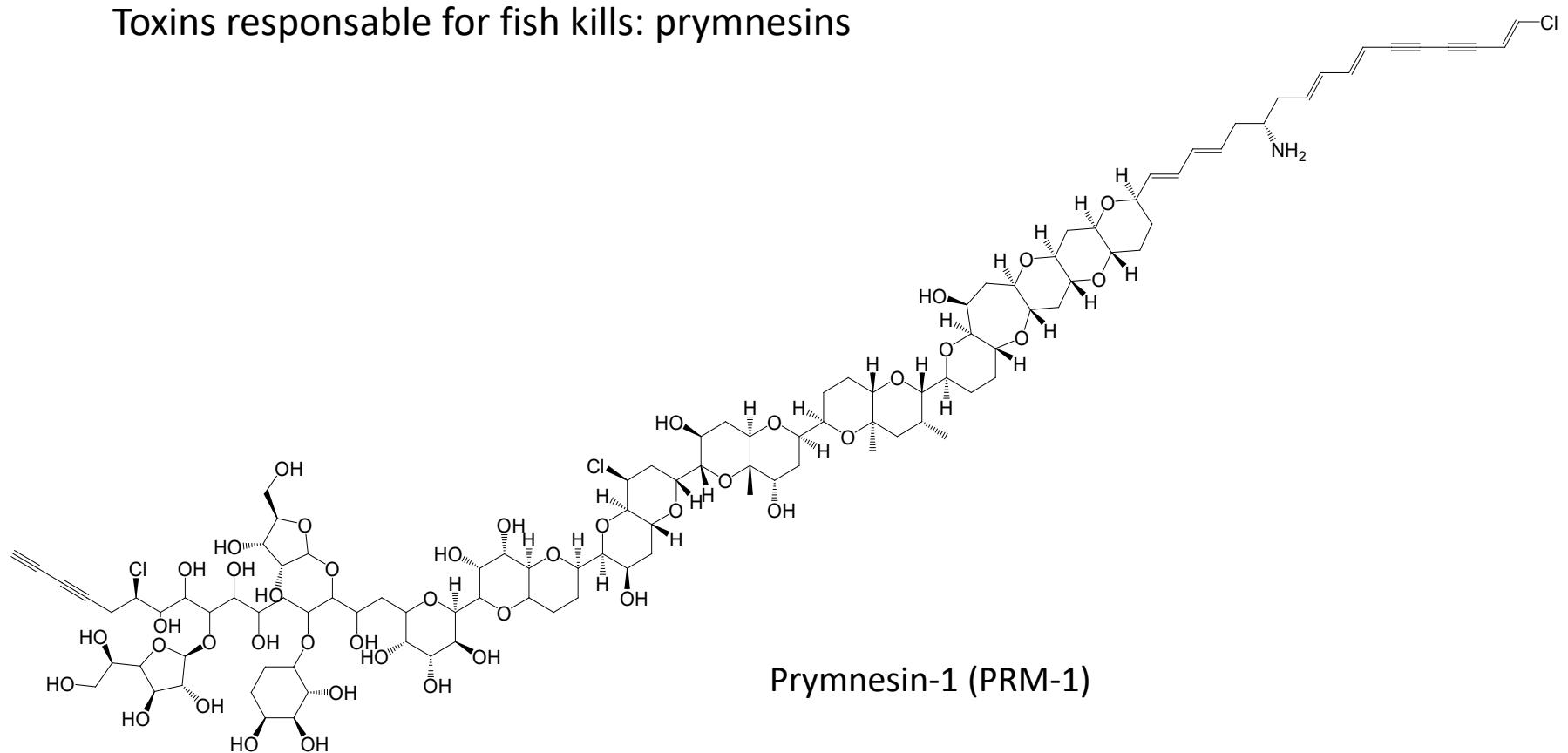
## Massive Fish Kill in the Oder River in August 2022



caused by the brackish haptophyte *Prymnesium parvum*

# Introduction

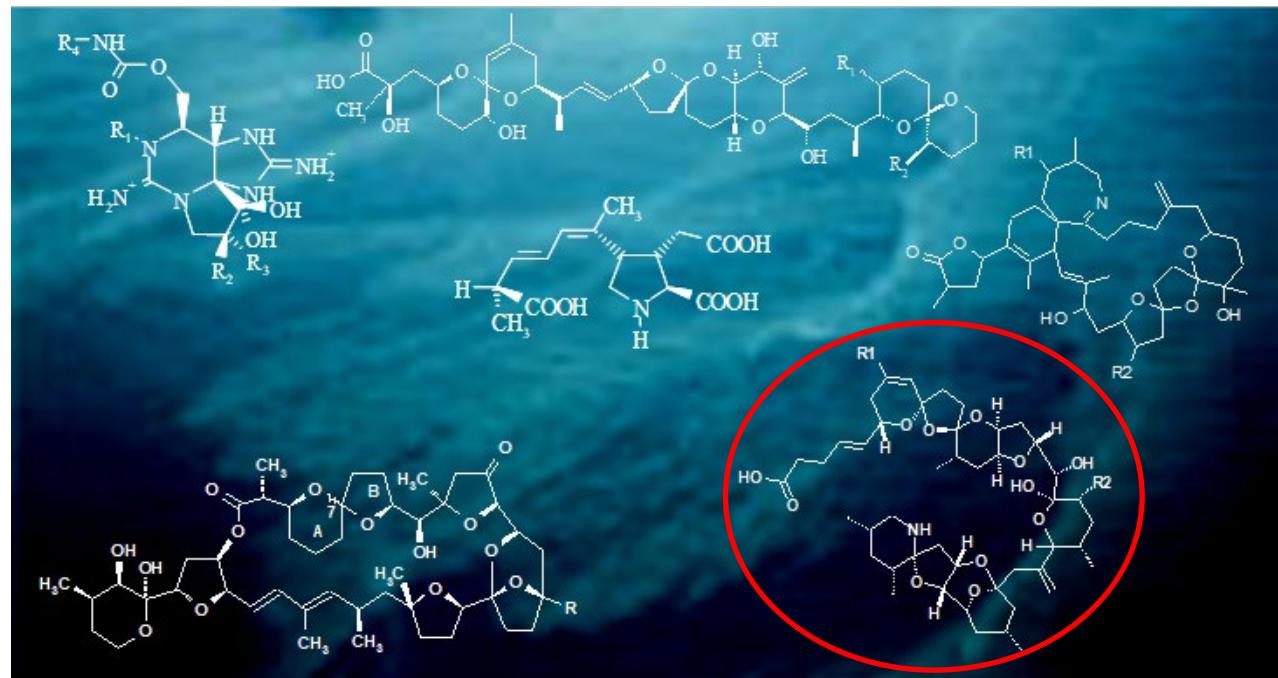
Toxins responsible for fish kills: prymnesins



# Introduction

Amphidinol  
Anatoxin  
**Azaspiracid**  
Brevetoxin  
Ciguatoxin  
Cylindrospermopsin  
Domoic Acid  
Goniodomin  
Gymnocin  
Gymnodimine  
Karenia brevisulcata toxin  
Karlotoxin  
Lyngbyatoxin  
Microcystin  
Nodularin  
Okadaic acid  
Palytoxin  
Pectenotoxin  
Pinnatoxin  
Prymnesin  
Saxitoxin  
Spirolide  
Yessotoxin

## High variability of phycotoxin classes



Need of monitoring for seafood safety

# Strategies for the Discovery of Source Organisms of Phycotoxins

## 1) Biological approach

- Plankton sampling
- Species isolation
- Establishment of a monoclonal culture
- Toxin testing

## 2) Chemical approach

- Plankton sampling
- Toxin testing
- Species isolation
- Establishment of a monoclonal culture

# Finding phycotoxin producers

- 1995: 8 people in the Netherlands became ill after consumption of Irish mussels (*Mytilus edulis*) harvested at Killary Harbour (Ireland). Symptoms were like DSP intoxication, but DSP toxins were hardly present in the mussels (MacMahon & Silke, 1996: Harmful Algae News, 14, 2)
- 1998: Satake et al. identified azapiracid-1 (AZA-1) as the causative compound in shellfish (J. Am. Chem. Soc., 120, 9967-9968)
- 1999: Ofuji et al. elucidated the structures of AZA-2 and AZA-3 from shellfish (Nat. Toxins, 7, 99-102)
- 2001: Ofuji et al. elucidated the structures of AZA-4 and AZA-5 from shellfish (Biosci. Biotechnol. Biochem., 65, 470-472)
- 2003: James et al. elucidated the structures of AZA-6 to AZA-11 from shellfish (Toxicon, 41, 277-283)

# Finding phycotoxin producers



ALFRED-WEGENER-INSTITUT  
HELMHOLTZ-ZENTRUM FÜR POLAR-  
UND MEERESFORSCHUNG



Toxicon 41 (2003) 145–151

## Toxicon

[www.elsevier.com/locate/toxicon](http://www.elsevier.com/locate/toxicon)

### Ubiquitous ‘benign’ alga emerges as the cause of shellfish contamination responsible for the human toxic syndrome, azaspiracid poisoning

Kevin J. James<sup>a,\*</sup>, Cian Moroney<sup>a</sup>, Cilian Roden<sup>a</sup>, Masayuki Satake<sup>b</sup>,  
Takeshi Yasumoto<sup>c</sup>, Mary Lehane<sup>a</sup>, Ambrose Furey<sup>a</sup>

<sup>a</sup>Proteobio, Department of Chemistry, Mass Spectrometry Centre for Proteomics and Biotoxin Research,  
Cork Institute of Technology, Bishopstown, Cork, Ireland

<sup>b</sup>Faculty of Agriculture, Tohoku University, Tsutsumidori-Amamiya, Aoba-ku, Sendai, Japan

<sup>c</sup>Japan Food Laboratories, Tama Laboratory, Nagayama, Tama-shi, Tokyo, Japan

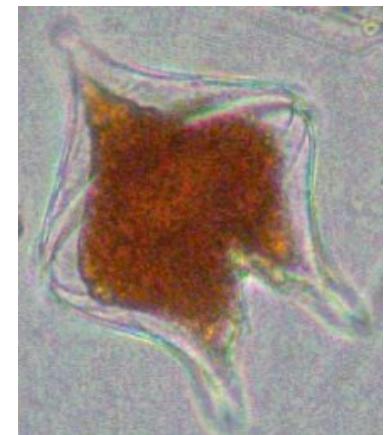
Received 5 June 2002; accepted 5 August 2002

#### Abstract

A new human toxic syndrome, azaspiracid poisoning (AZP), was identified following illness from the consumption of contaminated mussels (*Mytilus edulis*). To discover the aetiology of AZP, sensitive analytical protocols involving liquid chromatography–mass spectrometry (LC–MS) were used to screen marine phytoplankton for azaspiracids. Collections of single species were prepared by manually separating phytoplankton for LC–MS analysis. A dinoflagellate species of the genus, *Protoperidinium*, has been identified as the progenitor of azaspiracids. Azaspiracid-1, and its analogues, AZA2 and AZA3, were identified in extracts of 200 cells using electrospray multiple tandem MS. This discovery has significant implications for both human health and the aquaculture industry since this phytoplankton genus was previously considered to be toxicologically benign. The average toxin content was 1.8 fmol of total AZA toxins per cell with AZA1 as the predominant toxin, accounting for 82% of the total.

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**Keywords:** Marine toxins; LC–MS; AZP; *Protoperidinium*; Shellfish poisoning



*Protoperidinium crassipes*



# Finding phycotoxin producers



Correlations between the presence of known toxic phytoplankton species and toxin levels in shellfish in Irish waters 2002 – 2006

Siobhan Moran\*, J Silke, C Cusack, P Hess  
Marine Institute, Galway, Ireland  
[siobhan.moran@marine.ie](mailto:siobhan.moran@marine.ie)

The Irish National Monitoring Programme for phytoplankton is part of the Irish Shellfish Biotoxin Monitoring Programme, which fulfills Regulation (EC) 853/2004. The four main toxic syndromes found in Irish waters are Diarrhetic, Paralytic, Amnesic, and Azaspiracid Shellfish Poisoning.

Over a four year period (2002 – 2006) there was no correlation between the occurrence of *Protoperidinium* spp. in plankton and azaspiracids in shellfish in Irish waters.

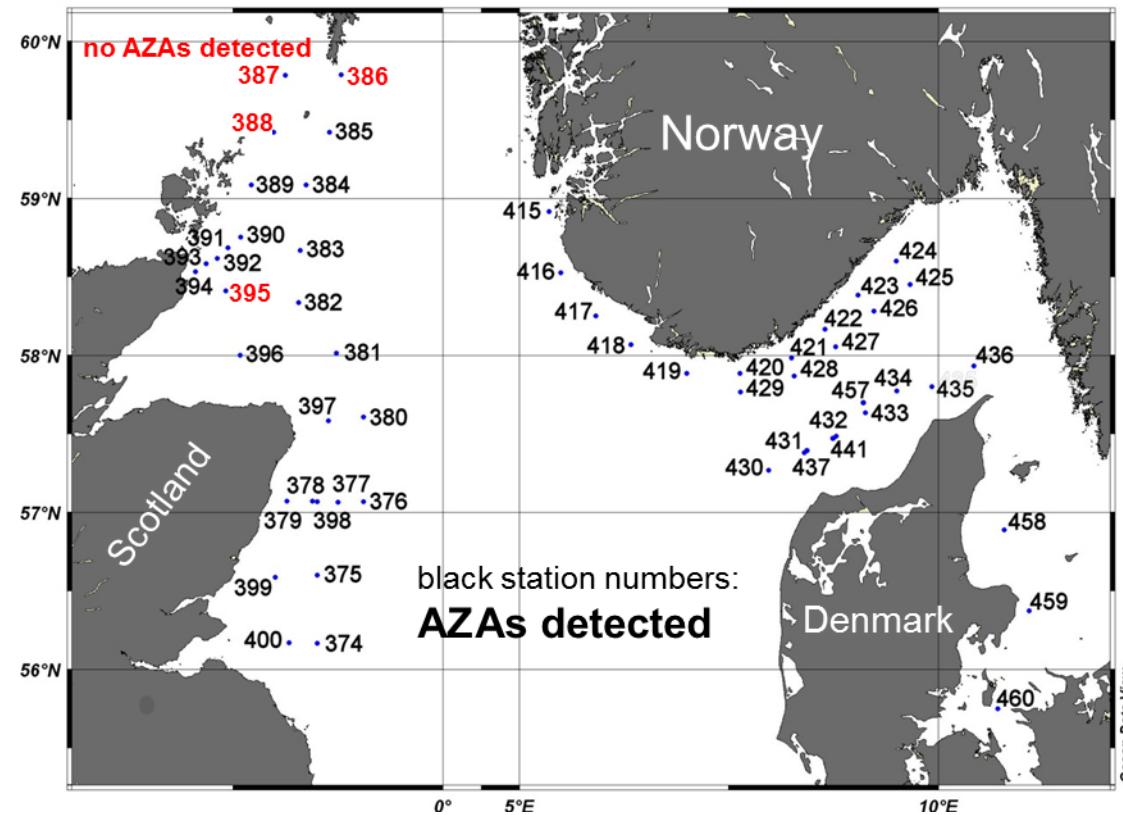
The authors exclude *Protoperidinium* as the source of azaspiracids

Possible reason for the misidentification of *Protoperidinium crassipes*:

*P. crassipes* as a heterotrophic dinoflagellate might have fed on the azaspiracid producing organism during a toxic event

# Finding phycotoxin producers

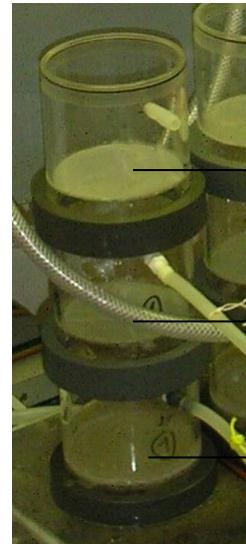
Survey on the North Sea in July 2007



# Finding phycotoxin producers



Plankton net  
pore size 20 µm



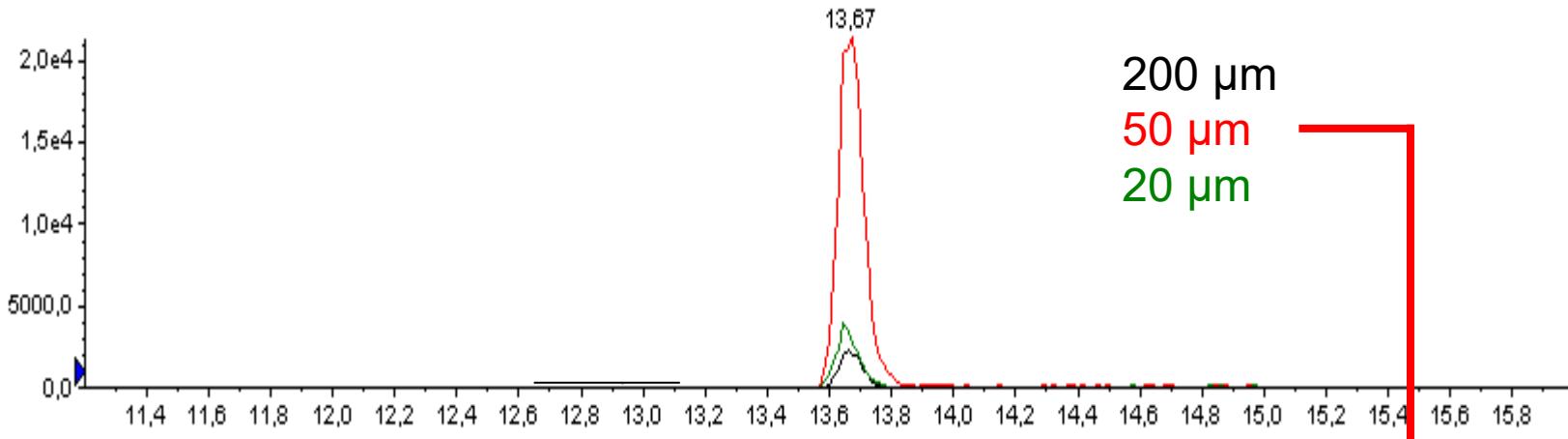
Filter array

LC-MS/MS



# Finding phycotoxin producers

## Size fraction test



Look into 50  $\mu\text{m}$  plankton fraction



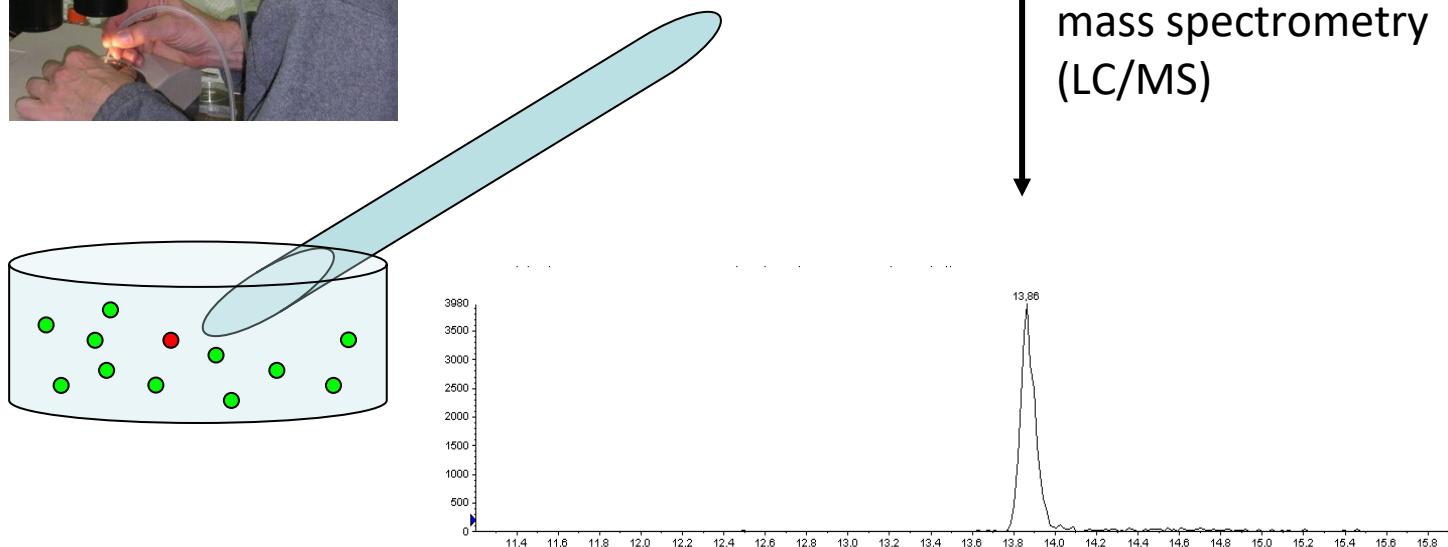
*Favella ehrenbergii*

# Finding phycotoxin producers



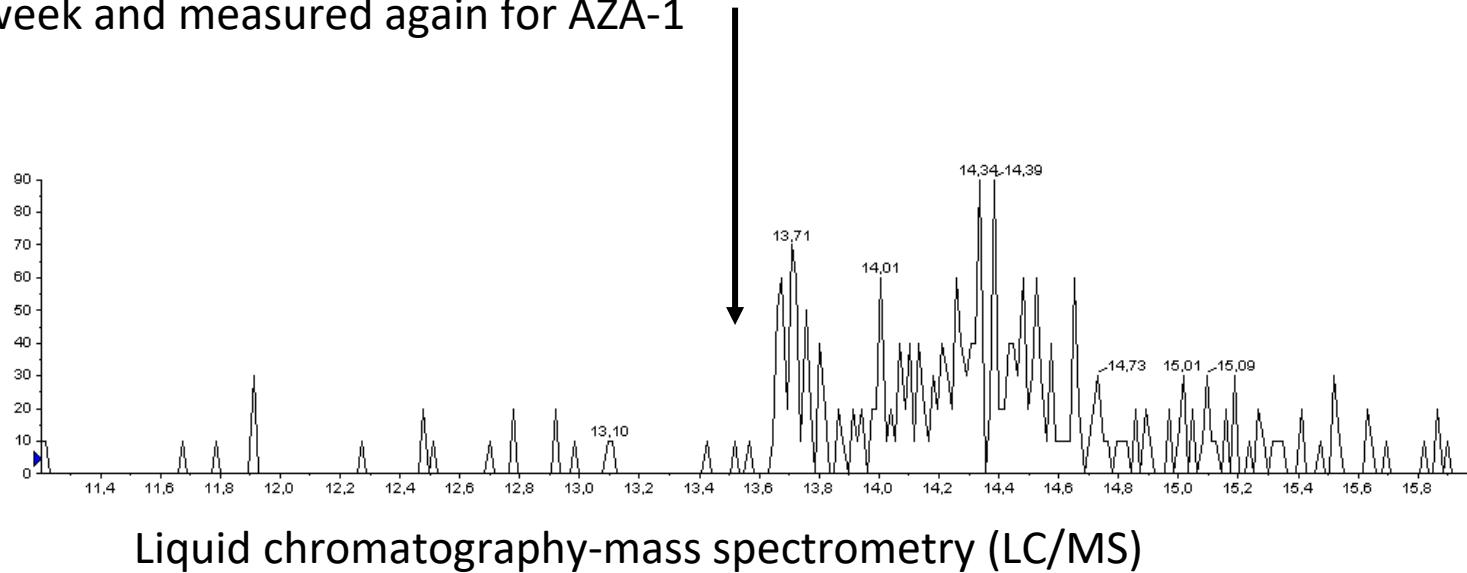
Single cell isolation of 160 *F. ehrenbergii* individuals with a microcapillary

Liquid chromatography-mass spectrometry (LC/MS)



# Finding phycotoxin producers

Azaspiracid containing *F. ehrenbergii* were fed with non-toxic *Scrippsiella* for one week and measured again for AZA-1



*Favella ehrenbergii*

=> *F. ehrenbergii* is not an azaspiracid producer!

# Finding phycotoxin producers



If *Favella* feeds on  
AZA producers,  
they cannot be very  
big!

=> screening of size  
fractions < 20 µm for  
AZA

# Finding phycotoxin producers

Screening of plankton < 20 µm for AZA



Rosette sampler  
(unfiltered water samples)

Filtration over 20 µm mesh

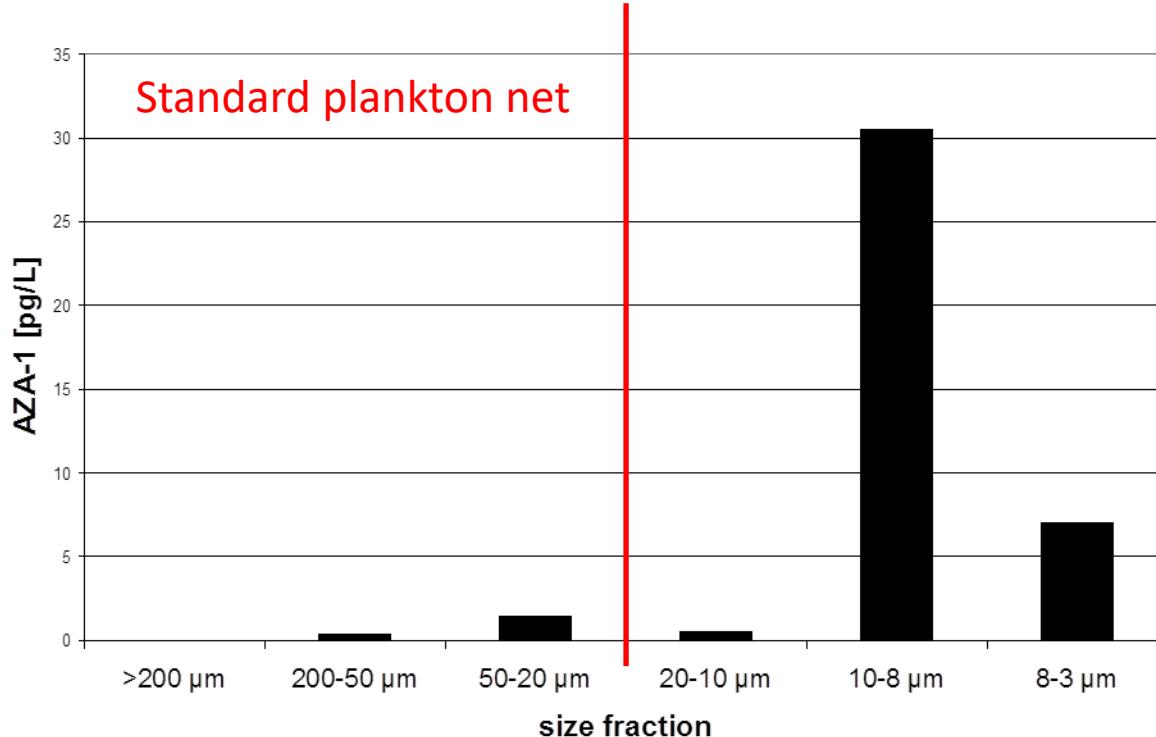


Removal of plankton > 20 µm



Filtration over 10 µm (mesh), 8 µm, 3 µm and 0.2 µm (polycarbonate filters)

# Finding phycotoxin producers

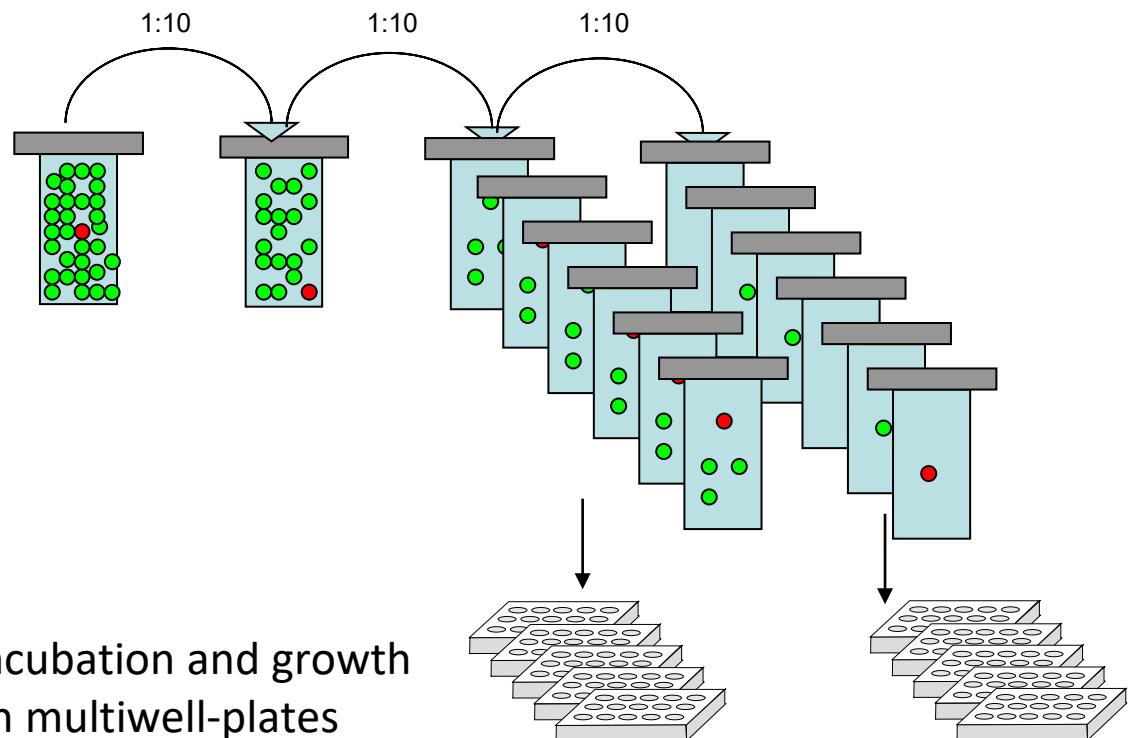


- => the AZA producer is smaller than 10 µm
- => the AZA producer can only be sampled by direct water collection, but not by phytoplankton net tows

# Finding phycotoxin producers

AZA containing plankton sample from the field

Serial dilution method



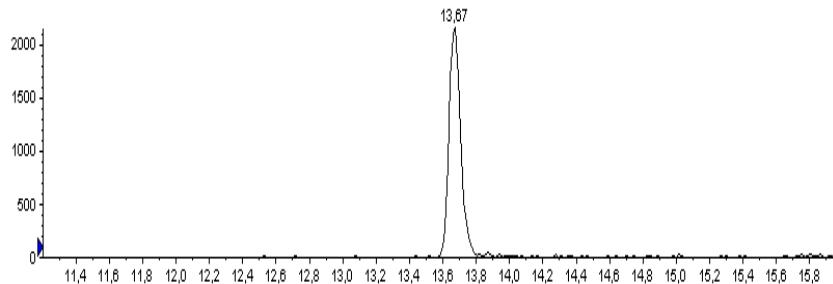
Incubation and growth  
in multiwell-plates

# Finding phycotoxin producers

Out of 240 isolates tested

only one culture “3 D9”  
contained AZA-1

AZA-1

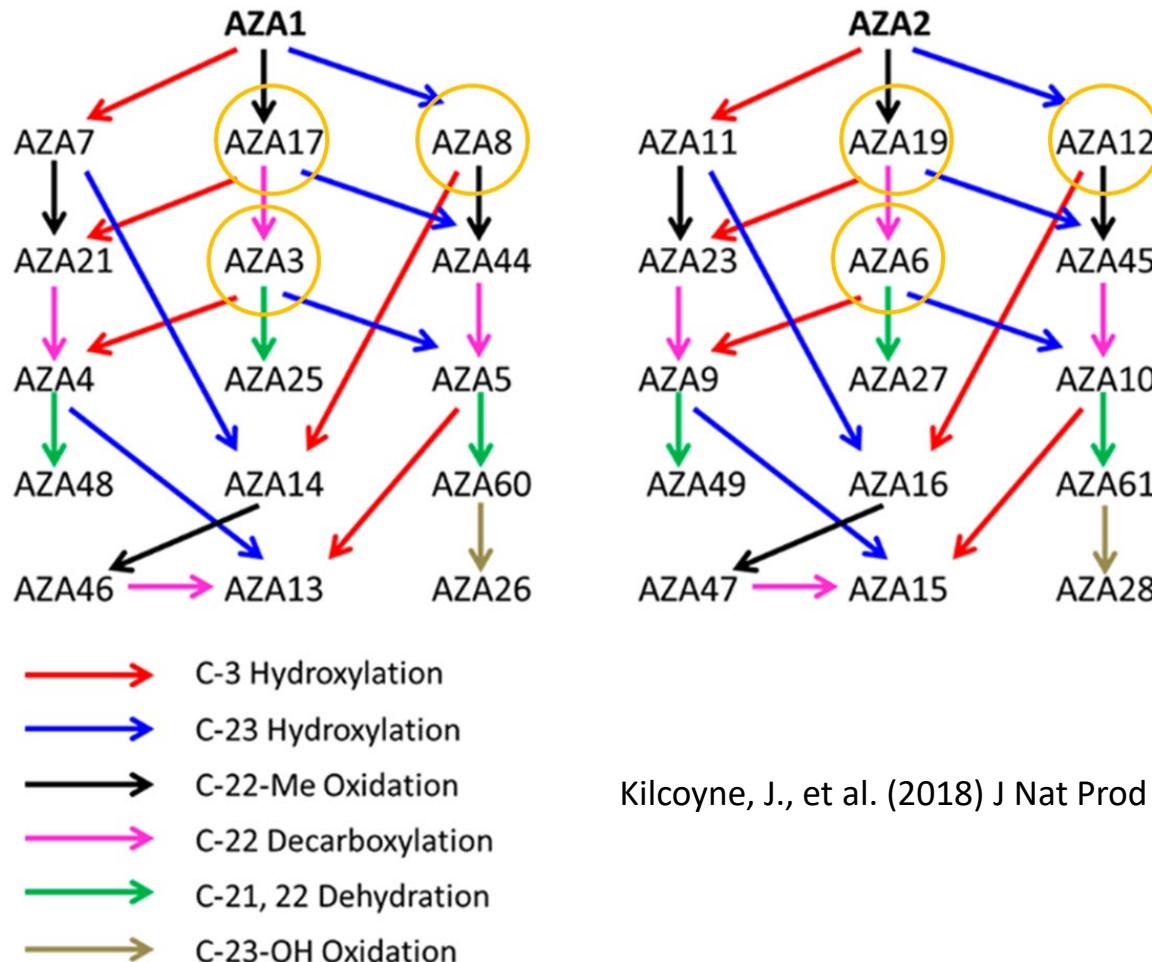


Krock, B., et al., 2009, Harmful Algae 8(2): 254-263.

# Elucidation of the High Structural Variability of Phycotoxins in Plankton and Higher Trophic Levels of the Food Chain

# Structural variability of phycotoxins

## Azaspiracids – Metabolism in Bivalves



Kilcoyne, J., et al. (2018) *J Nat Prod* 81(4), 885-893.

# Structural variability of phycotoxins

AZA-1	AZA-41
AZA-2	AZA-42
epi-AZA-7	AZA-43
AZA-11	AZA-50
AZA-33	AZA-51
AZA-34	AZA-52
AZA-35	AZA-53
AZA-36	AZA-54
AZA-37	AZA-55
AZA-38	AZA-56
AZA-39	AZA-57
AZA-40	AZA-58
AZA-59	AZA-62

AZA-3	AZA-14	AZA-25	AZA-47
AZA-4	AZA-15	AZA-26	AZA-48
AZA-5 <sup>+</sup>	AZA-16	AZA-27	AZA-49
AZA-6	AZA-17	AZA-28	AZA-60
AZA-7	AZA-18	AZA-29	AZA-61
AZA-8	AZA-19	AZA-30	
AZA-9	AZA-20	AZA-31	
AZA-10	AZA-21	AZA-32	
AZA-11*	AZA-22	AZA-44	
AZA-12	AZA-23	AZA-45	
AZA-13	AZA-24	AZA-46	

\* Tillmann, U., et al. (2017) J Plankt Res 39(2), 350-367.

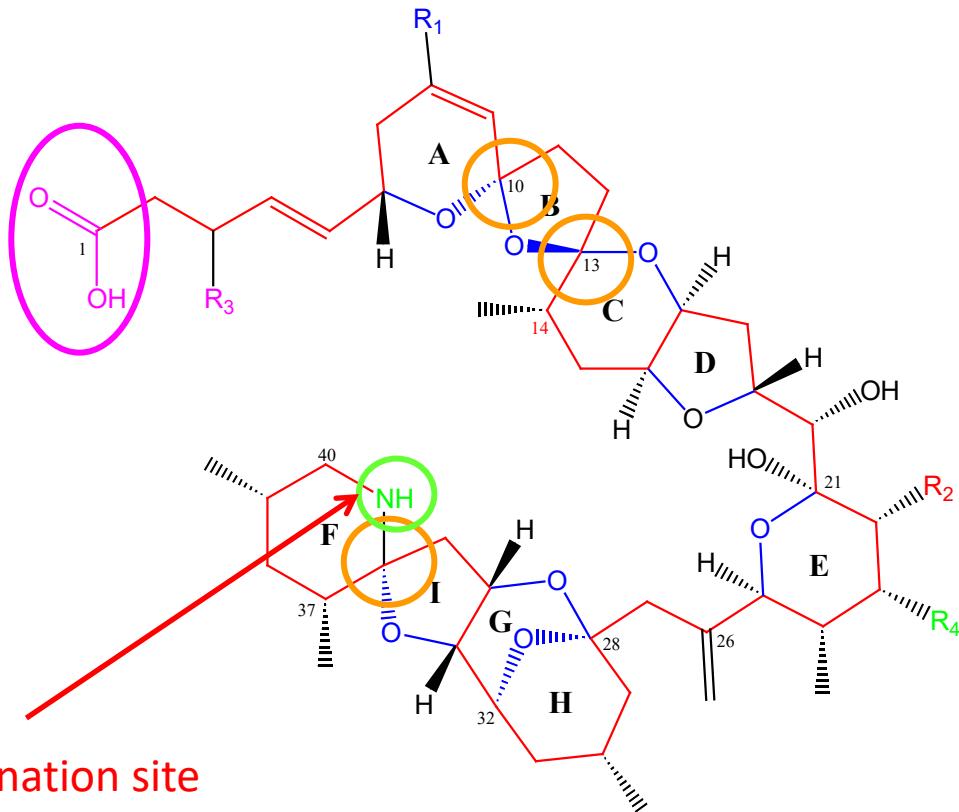
+ Montoya, N., pers. commun.

Krock, B., et al. (2019) Harmful Algae 82, 1-8.

<sup>1</sup> Additional planktonic AZA reported by Ozawa, M., et al. (2021) Toxicon 199: 145-155.

Two AZAs of phytoplankton origin result in 38 shellfish metabolites!

# Structural variability of phycotoxins



azaspiracid

Polyketide:  
linear carbon skeleton  
with cyclic ether  
bridges

amino function  
chemical nomenclature:  
aza = secondary amine

spiro function

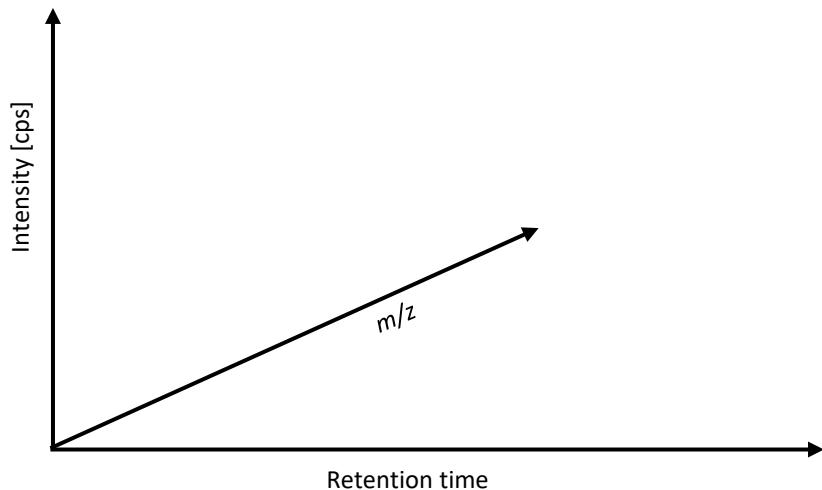
acid

# Structural variability of phycotoxins

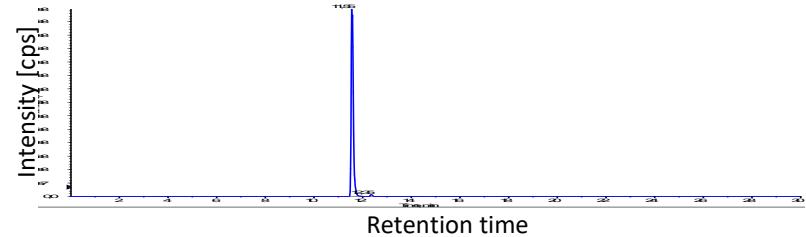
## LC-MS/MS data set

LC-MS data are a three dimensional data set:

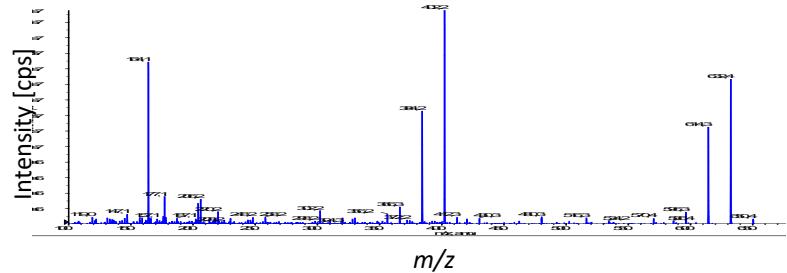
1. Retention time (chromatography)
2. Mass axis ( $m/z$ ) (mass spectrometry)
3. Ion intensity



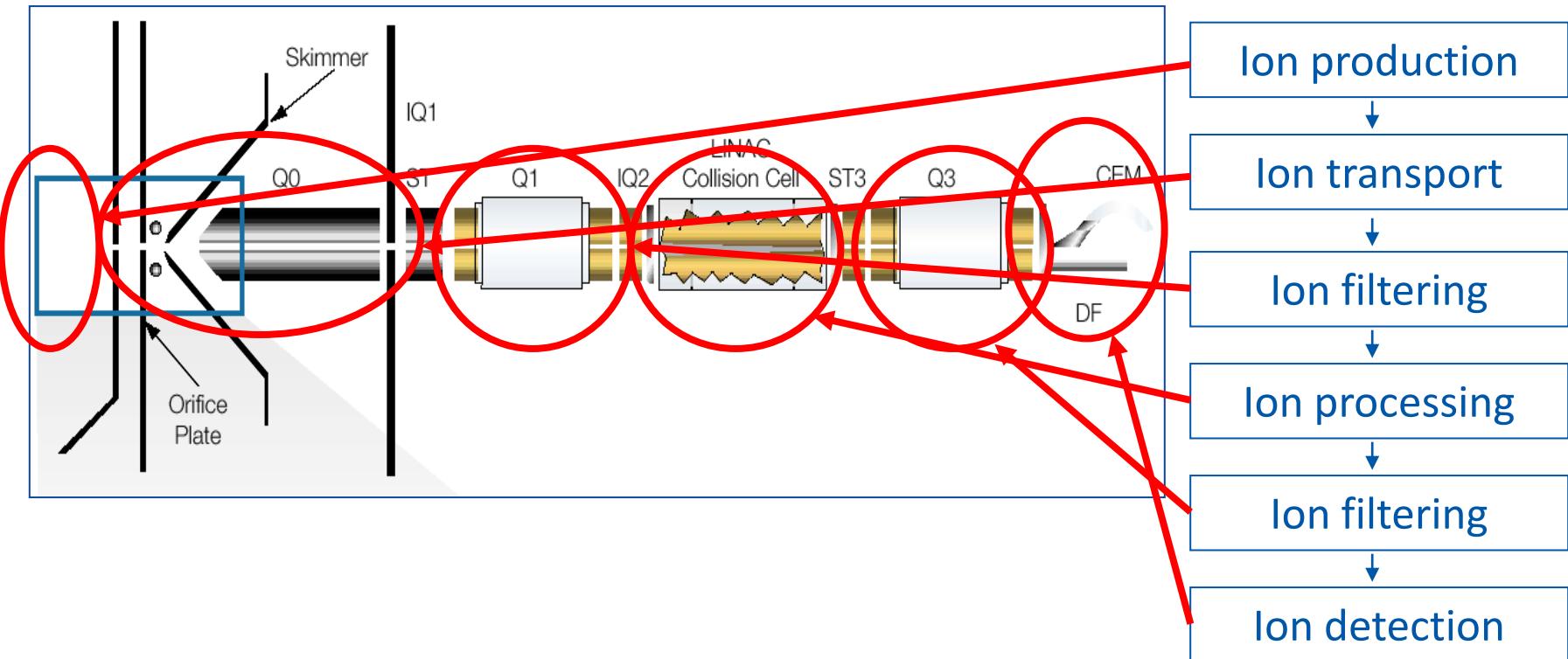
LC-MS chromatogram



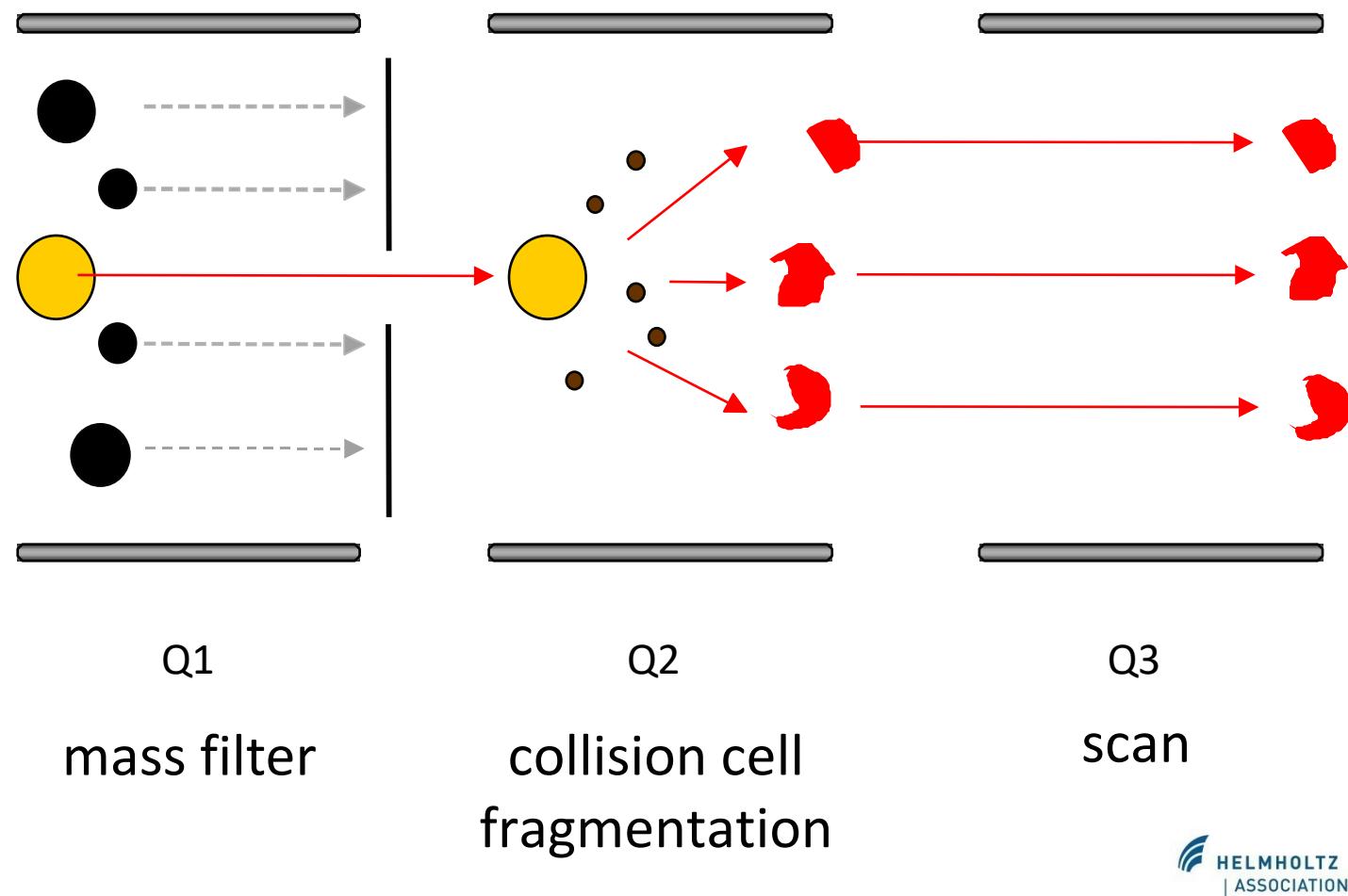
LC-MS spectrum



## Scheme of a Triple Quadrupole Instrument



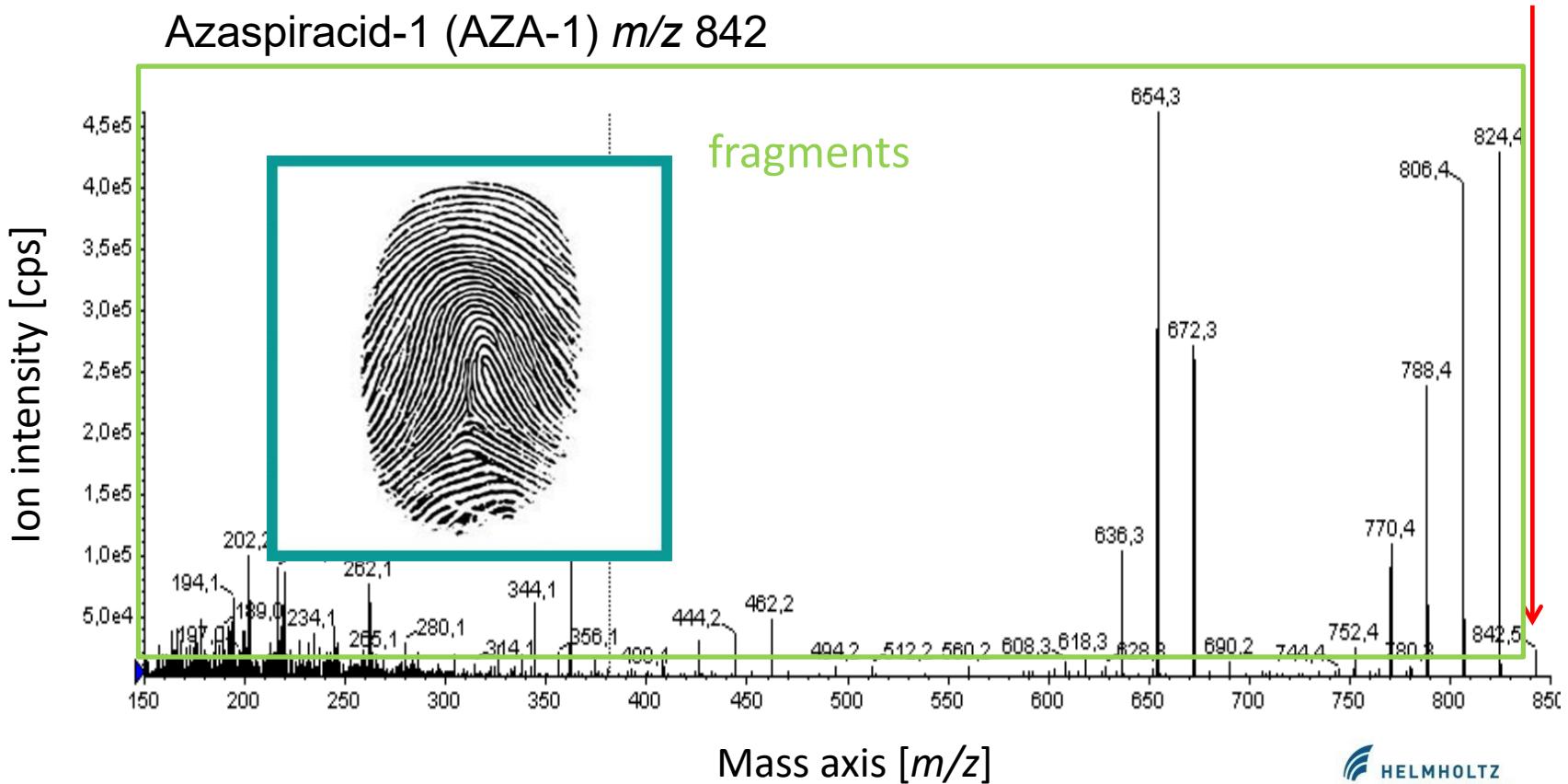
## Product (Daughter) Ion Scan



# Structural variability of phycotoxins

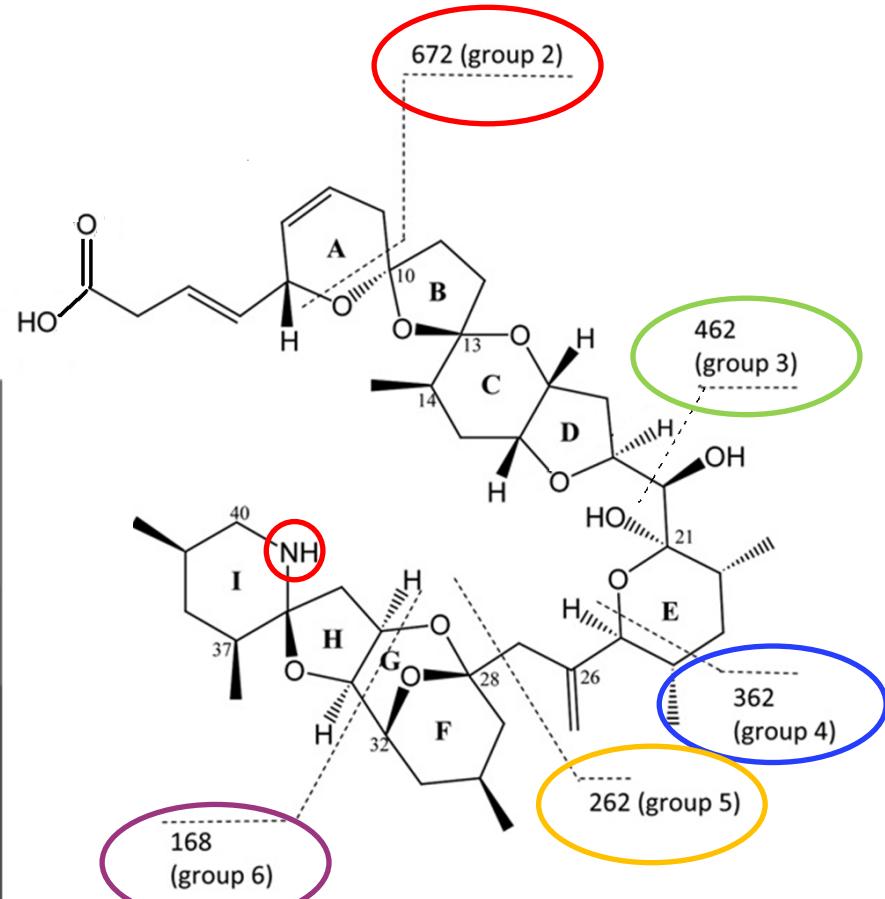
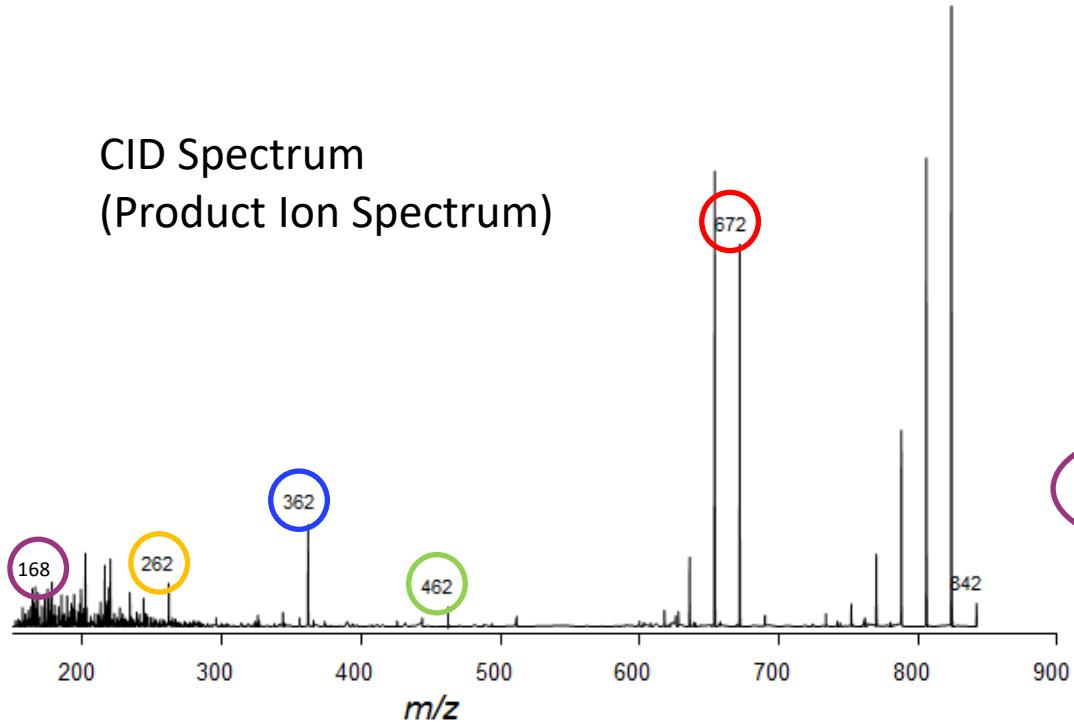
- Collision induced dissociation (CID) spectrum
- Product ion spectrum
- Mass spectrum

(pseudo)  
molecular  
ion



# Structural variability of phycotoxins

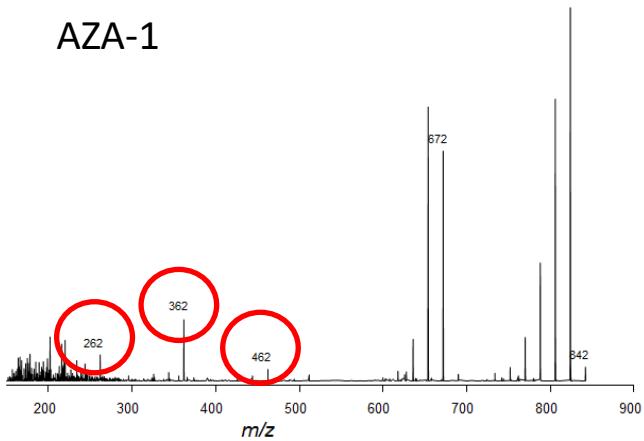
Azaspiracid-1 (AZA-1)  $m/z$  842



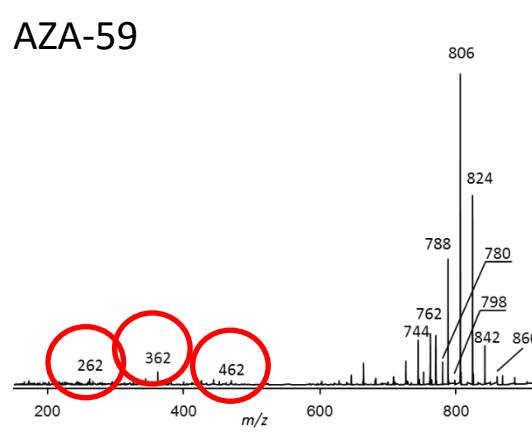
# Structural variability of phycotoxins

## Product Ion (CID) Spectra of Azaspiracids

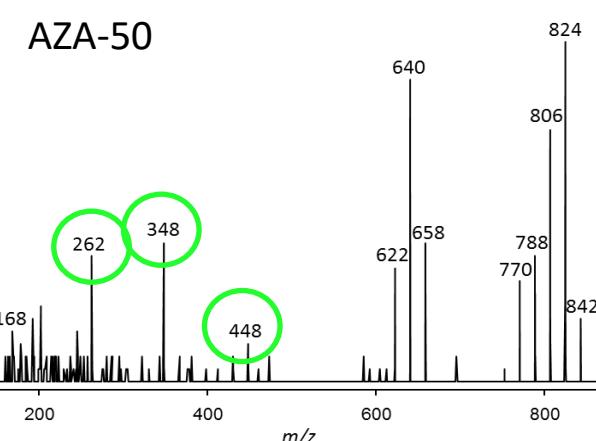
AZA-1



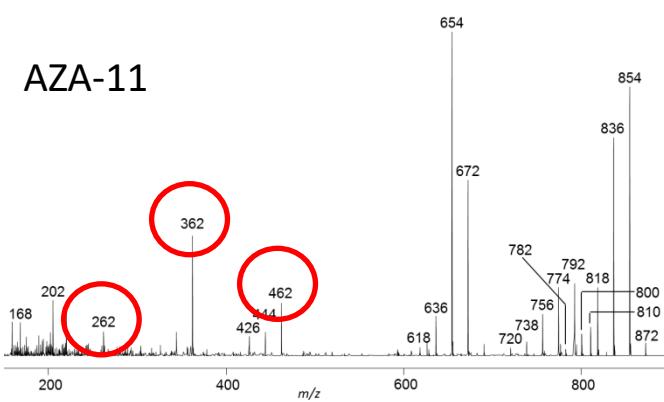
AZA-59



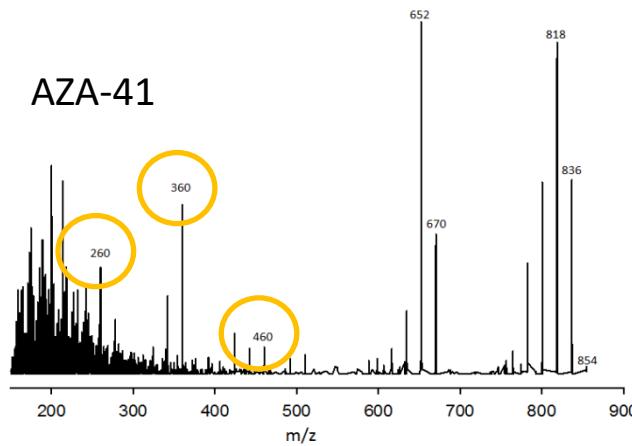
AZA-50



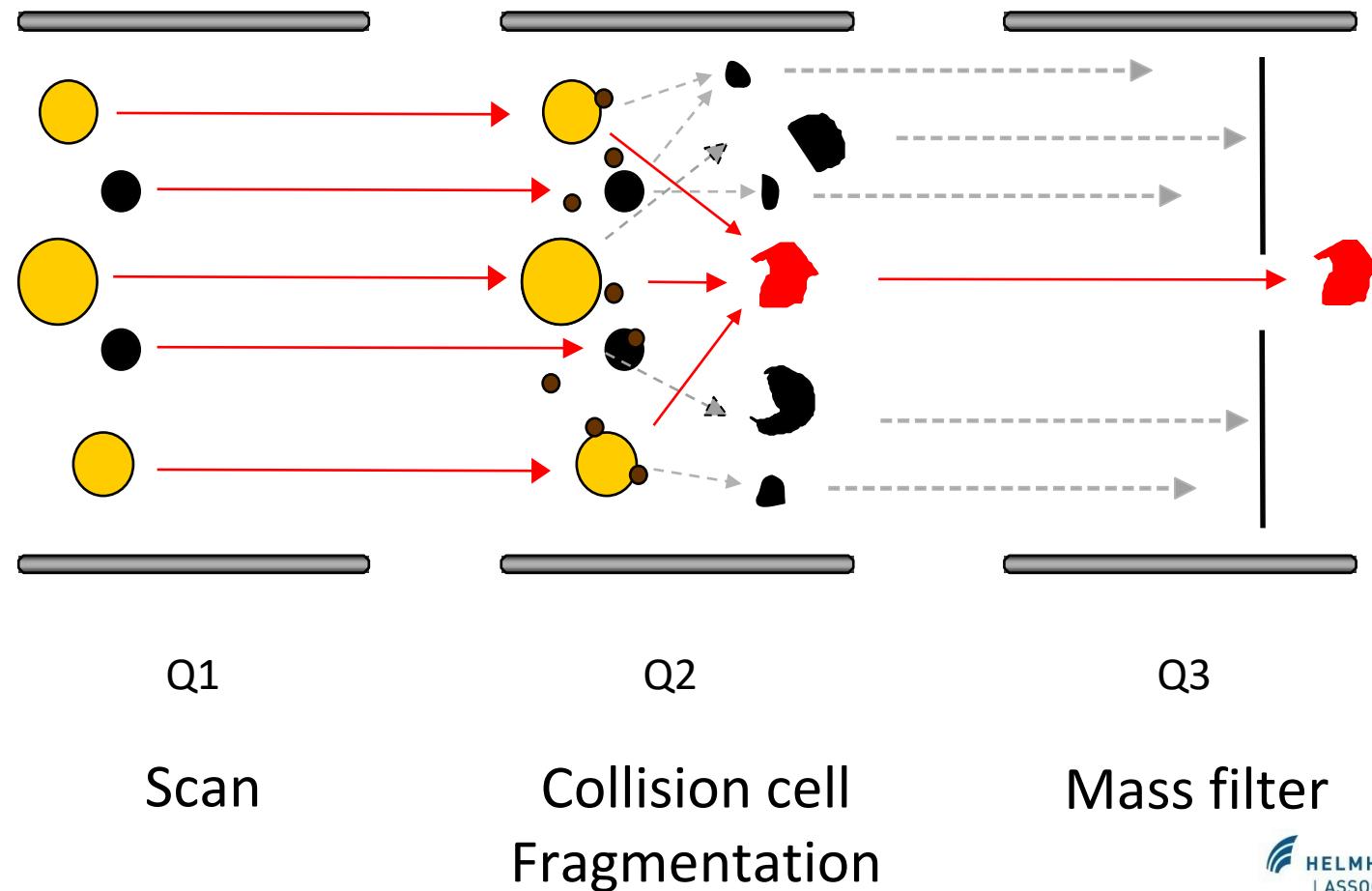
AZA-11



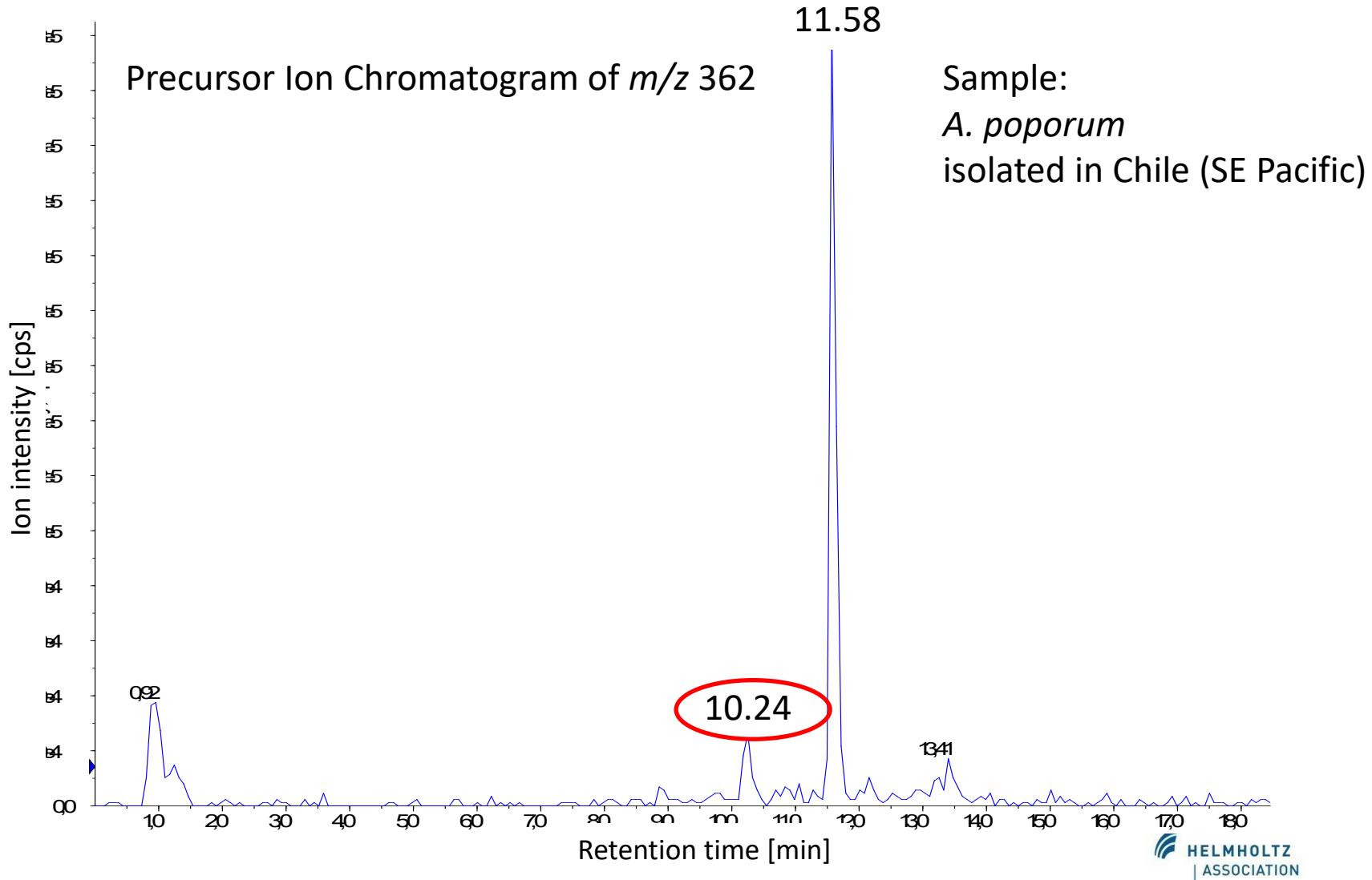
AZA-41



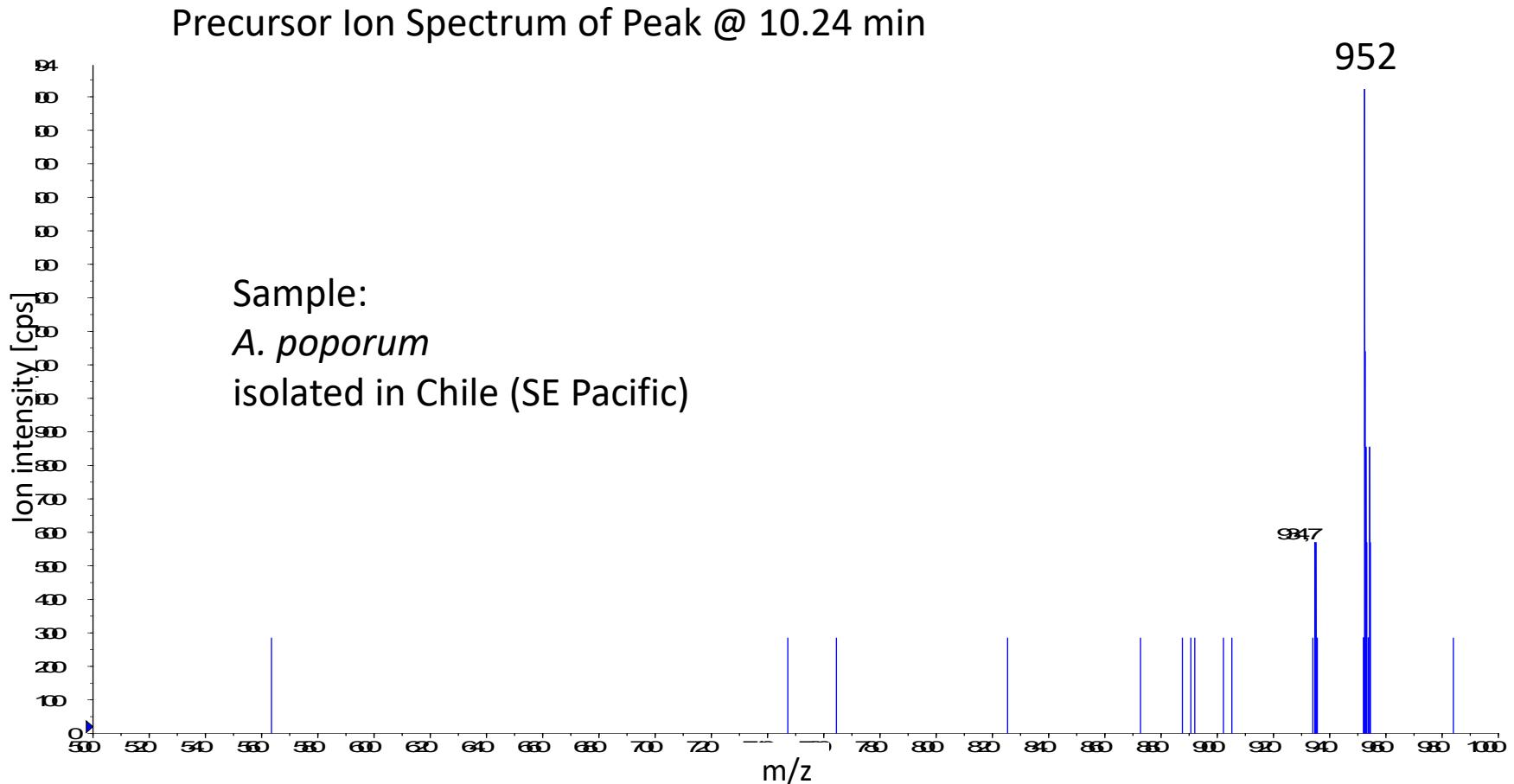
## Search for Unknowns – Precursor Scan



# Structural variability of phycotoxins



# Structural variability of phycotoxins



# Structural variability of phycotoxins

CID Spectrum of  $m/z$  952

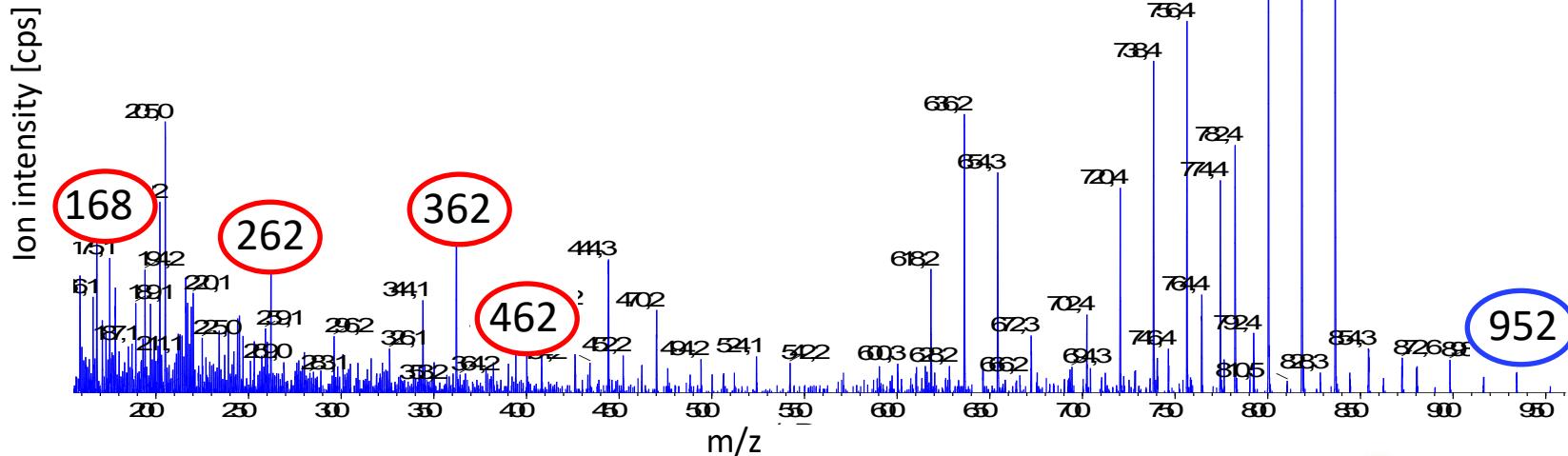
AZA-11 phosphate

Sample:

*A. poporum*

isolated in Chile (SE  
Pacific)

Tillmann, U. et al. (2017) J. Plankt. Res. 39(2): 350-367.



# Occurrence and Spatial Distribution of Phycotoxins and Toxigenic Microalgal Species

# Occurrence of Toxins and their Producers

#	AZA	m/z [M+H] <sup>+</sup>	m/z group 4 fragment	m/z group 5 fragment	Producer	Reference
1	AZA-1	842	362	262	<i>A. spinosum</i>	Krock et al. 2009
2	AZA-2	856	362	262	<i>A. spinosum</i>	Krock et al. 2009
					<i>A. poporum</i>	Krock et al. 2014
					<i>Am. languida</i>	Tillmann et al. 2017
3	epi-AZ				<i>A. dexteroporum</i>	Rossi et al. 2017
4	AZA-1				<i>A. poporum</i>	Krock et al. 2014
5	AZA-3				<i>A. spinosum</i>	Kilcoyne et al. 2014
6	AZA-34	818	362	262	<i>A. spinosum</i>	{Kilcoyne et al. 2014}
7	AZA-35	830	362	262	<i>A. spinosum</i>	Kilcoyne et al. 2014
					<i>A. dexteroporum</i>	Rossi et al. 2017
8	AZA-36	858	340	248	<i>A. poporum</i>	Krock et al. 2015
9	AZA-3				<i>A. poporum</i>	Krock et al. 2015
10	AZA-3				<i>Am. languida</i>	Krock et al. 2012
11	AZA-3				<i>Am. languida</i>	Krock et al. 2012
12	AZA-4				<i>A. poporum</i>	Krock et al. 2014
13	AZA-41	854	360	260	<i>A. poporum</i>	Krock et al. 2014
14	AZA-42	870	360	260	<i>A. poporum</i>	Krock et al. under review
15	AZA-43	828	360	260	<i>Am. languida</i>	Tillmann et al. 2017
16	AZA-5				<i>A. spinosum</i>	Tillmann et al. 2018
17	AZA-5				<i>A. spinosum</i>	Tillmann et al. 2018
18	AZA-5				<i>Am. languida</i>	Tillmann et al. 2018
19	AZA-5				<i>Am. languida</i>	Tillmann et al. 2018
20	AZA-5				<i>A. dexteroporum</i>	Rossi et al. 2017
21	AZA-5				<i>A. dexteroporum</i>	Rossi et al. 2017
22	AZA-5				<i>A. dexteroporum</i>	Rossi et al. 2017
23	AZA-57	844	362	262	<i>A. dexteroporum</i>	Rossi et al. 2017
24	AZA-58	828	362	262	<i>A. dexteroporum</i>	Rossi et al. 2017
25	AZA-59	860	362	262	<i>A. poporum</i>	Kim et al. 2017
26	AZA-62	870	362	262	<i>A. poporum</i>	Krock et al. 2019

Currently Known AZA  
from Dinoflagellate  
(Algal) Origin

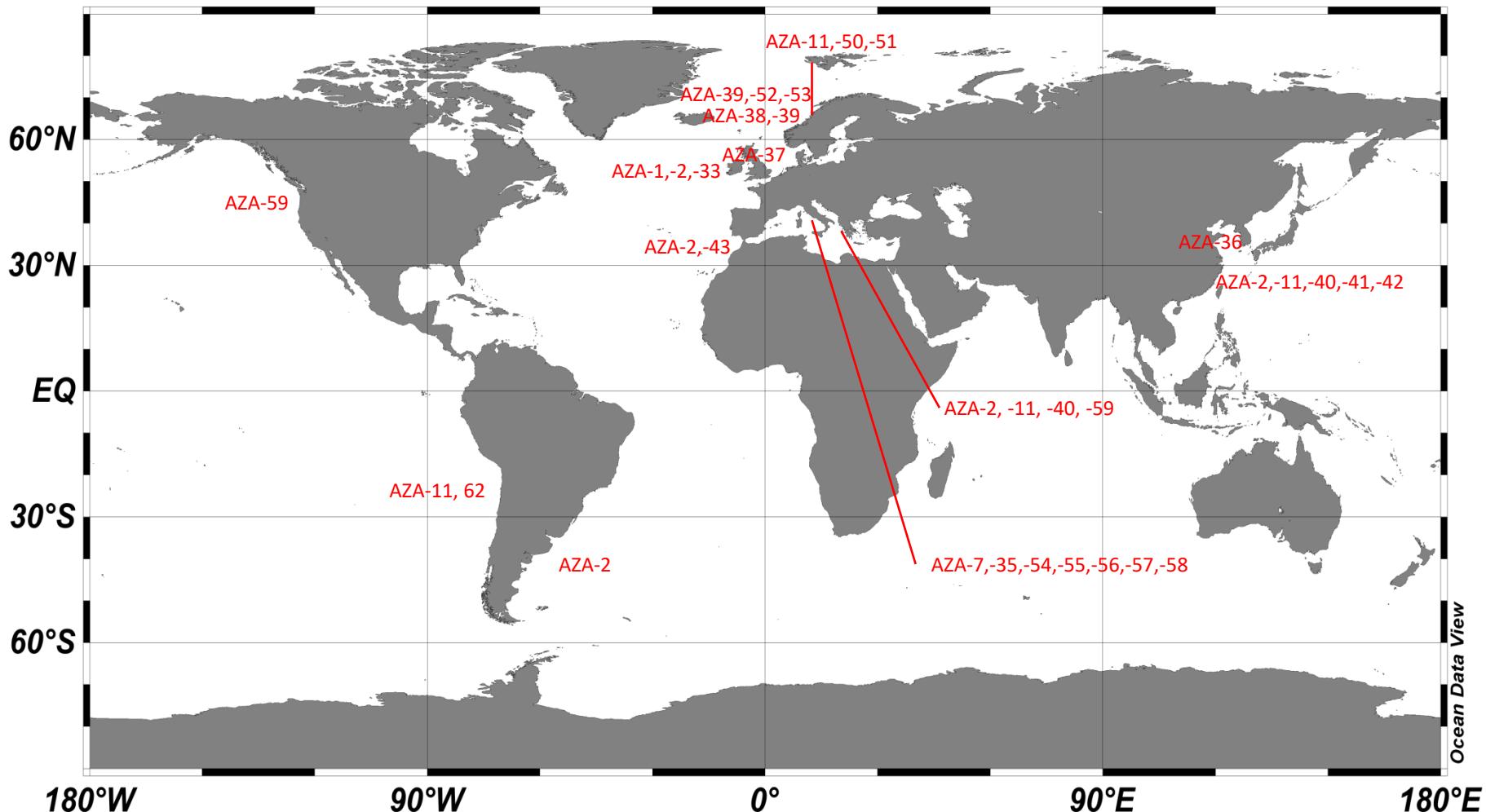
26 AZAs from  
planktonic origin

Currently  
62 published AZAs

And at least  
additional  
10 known AZAs

# Occurrence of Toxins and their Producers

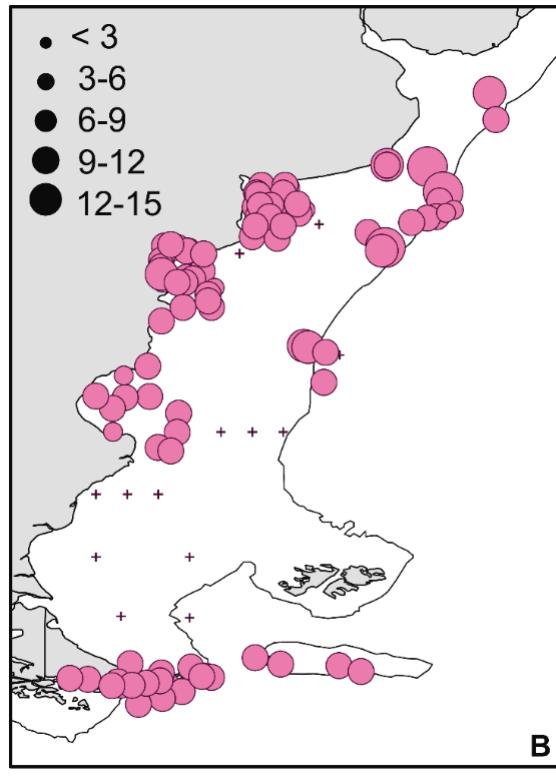
## Azaspiracids – Geographic distribution



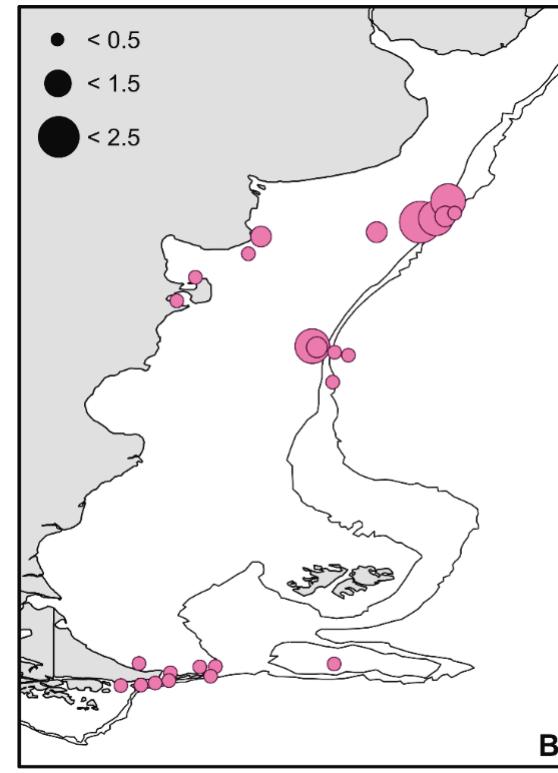
## Funded projects with AWI & IADO participation:

- |           |   |
|-----------|---|
| 2012-2014 | BMBF-Mincyt “Interactions between biogeochemistry and plankton composition of the Argentine Continental Shelf”  |
| 2013-2016 | EU: FP7-people-2012-IRSES, “IMCONet”  |
| 2015-2016 | DAAD-MINCYT “Detection of phycotoxins in Monte Hermoso, Argentina”  |
| 2015-2017 | BMBF-MINCYT “RETCEL”  |
| 2014      | FONCyT PICT “Dinámica planctónica y vectorización de ictiotoxinas hacia consumidores de niveles superiores en tramas tróficas pelágicas de la región de El Rincón - Golfo Norpatagónicos” |
| 2017-2024 | BMBF “DynAMo”   |
| 2020-2025 | EU: H2020-MSCA-RISE-2019 “CoastCarb”  |
| 2021-2023 | DAAD-CONICET, “PhytoxNorpat”  |

# Occurrence of Toxins and their Producers



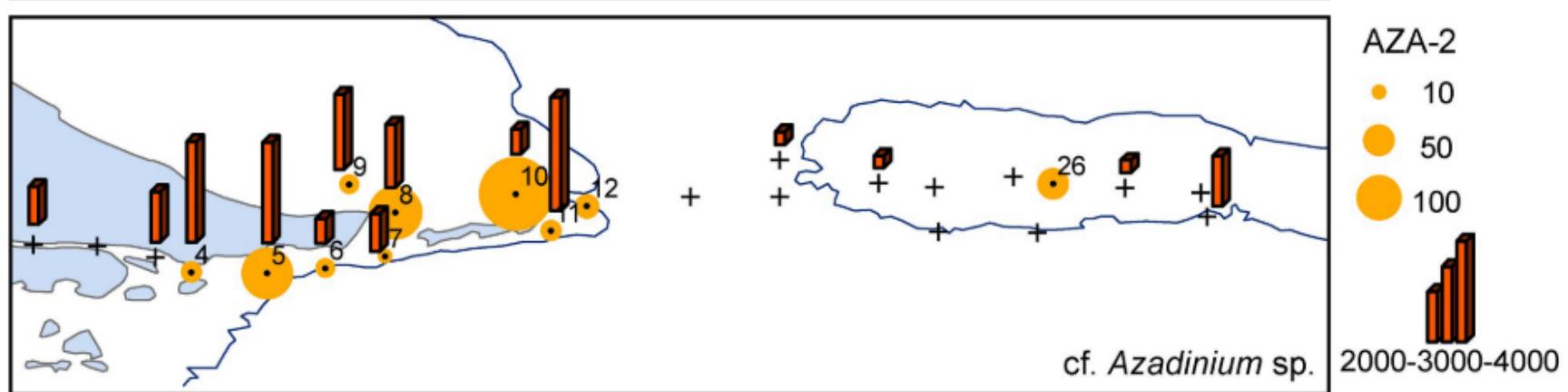
● *Amphidomataceae*



● AZAs

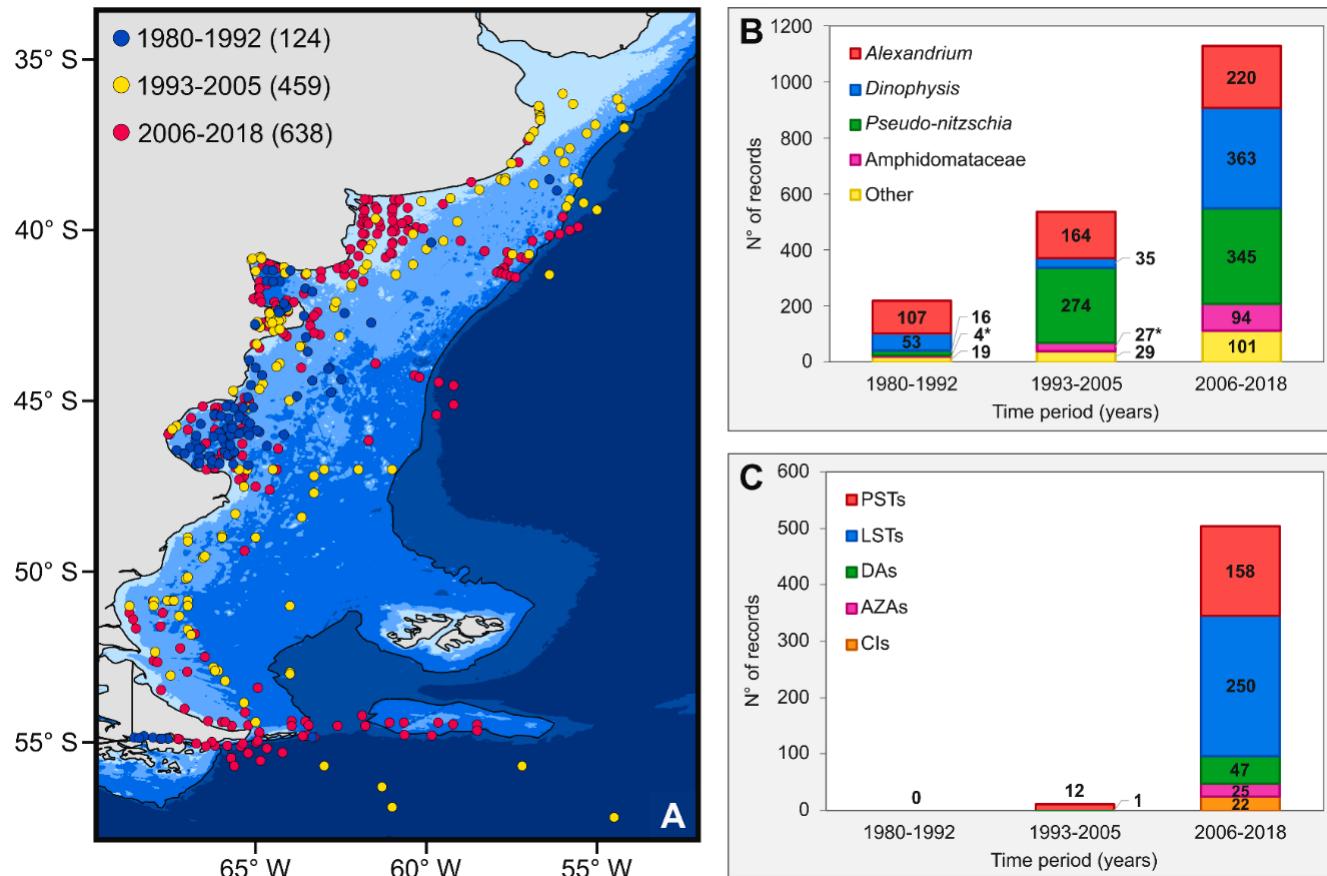
Ramírez, F.J. et al. 2022 Harmful Algae 118: 102317.

# Occurrence of Toxins and their Producers



Guinder, V.A., et al. (2020). PLOS ONE 15(5): e0233156.

# Occurrence of Toxins and their Producers



Ramírez, F.J. et al. 2022 Harmful Algae 118: 102317.

# Occurrence of Toxins and their Producers

- 1) Krock, B., et al. 2013, 9th International Conference on Molluscan Shellfish Safety, Sydney, Australia, FAO.
- 2) Akselman, R., et al. 2015, Harmful Algae 45(0): 40-52.
- 3) Fabro, E., et al. 2015, Harmful Algae 42(0): 25-33.
- 4) Gracia Villalobos, L., et al. 2015, Journal of Shellfish Research 34(3): 1141-1149.
- 5) Krock, B., et al. 2015, Journal of Marine Systems 148(0): 86-100.
- 6) Fabro, E., et al. 2016 Harmful Algae 59: 31-41.
- 7) Giannuzzi, L., et al. 2016, Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology 189: 22-30.
- 8) Hernando, M.P., et al. 2016, Biocell 40(1): 23-25.
- 9) Tillmann, U., et al. 2016, Harmful Algae 51: 40-55.
- 10) Almundoz, G.O., et al. 2017, Harmful Algae 63: 45-55.
- 11) Barrera, F., et al. 2017 Journal of Marine Systems 175: 36-45.
- 12) D'Agostino, V.C., et al. 2017, Harmful Algae 68: 248-257.
- 13) Fabro, E., et al. 2017, Journal of Phycology 53(6): 1206-1222.
- 14) Fabro, E., et al. 2018, Boletín de la Sociedad Argentina de Botánica 53(4): 551-566.
- 15) Guinder, V.A., et al. 2018, Frontiers in Marine Science 5(394).
- 16) Fabro, E., et al. 2018, Oceanography 31(4): 145-153.
- 17) Krock, B., et al. 2018, Oceanography 31(4): 132-144.
- 18) Tillmann, U., et al. 2018, European Journal of Phycology 53(1): 14-28.
- 19) D'Agostino, V.C., et al. 2019, Environmental Toxicology and Chemistry 38(10): 2209-2223.
- 20) Garzón-Cardona, J.E., et al. 2019, Journal of Marine Systems 195: 74-82.
- 21) Tillmann, U., et al. 2019, Harmful Algae 84: 244-260.
- 22) Fabro, E., et al. 2020, Marine and Freshwater Research 71(7): 832-843.
- 23) Guinder, V.A., et al. 2020, PLOS ONE 15(5): e0233156.
- 24) Hoffmeyer, M.S., et al. 2020, Journal of Marine Systems 212: 103448.
- 25) D'Agostino, V.C., et al. 2022, Oecologia 198(1): 21-34.
- 26) Ramírez, F.J., et al. 2022, Harmful Algae 118: 102317.



Any  
Questions?

Thanks for  
Your Attention!

